

REVIEW ARTICLE

In vivo Near Infrared Spectroscopy: a novel approach for simultaneously estimating molecules and hemodynamic parameters in the human and rat brain: a review

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Abstract

There have been great advances in optical brain imaging over the last 50 years and the technique has grown into a richly diverse field. *In vivo* recording and imaging using light provides extraordinary sensitivity to functional changes through intrinsic contrast, blood, and can even exploit the growing availability of exogenous optical contrast agents. Light can be used to analyze microscopic structures and function *in vivo* in the exposed animal brain, while also allowing noninvasive imaging of hemodynamics and metabolism in a clinical setting. This review is an overview of approaches that have been applied *in vivo* optical brain recording, in both animals and humans. The basic principles of each technique are described, emphasizing the techniques used in our laboratory.

Techniques include imaging of exposed cortex, *in vivo* functional spectroscopy of the living brain using optic fibers, and the broad range of noninvasive topography and tomography approaches to near-infrared imaging of the human brain. The basic principles of each technique are described, followed by examples of current applications to cutting-edge neuroscience research. In summary, it is shown that optical brain recording continues to grow and evolve, embracing new technologies and advancing to address ever more complex and important neuroscientific questions.

KEY WORDS: Spectroscopy, optical imaging; two-photon microscopy; near-infrared spectroscopy; diffuse optical tomography; neuroimaging; neurovascular coupling

1. Introduction

The interaction of light in tissue to recognize disease has been widely researched since the mid-19th century when Joseph von Fraunhofer developed diffraction grating. A large number of scientists have brought optical spectroscopy forward and enabled it to become a precise and quantitative scientific technology.

In 1963, when Franz F. Jöbsis published a new optical method in an original article(1), near-infrared spectroscopy (NIRS) was seen as the technique which could deliver a solution to a clinical need. In 1977, this author demonstrated the possibility of detecting changes in adult cortical oxygenation during hyperventilation(2). NIRS has become an established research and clinical tool for measuring changes in cerebral oxygenation, in particular, changes in oxygenated (HbO₂) and deoxygenated (HbR) hemoglobin concentration.

The technology has gained interest in the medical field in numerous biomedical applications for its advantages over existing conventional techniques. Optical spectroscopy at infrared and visible wavelengths avoids the use of ionizing radiation, is non-destructive, utilizes relatively inexpensive equipment, and can be performed near real-time without pharmaceutical means to enhance contrast, i.e., contrast agents.

Different optical recording techniques, both in visible and near infrared, have been used in animal experimentation. One of the most widely used techniques has been the exposed-cortex imaging; its use in animal studies has been widespread. Although it has been widely used by many groups, optical imaging in experimental animals has been only one step towards the study of imaging in clinical diagnosis and an excellent tool to learn much more about the basic mechanisms of brain function both in

physiology and in pathology. These results can be useful to help the development of new drugs and treatments. These studies can also contribute to the interpretation and better comprehension of results from other imaging modalities such as functional magnetic resonance imaging (fMRI) or positron emission tomography (PET). Some of these applications of animal imaging have included studies of Alzheimer's disease(3), stroke(4), epilepsy(5) and published by our group, the mechanisms of neurovascular coupling(6).

The obvious advantage of optical imaging over other modalities is its reduced cost and infrastructure requirements (such as shielded rooms, synchrotrons etc...).

This review describes a selection of optical approaches to detect functional brain activity. The basic principles of each technique are described, highlighting the techniques used in our laboratory, 1) Invasive optical brain techniques, including: a) optical techniques for exposed cortex imaging, b) recording functional activity using optic fibers, 2) noninvasive clinical optical imaging of the living brain.

2. Invasive optical brain techniques.

2.1. Optical techniques for exposed-cortex imaging.

The exposed-cortex imaging in animals rather than humans provides significantly more flexibility, since preparations can be much better controlled and all types of experiments can be systematically compared. Extrinsic dyes and cross-validation techniques such as voltammetry, amperometry or electrophysiology can even be used simultaneously(6).

This technique therefore also offers significant technical advantages for small animals, allowing higher resolution imaging and improved sensitivity. The cortex can be

surgically exposed to obtain high resolution imaging, allowing direct optical imaging of the brain's surface with only minimal disturbance to brain activity. Exposed cortex is highly accessible, and most commonly performed in experimental animals, although it has also been achieved on the intra operator human brain(7)(8).

In the neuroscience literature, the exposed-cortex is the simplest optical imaging techniques. The most useful are, HbO₂ and HbR dynamics(6); imaging extrinsic voltage sensitive dyes(9); speckle-flow imaging(4), capable of imaging the blood flow dynamics in the superficial cortex.

Exposed-cortex imaging has been applied to an extensive range of research areas. These can be summarized as: 1. functional imaging to improve understanding of the basic mechanisms of the hemodynamic and neuronal response to stimulus, 2. functional imaging to investigate the sensory and cognitive processing functions of the brain, and 3. a study of the effects of diseases and treatments on normal brain behavior.

2.1.1. Technical procedure.

Surgical preparation for exposed-cortex recordings, the experimental subject, commonly a rat, mouse, cat or primate is anesthetized while the scalp is retracted and the skull is carefully removed from over the brain area of interest. In some cases, the skull may be carefully thinned to obtain a good vision of an area of the brain's surface, avoiding contact with the brain cortex. Many imaginative tricks have been developed to ensure that correct brain function is not affected. Most often, these techniques are performed with anesthetized animals. However, in many other cases a cranial window can be implanted in the animal to perform chronic optical studies.

While the techniques described above make it possible to obtain information from the cortex (2D-imaging), there is the possibility of recording images with tomographic information (3D) using Optical Coherence Tomography (OCT)(10) of functional brain activation, Laminar Optical Tomography(LOT)(11), Fluorescence Lifetime Imaging and Microscopy (FLIM)(12) and Two-Photon Tomography(13)(14).

Brain *in vivo* imaging for research using Two-Photon Tomography has also found applications in areas of functional mechanisms, functional processing and pathology/treatment research(13)(14). These applications exploit a variety of methods to introduce fluorescent contrast into the brain, including intravenous injection of dextran-conjugated dyes to show blood vessels, topical application, or pressure injection of dyes into the cortex, transgenic mutation of cells to express fluorescent proteins and systemic delivery of dyes.

2.2. Recording functional activity using optic fibers.

We will now review fiber optic probe scattering spectroscopy of turbid tissues using visible and infrared light. A spectroscopic system incorporates a light source, an optical analyzer with a detector, and a light transport conduit, which, in many cases, is made of optical fibers, figure 1. The excitation or illumination light source is usually a laser or a white light source, such as a xenon or incandescent lamp. The coupling optics adapts the f-number of the light source to the numerical aperture of the fiber and guarantees optimal irradiance into the fiber.

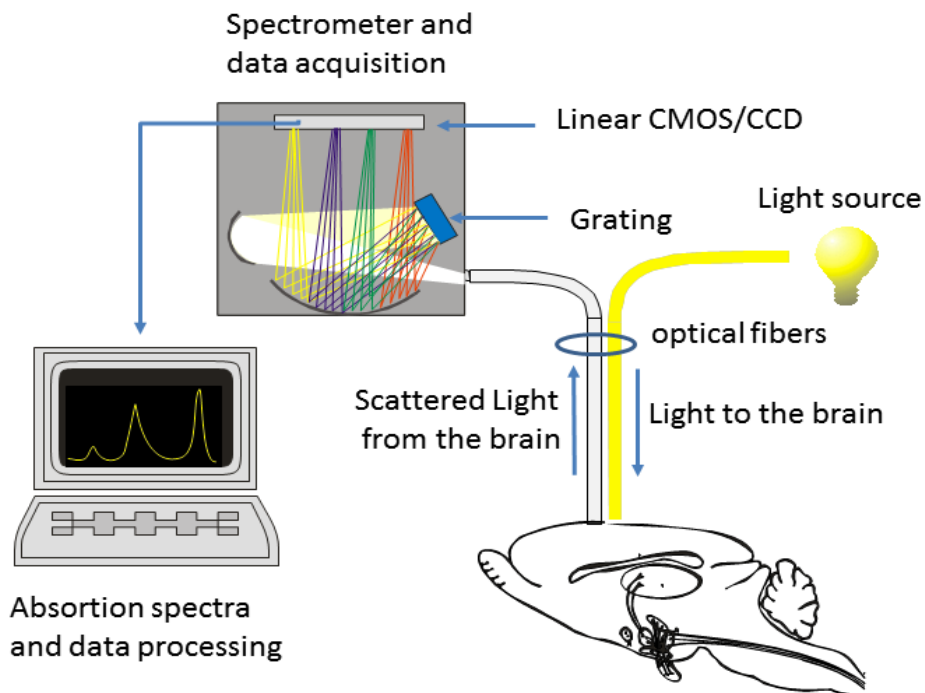


Figure 1. Fiber optic-based spectroscopy system, with separate illumination (excitation source, LED or incandescent light bulb). Optical elements couple the excitation light into the flexible probe, a probe collects the emitted light, coupling optics adapt the numerical aperture of the probe to the miniature spectrometer and an optical detector (CCD, or CMOS linear element) is read out and digitized.

Single fiber solutions are used and well-aligned coupling optics to achieve the smallest probe diameters. Single-fiber solutions are the most commonly used because of the small diameter 50-100 μ m and the fact that single fiber-based probes require a minimal amount of components for the probe and can be used to create the smallest illumination spots as well as

having excellent light collection efficiency. The simplest way to setup is Y-shaped assemblies with two fibers of the same diameter side-by-side in the common end, which then diverges into two separate legs. The fibers in the assembly may be UV-VIS, VIS-NIR or one of each in a mixed bifurcated assembly(15). See figure 2 for more details.

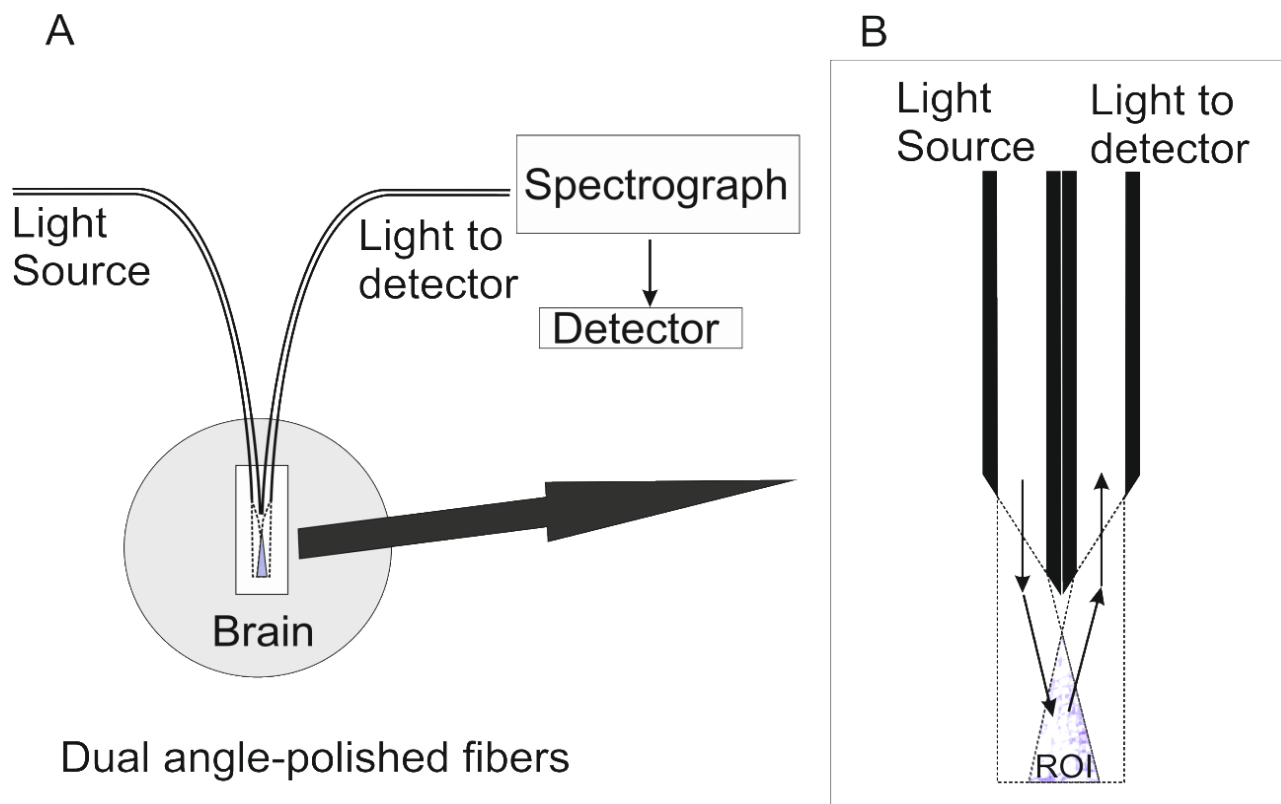


Figure 2. A. Y-shaped assembly with two fibers of the same diameter side-by-side in the common end implanted in brain, which then diverge into two separate legs. B. Zoom of distal tips of fiber optic in dual angle-polished fibers configuration. The arrangement of the fibers and their angle of polishing try to prevent damage to the cerebral parenchyma as much as possible.

Another of the additional advantages of simultaneously using optical fibers is the possibility of using other neurotechniques (voltammetry(6), amperometry(16), electrophysiology(17), microdialysis(18), etc.), that complement the obtained information. The optical registration techniques do not generate interferences or electronic noise that could alter the results.

As an example of what was said above, using microdialysis and fNIRS, we found that intracerebral infusions of amphetamine increase the extracellular concentration of glutamate, dopamine, aspartate, GABA, and taurine. This study(18) also shows that an alpha-noradrenergic receptor antagonist is able to attenuate the effects of amphetamine on the release of glutamate, dopamine,

GABA and taurine, which further suggests a vasoconstrictor effect of amphetamine as a result of which hypoxia could develop.

2.2.1. In vivo spectroscopy: for simultaneously estimating nitric oxide and hemodynamic parameters

Nitric oxide (NO) is a well-known signaling molecule involved in a wide range of biological processes. Under physiological conditions, NO reacts with HbO₂ to form methemoglobin (MetHb) at a very high rate. Microdialysis studies have used hemoglobin solutions as a trapping method to quantify NO *in vivo*. The methodology described here uses the microcapillary network (capillary bed) with endogenous

HbO₂ instead of a microdialysis probe with exogenous HbO₂ for monitoring MetHb as an indirect index of NO levels by *in vivo* spectroscopy using optical fibers.

This method has been validated using *in vivo* voltammetry and selective NO microelectrodes. We have used *in vivo* local infusion of NO into the tissue surrounding the probe (optodes) in both methods, NOS inhibitors to decrease the NO production and local infusion of NMDA agonists to increase NO production in the cerebral cortex. Thus, the association between *in vivo* voltammetry and *in vivo* spectroscopy as we have described for our group could be very advantageous, because by using both methodologies it is possible to measure the NO directly in the extracellular fluid (voltammetry) and its deactivation by its principal *in vivo* scavenger (spectroscopy). Moreover, the latter technique makes simultaneous measurements of hemodynamic parameters such as oxygenation rate and blood volume (cerebral blood flow) possible, see figure 3.

NO is extremely unstable *in vivo* and its half-life has been estimated as a few seconds(18). In accordance with its role as a paracrine mediator, NO can travel to reach target cells in neighboring areas of the NO-generating cell. During the paracrine migration, in particular at high concentrations, this reactive molecule can interact with molecular oxygen to form higher nitrogen oxides (e.g. NO₂ and N₂O₃), which can either react with other biomolecules such as thiols and amines or be hydrolyzed to nitrite (NO₂⁻) and nitrate (NO₃⁻)(16). However, the most important reactions are with ferrous hemoproteins and especially with hemoglobin (Hb)(19), (figure 3A), such as those which yield nitrosylhemoglobin or methemoglobin. Nitrosylhemoglobin formation has a very low rate(16), whereas the interaction with HbO₂ is characterized by a very high rate

even under saturating oxygen concentrations, and it has been estimated to be at least 26 times faster than the auto-oxidation of NO in aqueous solution. Thus, MetHb levels are proportional to the NO concentration and they can be used as an indirect index of NO (20)(21)(22), as we can observe when this technique is compared with another technique, see figure 3, C and D.

Many results indicate that this spectroscopy technique is able to record large increases in MetHb levels and to detect reductions of its basal levels(16)(17)(23)(24)(22). In addition, data show that similar changes and kinetics can be observed with both techniques. Thus, intravascular MetHb can be used as an indirect index of NO levels. It is proposed that *in vivo* spectroscopy may be a useful tool to gain insight into the roles of NO in hemodynamic parameters and in other physiological processes such as the regulation of the mitochondrial respiratory chain(18)(16).

Finally, this technique offers the possibility of monitoring the neuronal activity, bearing in mind that it is widely accepted that changes or alterations in regional cerebral blood flow and cerebral oxygen consumption rate can be used as an index of neuronal activity. Other groups have made interesting contributions using our method(23)(24).

3. Non-invasive optical imaging of the human brain

Functional near infrared spectroscopy (fNIRS) commonly used the topography approach in brain research covering different fields such as physiology (25), psychiatry (26), alterations in disease (27), and its application in the brain computer interface (BCI) (28), in neuroimaging studies has recently been developed. However, the method has some

disadvantages as the relative positions of the measurement channels to brain anatomy vary between subjects or fNIRS measurements in cortical areas are always

affected by the hemodynamic changes in the scalp layer affecting the interpretation of results in cortical activity(29)(30).

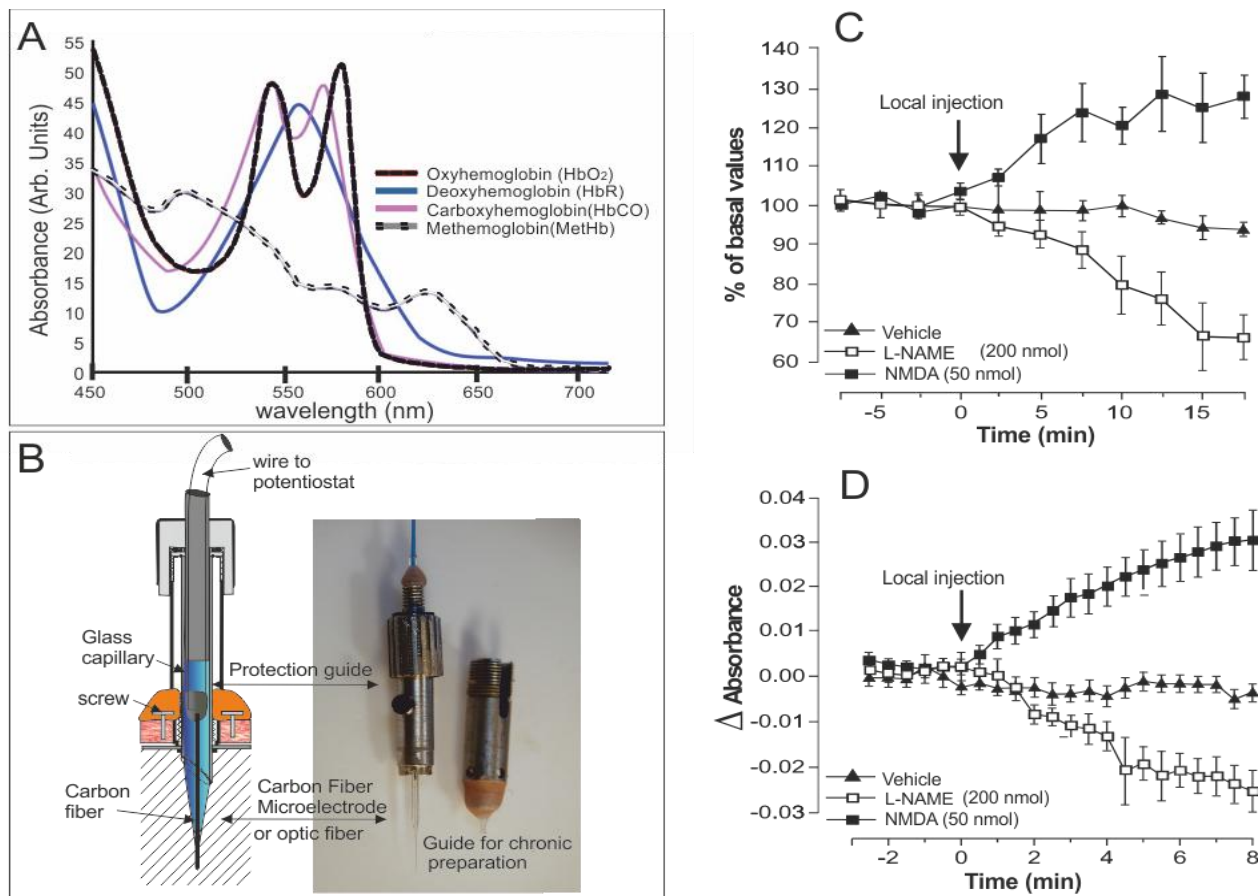


Figure 3. A, Absorbance in arbitrary units of oxyhemoglobin (HbO₂), deoxyhemoglobin (HbR), carboxyhemoglobin (HbCO) and methemoglobin (MethHb) as a function of wavelength. B, Mechanical device for selective microelectrode brain implantation for nitric oxide (NO) and fiber optics. Schematic drawing and photography. C and D, Kinetic effects of the manipulation of NO synthesis on MetHb band and free NO levels in rat cerebral cortex determined by *in vivo* spectroscopy (D) and *in vivo* voltammetry (C), respectively. Absorbance values at 635 nm (A) were averaged every 30 s (scan rate: 40 spectra min⁻¹). Voltammetric peaks of NO (oxidation peak at approximately 650 mV) were recorded every 2 min and percentages of modification from basal values were calculated. Data represent mean \pm SEM (n = 5-8 rats per group) and arrows indicate the infusion of drugs close to the microsensors (see publication 17 for more detail) As illustrated in C and D, NO and MetHb levels were decreased by the potent NO-synthase inhibitor L-NAME, white squares, but were not modified by vehicle solution (PBS), black triangles. In addition, NO and MetHb levels were markedly enhanced by the administration of NMDA, black squares. Time scales for spectroscopic and voltammetric studies differ due to the differences in scan rate between both techniques.

3.1. Advantages of optical techniques

Functional brain imaging has provided substantial information about how dynamic neural processes are distributed in space and time. A large number of brain studies based on task or resting state imaging studies of neural networks through healthy subjects have been reported(31)(32)(33). Some imaging modalities to study brain functioning use fMRI which require costly infrastructure, while optical imaging instruments are less expensive. Moreover, technique limitations in fMRI devices such as a fixed scanner, contraindication with metal implants, scanner noise and stress associated with fear are avoided or reduced in optical imaging devices.

fMRI measures changes in the *blood oxygen level dependence* (BOLD) signal associated with hemodynamic changes after the neural activity to visualize functional changes in the brain. Although the BOLD signal has been associated to a decrease of HbR(34), an increase of BOLD signal could be associated to an increase of HbO₂(35) or a combination of both. Initially, the HbR decreases and then the HbO₂ increases due to the vasodilatation that washes the local HbR. The above controversy disappears with the use of optical imaging techniques, which measure each hemoglobin state separately (HbR & HbO₂), using at least two wavelengths to measure each hemoglobin state. Moreover, the optical imaging techniques provide more comprehensive information of hemodynamic and metabolism than the BOLD signal, due to the complicated connection of the BOLD signal to the neurovascular coupling.

Optical imaging techniques can measure changes in HbO₂, HbR and HbT at a much higher sampling rate than fMRI, and this could be a fundamental tool for the study of the neurovascular coupling in humans,

especially when the neurovascular coupling is either unknown or altered. The principal advantage of fMRI measurements is that they can cover the whole brain, while optical measurements only reach the cerebral cortex because its penetration depth is around of 3-4 centimeters could anatomically reach the gyral level(36).

Finally, unlike other imaging modalities such as the PET(37) or x-ray computed tomography(38), functional brain measurements using optical imaging do not need a contrast agent, whose doses are limited in infants and could induce anaphylactic reactions in certain populations.

All these aforementioned circumstances have potentiated the use and developments of optical imaging techniques in recent years for research, diagnosis and prognostic studies.

3.2. Instrumentation

A wide variety of NIRS instruments has been created for different types of measurements, with the most common being the following: continuous wave (CW), time domain (TD) and frequency domain (FD).

- In CW measurements, the light is emitted at a constant intensity by sources into the tissue, and the same device detects the transmitted light intensities. CW uses frequency-encoded intensities to acquire data and can simultaneously measure wavelength(39) or light sources(40).
- TD uses ultrashort laser pulses to irradiate the tissue, and the light intensity detected is recorded over time to show a temporal point spread function (TPSF) with a resolution of picoseconds (41)(42).

- In FD measurements, the light source is modulated at radio frequencies (100-1000 MHz)(43), and measures the phase delay of the light detected from the tissue(44). The parameters of FD measurements are phase shift, the intensity of light (DC component) and the amplitude of the intensity oscillations (AC component) at given wavelengths and for different distances between the light source and detector. The FD instrumentation is more complex and expensive than CW systems, thus a combination of CW measurements and small frequency-domain measurements have been proposed to provide good spatial resolution and quantitative accuracy(45).

3.3. Tomography approach

The most significant improvements in optical imaging came when image reconstruction techniques were proposed in the 1990s using diffusive photons(46)(47). Diffuse optical tomography (DOT) is an fNIRS approach that transforms the detected light from different measuring distances on the surface of the head into depth information providing three-dimensional images of cerebral activations(48). DOT uses the multi-distance approach with the purpose of increasing spatial resolution and positional accuracy of optical brain imaging(49). Unlike the topography approach which directly maps the changes in optical properties from the midway between a source and detector into a 2-D image(50).

3.3.1. Image reconstruction

The **forward model** is used by DOT to model light migration processes to create functional images. The forward model relates the activity inside the head tissue

with the measured light intensity changes, using the radiative transfer equation (RTE) or diffusion approximation (DA). The mathematical forward model must be implemented in computational models. There are three types of computational modeling approach, which are the following: analytical modeling, stochastic modeling and deterministic modeling.

- Analytical modeling uses Green's function for the solution of partial differential equations such as the RTE or DA in a homogeneous semi-infinite medium(51) and simple geometries. It has been used to validate stochastic and deterministic models.
- Stochastic modeling whose distribution of optical properties is calculated by Monte Carlo simulations which model the light propagation inside a 3-D realistic head obtained from MRI scans, where heterogeneous structures are incorporated to the simulation(52). This method allows heterogeneity and a flexible shape of the medium.
- Deterministic modeling is based on the finite element method to solve the DA. FEM is capable of dealing with heterogeneity in arbitrary geometries. This is the most commonly used system in diffuse optical imaging.

The relationship between the measurement of light intensity and optical property is non-linear. However, the relationship is assumed to be linear for DOT images, and is known as the **inverse problem**. Two approaches are used to solve the inverse problem: linearization and nonlinear iterative approaches.

The linearization approach does not correctly predict changes in the optical properties, showing as results qualitative

image reconstructed of measured changes in the brain(53). This approach is good enough for neuroimaging studies, but not for clinical studies because, for it, it's necessary to have quantitative images of hemoglobin states, whereas the nonlinear iterative approach applies an iterative optimization method to minimize the differences between the calculated and measured data of the distribution of optical properties. In addition, a Jacobian matrix or sensitivity is found as a product of both approaches, which relates the number of measurements on the surface and changes in the optical properties, and must be computed. Methods have been proposed such as the perturbation method(54) or the gradient-based method(55) to compute the Jacobian matrix

Finally, the inverse problem is ill conditioned suggesting that the reconstructed images are sensitive to noise during the measurements. Some research groups have attempted to solve this using regularization methods(56)(57) such as the use of decomposition singular values(58).

In addition, anatomical information of a subject can be a problem during the image reconstruction. The DOT technique cannot provide anatomical information making it difficult to solve the forward model, unlike fMRI or x-ray CT which provide anatomical information. In neuroimaging studies using DOT technology, the optical model is constrained to the tissue geometry through segmented MRI scans. In order to solve this problem, some research groups have proposed other methods to perform DOT studies without MRI scans such as MNI-guided DOT(59) or DOT images reconstructed on a generic head model(60).

The same problem occurs when NIRS technology is applied on other tissues such as prostate or breast. X-ray CT(61) or

ultrasound(62) or MRI scans can be used to improve the image reconstruction.

3.4. Applications in functional brain imaging

Optical measurements can play a role in determining underlying brain physiology, especially when investigating the relationship between neural activity and hemodynamic changes known as neurovascular coupling from animals(63) to humans(64). fNIRS has been used in a wide variety of applications in neuroimaging studies to measure functional changes associated to a stimuli or paradigm. Some examples are listed below:

- Cognitive stimuli based on go/no-go paradigms(65). Different letters are presented on a screen for a few seconds followed by an inter-stimulus period of 1-2 seconds. On the one hand, the participants are instructed to press a button with their right index finger each time a letter appears on the screen (during the go). On the other hand, the participants are instructed to push the button for all letters except X (during the no-go). Although, the go/no-go paradigm is one of the most used in cognitive studies, some authors, as is the case of our laboratory, have used other cognitive tasks such as mental arithmetic tasks(66), see figure 4.
- Somatosensory and motor stimuli based on finger tapping(67), tactile stimulation(68) or finger flexion/extension(69) tasks are some of the most used examples in neuroimaging studies. Due to the fact that cerebral activation amplitudes are higher than the amplitudes given by other paradigms e.g. cognitive, the cerebral activations are reproducible and spatial localizations of the motor

activity are known, the motor paradigms are especially used in the new method applications such as image reconstruction algorithms(56),

filtering procedures(70) or corroboration of simulated models(57).

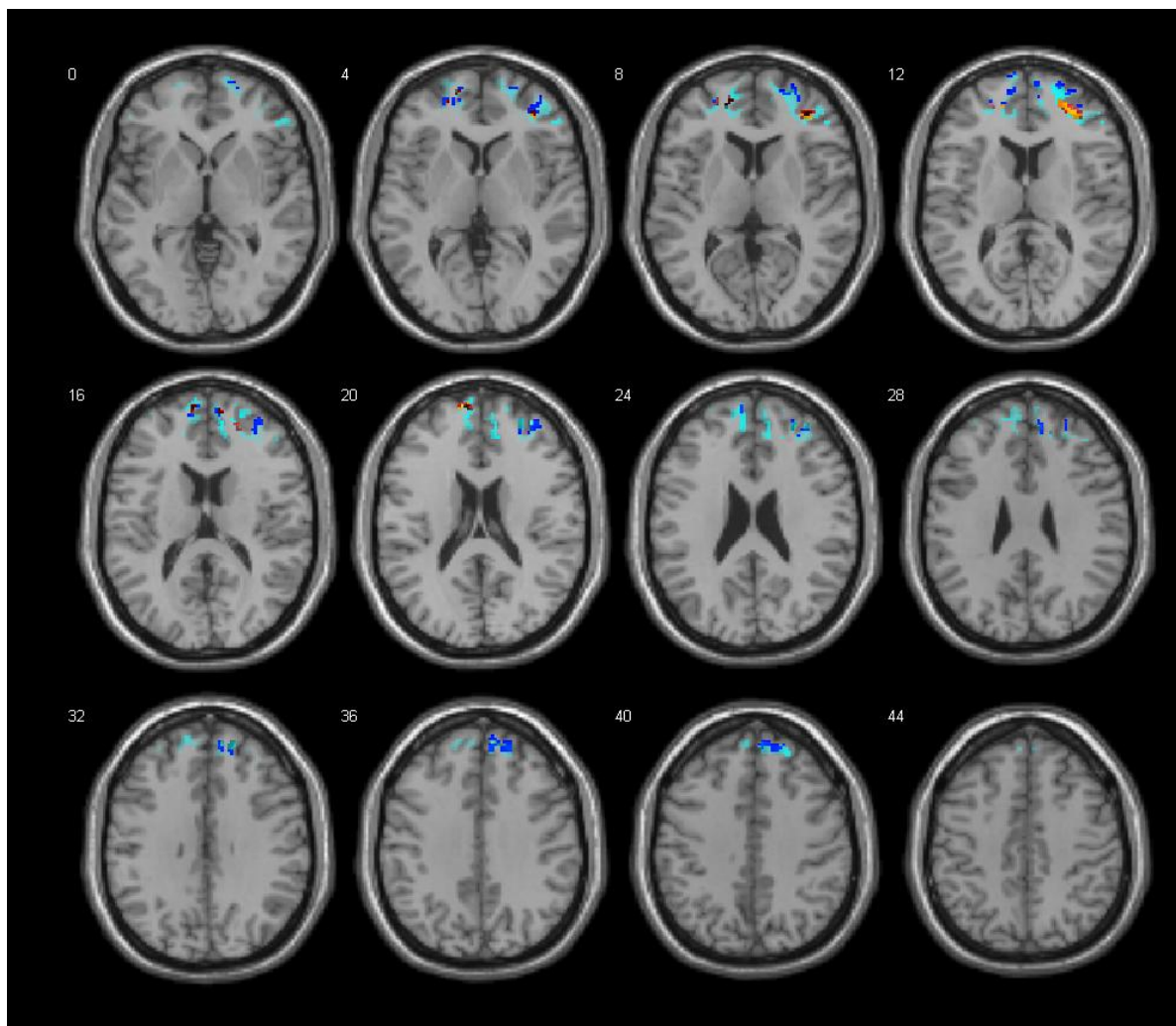


Figure 4. Spatial conjunction of HbO (yellow), HbR (red), HbT (blue) and BOLD (cyan) signals during a mental arithmetic task based on easy count<difficult count, performance by a subject in 6 sessions on subsequent days. Easy count refers to counting backwards from a 3-digit number for 1s. e.g. “136 for 1s”. Difficult count refers to counting backwards from a 3-digit number for 7s, e.g. “136 for 7s”, for 30 seconds of the task period. Each condition was repeated twelve times in both DOT and fMRI devices with a random order of the instructions. All resulting t-images of contrast selected were fitted a normalized anatomical space. Threshold p-value < 0.05, FDR corrected.

- Visual stimuli based on random dot stereo pairs where the stimuli are presented as a pair of images, one to each eye, then when viewed

binocularly a strongly fused perception of depth is produced(71). A visual paradigm is used because binocular vision allows the fusion of

each image presented from our retinas using the difference between them to estimate relative depths. A wide variety of studies to measure the relationship of HbO₂/HbR with perception have been performance(72).

- Resting-state based on the study of the functional architecture of the brain(73). Monitoring HbO₂ and HbR using fNIRS during a rest state of the participants can exhibit patterns of functional connectivity. Given that fNIRS uses a sampling rate higher than classical fMRI, optical measurements allow the study of low frequency components (aim of resting-state studies), thereby avoiding the mix with high frequency components(74).

3.4.1. Applications in infants and neonates

Portable and noninvasive measurements are characteristic of NIRS devices, which allow blood flow and oxygenation monitoring in infants and neonates especially in brain injuries. It is essential to control hemodynamic changes during the development of neurological disorders in these patients. Various studies have reported the suitability of fNIRS to measure changes in saturation, blood volume and relative cerebral metabolic rate of oxygen(75), even in hemorrhage in a premature baby(76).

3.4.2. Clinic applications

NIRS has been used in populations for which other imaging modalities are impractical, such as the elderly and infants, because fNIRS devices are flexible, minimally invasive and can be portable. Despite its limited depth penetration and difficulty to apply on darker skins or hairs,

fNIRS devices can still be used for neurologic and psychiatric disorders studies such as the following:

Neurologic disorders such as Parkinson's(77), Alzheimer's(78), epilepsy(79), ischemia(80) or aging(81), have been evaluated using fNIRS devices. Moreover, psychiatric disorders such as schizophrenia(82) or anxiety disorders(83), have also been monitored by fNIRS devices.

In spite of the wide use of fNIRS devices for brain imaging, an essential application is, without doubt, for breast cancer imaging, one the most common cancers in women. Currently, the most usual screening is x-ray mammography combined with physical examination. Tumors are normally associated with an increase in vascularization. In these cases, NIRS can play an important role because it can measure blood volume and oxygenation to determine the presence of a tumor. A variety of studies have shown the capability and feasibility of fNIRS to identify the increased vascularization associated with a tumor(84)(85), despite the fact that the poor spatial resolution is still insufficient for a diagnosis.

3.4.3. Other applications

fNIRS not only offers potential applications in diagnosis and evaluation of diseases, but can also be used to monitor the saturation of oxygen in the blood during the neurorehabilitation in stroke patients, allowing the evaluation of the recovery degree. Some research reports the use of fNIRS devices to monitor functional changes during neurorehabilitation processes in cognitive disabilities(86), motor disabilities(87) or aphasia(88). A new application of fNIRS is being developed based on the application of transcranial magnetic stimulation (TMS)

whose electrical changes produced inside the brain lead to hemodynamic changes which can be monitored by fNIRS. Electroencephalography (EEG) has been used to monitoring the cerebral changes generated during the TMS (89). The EEG allows the measurement of changes in the bioelectric activity while the TMS is applied, with a good temporal resolution but does not provide hemodynamic information. Some authors have stimulated previous to or between intervals of radio frequency pulses inside fMRI(90) to measure cerebral functional changes generated by TMS. It is not possible to monitor functional changes during the stimulation period because of the electromagnetic incompatibility with fMRI.

In these cases, NIRS offers the possibility of functional change monitoring while the TMS is applied without interferences that can affect the results, an example of this, recorded in our laboratory can be seen in figure 5.

Although, there are some discrepancies about the hemodynamic changes measured by fNIRS during the TMS application, which depending on the cerebral area stimulated, TMS coil angulation, intensity, and frequency of the stimulation, the results are variable. In spite of these discrepancies, simultaneous fNIRS and TMS are potential tools for the study of the physiology that underlies the stimulation, neurovascular coupling and as clinic tools.

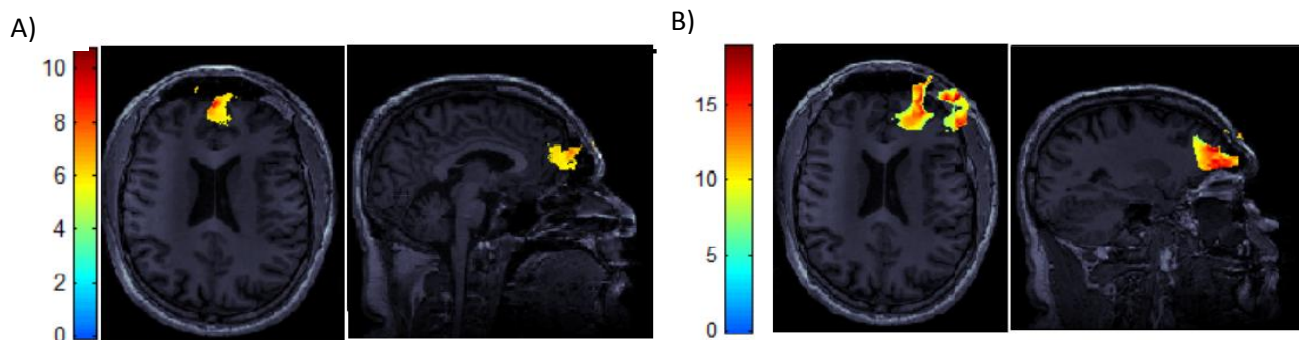


Figure 5. T-maps of brain activation during rTMS at high-frequency (>10Hz) vs resting period, simultaneously measured by DOT on A) the middle of the prefrontal cortex and B) the right lateral prefrontal cortex. All results were mapped onto the subject's anatomical scans. Threshold $p < 0.001$; $p < 0.05$ corrected FDR, at the voxel level for HbO signals. Color bars show HbO changes during rTMS.

In summary, for clinical applications, noninvasive optical imaging can provide complimentary information to other modalities such as fMRI and provide a low-cost alternative in some cases. This is in addition to serving populations often unable to receive MRI or PET scans such as young infants or the critically ill. Clinical optical brain imaging is generally noninvasive and uses NIR light to obtain improved penetration through the scalp, skull, and brain. To conclude, optical imaging's key

advantage is the ability to measure a range of functional contrasts, it can readily be exploited in functional brain imaging via a wide range of approaches from animal studies of the intricate cellular mechanisms of normal and diseased brain to *in vivo* noninvasive clinical brain imaging.

In addition, optical recordings or brain imaging is finding widespread applications as a research tool for both clinical and animal studies of basic brain function and

disease. At present, so little is known about the way that the normal brain functions, in part due to the difficulties of measuring such a complex organ without disturbing or damaging the brain's *in vivo* functioning. Optical imaging allows the living brain to be closely observed, as well as investigation into many functional interactions and changes over many length scales.

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5. References

1. Jöbsis FF. Spectrophotometric Studies on Intact Muscle: II. Recovery from contractile activity. *J Gen Physiol* [Internet]. 1963 May;46(5):929–69. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2195306/>
2. Jobsis FF. Noninvasive, infrared monitoring of cerebral and myocardial oxygen sufficiency and circulatory parameters. *Science* (80-). 1977;198(4323):1264–7.
3. Spires TL, Meyer-Luehmann M, Stern EA, McLean PJ, Skoch J, Nguyen PT, et al. Dendritic spine abnormalities in amyloid precursor protein transgenic mice demonstrated by gene transfer and intravital multiphoton microscopy. *J Neurosci*. 2005;25(31):7278–87.
4. Ayata C, Dunn AK, Gursoy-Özdemir Y, Huang Z, Boas DA, Moskowitz MA. Laser speckle flowmetry for the study of cerebrovascular physiology in normal and ischemic mouse cortex. *J Cereb Blood Flow Metab*. 2004;24(7):744–55.
5. Bahar S, Suh M, Zhao M, Schwartz TH. Intrinsic optical signal imaging of neocortical seizures: the “epileptic dip.” *Neuroreport*. 2006;17(5):499–503.
6. Roche R, Salazar P, Martín M, Marcano F, González-Mora JL. Simultaneous measurements of glucose, oxyhemoglobin and deoxyhemoglobin in exposed rat cortex. *J Neurosci Methods*. 2011;202(2):192–8.
7. Pouratian N, Cannestra AF, Martin NA, Toga AW. Intraoperative optical intrinsic signal imaging: a clinical tool for functional brain mapping. *Neurosurg Focus*. 2002;13(4):1–9.
8. Cannestra AF, Pouratian N, Bookheimer SY, Martin NA, Becker DP, Toga AW. Temporal spatial differences observed by functional MRI and human intraoperative optical imaging. *Cereb Cortex*. 2001;11(8):773–82.
9. Kleinfeld D, Delaney KR. Distributed representation of vibrissa movement in the upper layers of somatosensory cortex revealed with voltage-sensitive dyes. *J Comp Neurol*. 1996;375(1):89–108.
10. Maheswari RU, Takaoka H, Kadono H, Homma R, Tanifuji M. Novel functional imaging technique from brain surface with optical coherence tomography enabling visualization of depth resolved functional structure in vivo. *J Neurosci Methods*. 2003;124(1):83–92.
11. Hillman EMC, Boas DA, Dale AM, Dunn AK. Laminar optical

- tomography: demonstration of millimeter-scale depth-resolved imaging in turbid media. *Opt Lett*. 2004;29(14):1650–2.
12. Bacsikai BJ Hickey GA, Allen R, Hyman BT SJ. Fluorescence resonance energy transfer determinations using multiphoton fluorescence lifetime imaging microscopy to characterize amyloid-beta plaques. *J Biomed Opt*. 2003;8:368–75.
 13. Ohki K, Chung S, Ch'ng YH, Kara P, Reid RC. Functional imaging with cellular resolution reveals precise micro-architecture in visual cortex. *Nature*. 2005;433(7026):597–603.
 14. Chaigneau E, Oheim M, Audinat E, Charpak S. Two-photon imaging of capillary blood flow in olfactory bulb glomeruli. *Proc Natl Acad Sci*. 2003;100(22):13081–6.
 15. Hernández SE, Rodríguez VD, Pérez J, Martín FA, Castellano MA, Gonzalez-Mora JL. Diffuse reflectance spectroscopy characterization of hemoglobin and intralipid solutions: in vitro measurements with continuous variation of absorption and scattering. *J Biomed Opt*. 2009;14(3):34026.
 16. González-Mora JL, Martín FA, Rojas-Díaz D, Hernández S, Ramos-Pérez I, Rodríguez VD, et al. In vivo spectroscopy: a novel approach for simultaneously estimating nitric oxide and hemodynamic parameters in the rat brain. *J Neurosci Methods*. 2002;119(2):151–61.
 17. Martín FA, Rojas-Díaz D, Morales CA, Camacho J, González-Mora JL, Castellano MA. Simultaneous In Vivo Measurements of Methemoglobin and Other Endogenous Chromophores by Visible Spectroscopy.
 18. Del Arco A, González-Mora JL, Armas VR, Mora F. Amphetamine increases the extracellular concentration of glutamate in striatum of the awake rat: involvement of high affinity transporter mechanisms. *Neuropharmacology*. 1999;38(7):943–54.
 19. Kharitonov VG Sharma VS BJ. Interactions of nitric oxide with heme proteins using UV/VIS spectroscopy. In: *Methods in nitric oxide research*. 1996. p. 39–45.
 20. Felipe A. Martín Simeona J. Alonso, Eduardo Navarro, Miguel A JLG-M. Fiber optic spectroscopy to study neurovascular coupling in small areas of the rat brain. In: *Progress in Optical Fibers Research*. 2007. p. 369–89.
 21. Feelisch M Werringloer J KD. The oxyhemoglobin assay. In: *Methods in nitric oxide research*. 1996. p. 455–78.
 22. Martín FA, Rojas-Díaz D, Luis-García ML, González-Mora JL, Castellano MA. Simultaneous monitoring of nitric oxide, oxyhemoglobin and deoxyhemoglobin from small areas of the rat brain by in vivo visible spectroscopy and a least-square approach. *J Neurosci Methods*. 2004;140(1):75–80.
 23. Espinosa N, Cudeiro J, Mariño J. Spectroscopic measurement of cortical nitric oxide release induced by ascending activation. *Neuroscience*. 2015;285:303–11.
 24. de Labra C. Different sources of nitric oxide mediate neurovascular coupling in the lateral geniculate nucleus of the cat. *Front Syst Neurosci* [Internet]. 2009;3(September):2–3. Available from: <http://journal.frontiersin.org/article/10>.

- 3389/neuro.06.009.2009/abstract
25. Hyde DC, Boas DA, Blair C, Carey S. Near-infrared spectroscopy shows right parietal specialization for number in pre-verbal infants. *Neuroimage* [Internet]. 2010;53(2):647–52. Available from: <http://www.sciencedirect.com/science/article/pii/S1053811910008748>
 26. Okada F, Tokumitsu Y, Hoshi Y, Tamura M. Impaired interhemispheric integration in brain oxygenation and hemodynamics in schizophrenia. *Eur Arch Psychiatry Clin Neurosci* [Internet]. 1994;244(1):17–25. Available from: <http://dx.doi.org/10.1007/BF02279807>
 27. Ferrari M, Quaresima V. A brief review on the history of human functional near-infrared spectroscopy (fNIRS) development and fields of application. *Neuroimage*. 2012;63(2):921–35.
 28. Afergan DA. *Implicit Brain-Computer Interfaces for Adaptive Systems: Improving Performance through Physiological Sensing*. Tufts University; 2015.
 29. Kirilina E, Jelzow A, Heine A, Niessing M, Wabnitz H, Brühl R, et al. The physiological origin of task-evoked systemic artefacts in functional near infrared spectroscopy. *Neuroimage*. 2012;61(1):70–81.
 30. Saager R, Berger A. Measurement of layer-like hemodynamic trends in scalp and cortex: implications for physiological baseline suppression in functional near-infrared spectroscopy. *J Biomed Opt*. 2008;13(3):10.
 31. Turkeltaub PE, Coslett HB. Localization of sublexical speech perception components. *Brain Lang*. 2010;114(1):1–15.
 32. Yang J, Andric M, Mathew MM. The neural basis of hand gesture comprehension: a meta-analysis of functional magnetic resonance imaging studies. *Neurosci Biobehav Rev*. 2015;57:88–104.
 33. Samara Z, Evers EAT, Goulas A, Uylings HBM, Rajkowska G, Ramaekers JG, et al. Human orbital and anterior medial prefrontal cortex: Intrinsic connectivity parcellation and functional organization. *Brain Struct Funct*. 2017;1–20.
 34. Toronov V, Webb A, Choi JH, Wolf M, Safonova L, Wolf U, et al. Study of local cerebral hemodynamics by frequency-domain near-infrared spectroscopy and correlation with simultaneously acquired functional magnetic resonance imaging. *Opt Express*. 2001;9(8):417–27.
 35. Toronov VY, Zhang X, Webb AG. A spatial and temporal comparison of hemodynamic signals measured using optical and functional magnetic resonance imaging during activation in the human primary visual cortex. *Neuroimage* [Internet]. 2007;34(3):1136–48. Available from: <http://www.sciencedirect.com/science/article/pii/S105381190600841X>
 36. Eggebrecht AT, White BR, Ferradal SL, Chen C, Zhan Y, Snyder AZ, et al. A quantitative spatial comparison of high-density diffuse optical tomography and fMRI cortical mapping. *Neuroimage*. 2012;61(4):1120–8.
 37. Bailey DL, Townsend DW, Valk PE, Maisey MN. *Positron emission tomography*. Springer; 2005.
 38. Kalender WA. *X-ray computed*

- tomography. *Phys Med Biol*. 2006;51(13):R29–R29.
39. Schmitz CH, Löcker M, Lasker JM, Hielscher AH, Barbour RL. Instrumentation for fast functional optical tomography. *Rev Sci Instrum*. 2002;73(2):429–39.
 40. Joseph DK, Huppert TJ, Franceschini MA, Boas DA. Diffuse optical tomography system to image brain activation with improved spatial resolution and validation with functional magnetic resonance imaging. *Appl Opt*. 2006;45(31):8142–51.
 41. Steinbrink J, Wabnitz H, Obrig H, Villringer A, Rinneberg H. Determining changes in NIR absorption using a layered model of the human head. *Phys Med Biol*. 2001;46(3):879.
 42. Liebert A, Wabnitz H, Steinbrink J, Obrig H, Möller M, Macdonald R, et al. Time-resolved multidistance near-infrared spectroscopy of the adult head: intracerebral and extracerebral absorption changes from moments of distribution of times of flight of photons. *Appl Opt*. 2004;43(15):3037–47.
 43. Hielscher AH, Bluestone AY, Abdoulaev GS, Klose AD, Lasker J, Stewart M, et al. Near-infrared diffuse optical tomography. *Dis Markers*. 2002;18(5–6):313–37.
 44. Hueber DM, Franceschini MA, Ma HY, Zhang Q, Ballesteros JR, Fantini S, et al. Non-invasive and quantitative near-infrared haemoglobin spectrometry in the piglet brain during hypoxic stress, using a frequency-domain multidistance instrument. *Phys Med Biol*. 2001;46(1):41.
 45. Culver JP, Choe R, Holboke MJ, Zubkov L, Durduran T, Slemo A, et al. Three-dimensional diffuse optical tomography in the parallel plane transmission geometry: Evaluation of a hybrid frequency domain/continuous wave clinical system for breast imaging. *Med Phys*. 2003;30(2):235–47.
 46. Singer JR, Grunbaum FA, Kohn P, Zubelli JP. Image reconstruction of the interior of bodies that diffuse radiation. *Science* (80-). 1990;248(4958):990–3.
 47. Arridge SR, Schweiger M, Delpy DT. Iterative reconstruction of near infrared absorption images. In: *Proc SPIE*. 1992. p. 372–83.
 48. Bluestone AY, Abdoulaev G, Schmitz CH, Barbour RL, Hielscher AH. Three-dimensional optical tomography of hemodynamics in the human head. *Opt Express*. 2001;9(6):272–86.
 49. Barbour RL, Graber HL, Pei Y, Zhong S, Schmitz CH. Optical tomographic imaging of dynamic features of dense-scattering media. *JOSA A*. 2001;18(12):3018–36.
 50. Tanosaki M, Hoshi Y, Iguchi Y, Oikawa Y, Oda I, Oda M. Variation of temporal characteristics in human cerebral hemodynamic responses to electric median nerve stimulation: a near-infrared spectroscopic study. *Neurosci Lett*. 2001;316(2):75–8.
 51. O’Leary MA, Boas DA, Chance B, Yodanis AG. Experimental images of heterogeneous turbid media by frequency-domain diffusing-photon tomography. *Opt Lett*. 1995;20(5):426–8.
 52. Boas DA, Culver JP, Stott JJ, Dunn AK. Three dimensional Monte Carlo code for photon migration through

- complex heterogeneous media including the adult human head. *Opt Express* [Internet]. 2002 Feb;10(3):159–70. Available from: <http://www.opticsexpress.org/abstract.cfm?URI=oe-10-3-159>
53. Eggebrecht AT, Ferradal SL, Robichaux-Viehoever A, Hassanpour MS, Dehghani H, Snyder AZ, et al. Mapping distributed brain function and networks with diffuse optical tomography. *Nat Photonics*. 2014;8(6):448–54.
 54. Arridge SR, Schweiger M. Photon-measurement density functions. Part 2: Finite-element-method calculations. *Appl Opt*. 1995;34(34):8026–37.
 55. Hielscher AH, Klose AD, Hanson KM. Gradient-based iterative image reconstruction scheme for time-resolved optical tomography. *IEEE Trans Med Imaging*. 1999;18(3):262–71.
 56. Habermehl C, Steinbrink J, Müller K-R, Haufe S. Optimizing the regularization for image reconstruction of cerebral diffuse optical tomography. *J Biomed Opt*. 2014;19(9):96006.
 57. Yamashita O, Shimokawa T, Aisu R, Amita T, Inoue Y, Sato M. Multi-subject and multi-task experimental validation of the hierarchical Bayesian diffuse optical tomography algorithm. *Neuroimage*. 2016;135:287–99.
 58. Hernandez-Martin E, Marcano F, Casanova O, Modrono C, Plata-Bello J, Gonzalez-Mora JL. Comparing diffuse optical tomography and functional magnetic resonance imaging signals during a cognitive task: pilot study. *Neurophotonics*. 2017 Jan;4(1):15003.
 59. Custo A, Boas DA, Tsuzuki D, Dan I, Mesquita R, Fischl B, et al. Anatomical atlas-guided diffuse optical tomography of brain activation. *Neuroimage*. 2010;49(1):561–7.
 60. Habermehl C, Holtze S, Steinbrink J, Koch SP, Obrig H, Mehnert J, et al. Somatosensory activation of two fingers can be discriminated with ultrahigh-density diffuse optical tomography. *Neuroimage*. 2012;59(4):3201–11.
 61. Yuan Z, Zhang Q, Sobel ES, Jiang H. Tomographic x-ray-guided three-dimensional diffuse optical tomography of osteoarthritis in the finger joints. *J Biomed Opt*. 2008;13(4):44006.
 62. Kavuri VC, Liu H. Hierarchical clustering method to improve transrectal ultrasound-guided diffuse optical tomography for prostate cancer imaging. *Acad Radiol*. 2014;21(2):250–62.
 63. Siegel AM, Culver JP, Mandeville JB, Boas DA. Temporal comparison of functional brain imaging with diffuse optical tomography and fMRI during rat forepaw stimulation. *Phys Med Biol*. 2003;48(10):1391.
 64. Franceschini MA, Joseph DK, Huppert TJ, Diamond SG, Boas DA. Diffuse optical imaging of the whole head. *J Biomed Opt*. 2006;11(5):54007.
 65. Boecker M, Buecheler MM, Schroeter ML, Gauggel S. Prefrontal brain activation during stop-signal response inhibition: an event-related functional near-infrared spectroscopy study. *Behav Brain Res*. 2007;176(2):259–66.
 66. Power SD, Kushki A, Chau T. Towards a system-paced near-infrared spectroscopy brain-computer interface: differentiating prefrontal

- activity due to mental arithmetic and mental singing from the no-control state. *J Neural Eng.* 2011;8(6):66004.
67. Huppert TJ, Hoge RD, Diamond SG, Franceschini MA, Boas DA. A temporal comparison of BOLD, ASL, and NIRS hemodynamic responses to motor stimuli in adult humans. *Neuroimage* [Internet]. 2006 Jan;29(2):368–82. Available from: <http://www.sciencedirect.com/science/article/pii/S1053811905005823>
 68. Becerra L, Harris W, Joseph D, Huppert T, Boas DA, Borsook D. Diffuse optical tomography of pain and tactile stimulation: Activation in cortical sensory and emotional systems. *Neuroimage* [Internet]. 2008 Jun;41(2):252–9. Available from: <http://www.sciencedirect.com/science/article/pii/S1053811908001006>
 69. Strangman G, Culver JP, Thompson JH, Boas DA. A quantitative comparison of simultaneous BOLD fMRI and NIRS recordings during functional brain activation. *Neuroimage.* 2002;17(2):719–31.
 70. Gagnon L, Yücel MA, Boas DA, Cooper RJ. Further improvement in reducing superficial contamination in NIRS using double short separation measurements. *Neuroimage* [Internet]. 2014 Jan;85:127–35. Available from: <http://www.sciencedirect.com/science/article/pii/S1053811913001201>
 71. Hisakata R, Nishida S, Johnston A. An Adaptable Metric Shapes Perceptual Space. *Curr Biol* [Internet]. 2016 Jul;26(14):1911–5. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4963211/>
 72. Plichta MM, Heinzl S, Ehlis A-C, Pauli P, Fallgatter AJ. Model-based analysis of rapid event-related functional near-infrared spectroscopy (NIRS) data: a parametric validation study. *Neuroimage.* 2007;35(2):625–34.
 73. Plata-Bello J, Modroño C, Hernández-Martín E, Pérez-Martín Y, Fariña H, Castañón-Pérez A, et al. The mirror neuron system also rests. *Brain Struct Funct.* 2017;222(5):2193–202.
 74. Sasai S, Homae F, Watanabe H, Taga G. Frequency-specific functional connectivity in the brain during resting state revealed by NIRS. *Neuroimage.* 2011;56(1):252–7.
 75. Grant PE, Roche-Labarbe N, Surova A, Themelis G, Selb J, Warren EK, et al. Increased cerebral blood volume and oxygen consumption in neonatal brain injury. *J Cereb Blood Flow Metab.* 2009;29(10):1704–13.
 76. Hebden JC, Gibson A, Yusof RM, Everdell N, Hillman EMC, Delpy DT, et al. Three-dimensional optical tomography of the premature infant brain. *Phys Med Biol.* 2002;47(23):4155.
 77. Maidan I, Bernad-Elazari H, Gazit E, Giladi N, Hausdorff JM, Mirelman A. Changes in oxygenated hemoglobin link freezing of gait to frontal activation in patients with Parkinson disease: an fNIRS study of transient motor-cognitive failures. *J Neurol.* 2015;262(4):899–908.
 78. Hock C, Villringer K, Müller-Spahn F, Wenzel R, Heekeren H, Schuh-Hofer S, et al. Decrease in parietal cerebral hemoglobin oxygenation during performance of a verbal fluency task in patients with Alzheimer's disease monitored by means of near-infrared spectroscopy (NIRS) - Correlation

- with simultaneous rCBF-PET measurements. *Brain Res* [Internet]. 1997 May;755(2):293–303. Available from: <http://www.sciencedirect.com/science/article/pii/S0006899397001224>
79. Sokol DK, Markand ON, Daly EC, Luerssen TG, Malkoff MD. Near infrared spectroscopy (NIRS) distinguishes seizure types. *Seizure*. 2000;9(5):323–7.
 80. Shidoh S, Akiyama T, Horiguchi T, Ohira T, Yoshida K. The process of change in hemodynamics after revascularization in the ischemic brain. *Neuroreport*. 2015;26(11):629–33.
 81. Herrmann MJ, Walter A, Ehlis A-C, Fallgatter AJ. Cerebral oxygenation changes in the prefrontal cortex: effects of age and gender. *Neurobiol Aging*. 2006;27(6):888–94.
 82. Koike S, Nishimura Y, Takizawa R, Yahata N, Kasai K. Near-infrared spectroscopy in schizophrenia: a possible biomarker for predicting clinical outcome and treatment response. *Front psychiatry*. 2013;4.
 83. Tuscan L-A, Herbert JD, Forman EM, Juarascio AS, Izzetoglu M, Schultheis M. Exploring frontal asymmetry using functional near-infrared spectroscopy: a preliminary study of the effects of social anxiety during interaction and performance tasks. *Brain Imaging Behav*. 2013;7(2):140–53.
 84. Zhang Q, Brukilacchio TJ, Li A, Stott JJ, Chaves T, Hillman E, et al. Coregistered tomographic x-ray and optical breast imaging: initial results. *J Biomed Opt*. 2005;10(2):24033–240339.
 85. Taroni P, Torricelli A, Spinelli L, Pifferi A, Arpaia F, Danesini G, et al. Time-resolved optical mammography between 637 and 985 nm: clinical study on the detection and identification of breast lesions. *Phys Med Biol*. 2005;50(11):2469.
 86. Arenth PM, Ricker JH, Schultheis MT. Applications of functional near-infrared spectroscopy (fNIRS) to neurorehabilitation of cognitive disabilities. *Clin Neuropsychol*. 2007;21(1):38–57.
 87. Suzuki M, Miyai I, Ono T, Oda I, Konishi I, Kochiyama T, et al. Prefrontal and premotor cortices are involved in adapting walking and running speed on the treadmill: An optical imaging study. *Neuroimage* [Internet]. 2004 Nov;23(3):1020–6. Available from: <http://www.sciencedirect.com/science/article/pii/S1053811904003672?via%3Dihub>
 88. Pedersen PM, Stig Jørgensen H, Nakayama H, Raaschou HO, Olsen TS. Aphasia in acute stroke: incidence, determinants, and recovery. *Ann Neurol*. 1995;38(4):659–66.
 89. Bonato C, Miniussi C, Rossini PM. Transcranial magnetic stimulation and cortical evoked potentials: a TMS/EEG co-registration study. *Clin Neurophysiol*. 2006;117(8):1699–707.
 90. Bohning DE, Shastri A, Nahas Z, Lorberbaum JP, Andersen SW, Dannels WR, et al. Echoplanar BOLD fMRI of brain activation induced by concurrent transcranial magnetic stimulation. *Invest Radiol*. 1998;33(6):336–40.