

RESEARCH ARTICLE

The potential role of the paxillin paralog Hic-5 in progression of hepatocellular carcinoma

Authors

Jia-Ru Wu[&], Chi-Tan Hu[#], Ren-In You[&], Chi-Wen Chen[%], Wen-Sheng Wu^{&1}

Affiliations:

Research Centre for Hepatology, Department of Internal Medicine, Buddhist Tzu Chi General Hospital. School of Medicine, Tzu Chi University, Hualien, Taiwan. &Institute of Medical Sciences, Department of Laboratory Medicine and Biotechnology, College of Medicine, Tzu Chi University, Hualien, Taiwan. % School of Chinese Medicine, China Medical University, Taiwan

Correspondence:

Wen-Sheng Wu. Institute of medical biotechnology, college of Medicine, Tzu Chi University No. 701, Chung Yang Rd, Sec 3, Hualien 970, Taiwan.

E-mail: wuwstcu1234@yahoo.com.tw ; Phone: 8867-03-8565301 ext 2327 ; Fax: 8867-03-8571917

ABSTRACT

Hepatocellular carcinoma (HCC) is one of the most common causes of death from cancer worldwide. The poor prognosis of HCC is due to high recurrence rate mainly caused by intrahepatic metastasis. Paxillin was known to be a central adaptor protein for mediating focal adhesion (FA) signal required for HCC progression. However, target therapy aiming at paxillin seems unfeasible due to its ubiquitous tissue expression and essential biological functions. Within the paxillin superfamily, hydrogen peroxide inducible clone-5 (Hic-5) is the most homologous to paxillin. This review summarises the recent findings relevant to the differential biochemical and biological roles of Hic-5 and paxillin. Given the structure similarity between Hic-5 and paxillin, Hic-5 shares many of the characteristics of paxillin, including the localization of Hic-5 at focal adhesions and similar FA binding factors. However, some of the regulatory mechanisms and molecular functions of Hic-5 are rather different from those of paxillin. These might explain the differential roles of both adaptors in regulating various pathophysiological processes. Interestingly, both adaptors might play distinct but complementary roles in tumor progression. Due to the more limited tissue distribution of Hic-5, it can be a more suitable therapeutic target for preventing HCC progression.

Key words: Hepatocellular carcinoma; Metastasis; Paxillin; Hic-5; focal adhesions.

Introduction

Hepatocellular carcinoma (HCC) is one of the most common causes of death from cancer worldwide. The mortality rate is very high in China, Taiwan and Southeast Asia[1]. Recently, the incidence of HCC was increasing in countries such as Japan, Italy, France, Switzerland, United Kingdom and the United States[2]. The poor prognosis of HCC is due to high recurrence rate mainly caused by intrahepatic metastasis (about 80%) or extrahepatic metastasis (about 20%)[3]. Therefore, prevention of metastasis is critical for HCC management. To address the issue, the suitable molecular targets within the molecular pathways leading to HCC metastasis are needed to be identified.

Tumor metastasis occurs *via* complicated processes, including epithelial mesenchymal transition (EMT), migration and invasion of primary tumor, followed by intravasation, extravasation and colonization at the metastatic loci. The tumor microenvironment in HCC contains a lot of metastatic factors produced by interaction of primary cell with inflammatory cells, stromal cell and extracellular matrix [4, 5]. Some of the metastatic factors including transforming growth factor β (TGF β) [6], hepatocyte growth factor (HGF) [7, 8], vascular endothelial growth factor (VEGF) and epidermal growth factor (EGF) [9] and integrin engagement [10-15], are capable of triggering HCC progression. A lot of signaling components in the FA such as RacGTPase [16, 17], focal adhesion kinase (FAK) [18-22], Src [23], Pyk2 [24] and paxillin [25] are known to be responsible for mediating HCC progression triggered by these metastatic factors.

Among the aforementioned FA signal components, Paxillin is responsible for mediating signal cross talk between integrin and

metastatic factors [27]. The role of paxillin in tumor progression of HCC has been demonstrated in a lot of studies. Paxillin phosphorylation at Ser178 mediated by Jun N-terminal kinase (JNK) was involved in the HGF [30] and P21-activated protein kinase (PAK) triggered tumor progression of HCC [31]. Recently, paxillin was found to be a mediator for the actopaxin-triggered HCC metastasis [25].

Within the paxillin superfamily, Hic-5 (hydrogen peroxide inducible clone-5), is the most homologous to paxillin. Hic-5 was initially identified as one of the TGF β 1 and hydrogen peroxide-inducible genes [32]. As paxillin, Hic 5 is also an adaptor molecule essential for triggering progression of tumors [33, 34] including HCC [35]. This review summarizes the recent findings relevant to differential biochemical and biological roles between Hic-5 and paxillin in mediating a lot of cellular phenotypes. Specifically the complementary role of both adaptors in tumor progression is addressed. Moreover, the possibility of Hic-5 to be a promising therapeutic target for prevention of HCC metastasis is highlighted.

Comparison of the biochemical properties between Hic-5 and paxillin

Differential structure and binding properties

Paxillin comprises numerous discrete structural domains for scaffolding FA components in response to integrin engagement and growth factors [36]. The C-terminal half of paxillin contains four LIM domains, serving as binding sites for several structural and regulatory proteins such as tubulin and the protein tyrosine phosphatase (PTP) PEST. The N-terminus of paxillin is composed of five

leucine- and aspartate-rich LD motifs (LD1-LD5), with multiple tyrosine, serine and threonine phosphorylation sites for recruiting a lot of signal molecules including Src tyrosine kinase, focal adhesion kinase (FAK), receptor for activated C kinase 1(RACK1), JNK, p-38 and Abl. Hic-5 shares the same 11-exon genomic organization as paxillin with minor differences in the number of LD domains in the N-terminal region (five for paxillin and four for HIC-5) [36].

Given the structure similarity between Hic-5 and paxillin, Hic-5 shares many of the characteristics of paxillin, including the localization at FA and similar FA binding factors such as protein tyrosine kinase 2 beta (PYK2), c-Src tyrosine kinase (Csk), FAK, Arf GAP1 (GIT-1) [37] and PTP-PEST [38]. However, there are paxillin interacting FA components such as Crk and Src which can not bind Hic5. One distinct binding activity of Hic-5 was ascribed to its LIM domain by which Hic-5 may form LIM-LIM hetero-oligomers with LIM-only proteins such as PINCH or CRP2 [41]. In contrast, LIM4 of paxillin cannot form oligomers and does not interact with PINCH or CRP2 [41].

Differential phosphorylation pattern involved in FA signaling

The phosphorylation patterns of Hic-5 and paxillin induced by extracellular stimuli are also rather different. Upon integrin engagement, paxillin becomes tyrosine phosphorylated, primarily on tyrosine residues 31 and 118 (Y31 and Y118, respectively at LD1), in a FAK- and Src-dependent manner, resulting in activation of a lot of critical signal cascades for cell spreading and motility [42, 43]. In contrast, Hic-5 does not exhibit the aforementioned phosphorylation patterns probably due to lack of cognate tyrosine

residues. However, phosphorylation of Hic-5 may occur through PYK2 following hyperosmotic stress [47] and platelet activation [48].

In addition to the discrepancies of the molecular structure, interaction and phosphorylation pattern for FA signaling as described above, there are a lot of different biochemical properties between both Hic-5 and paxillin including tissue specific distribution, regulation of gene expression, interaction with critical signal cascade, and the impacts on cellular phenotypes.

Differential tissue specific distribution between paxillin and Hic-5

Tissue expression of paxillin is broader than that of Hic-5

Whereas paxillin is ubiquitously expressed in most tissue and cell types, Hic-5 is enriched only in certain tissue such as smooth muscle (in particular the vasculature), large intestine and uterus and relatively high in the lung and spleen [33]. This implicates that the biological roles of paxillin are broader than Hic-5. Indeed, paxillin ablation causing early embryonic lethality, while Hic-5 knockout exhibits only very mild vascular defects [33].

Differential pattern of inducible gene expression between paxillin and Hic-5

One distinct regulatory mechanism of Hic-5 is the inducible gene expression by a lot of extracellular stimuli. In the most early studies, Hic-5 gene expression was found to be induced by reactive oxygen species (ROS) [59], as its name suggests. Hic-5 expression can also be induced during TGF β 1-induced senescence of

osteoblastic cell line [32], angiotensin II-induced abdominal aortic aneurysm (AAA) development [60], methylmercury-induced ER stress [61] and *Escherichia coli*-induced prostatic inflammation [62]. Also, epithelial expression of Hic-5 in mouse and human prostate tissues was elevated after castration, leading to epithelial regression through the repression of c-myc gene [63]. In contrast, the evidence regarding inducible gene expression of paxillin was very rare, with only one report demonstrating that paxillin mRNA is induced by TGF β as shown on a retrovirus-mediated gene trap screen [64].

Differential impacts of Hic-5 and paxillin on intracellular signal cascades

The impacts of Hic-5 gene expression on essential signal cascades are more prominent than those of paxillin. Whereas Hic-5 gene expression and nuclear translocation can be induced by ROS as described above, it appears that Hic-5 may also positively regulate ROS generation in the focal adhesion [65]. In this context, Hic-5 serves as an adaptor for association of TRAF4 and p47^{phox} which initiate Rho GTPase activation required for NADP oxidase-dependent-ROS production. The ROS generated in turn targets the redox-sensitive phosphatase PTP-PEST in FA, establishing a positive feedback cycle that facilitates Rac1 activation leading to sustained MAPK activation and cell migration [65]. Similarly, whereas TGF β was known to be an inducer of Hic-5 [66, 67], TGF β -induced signaling can also be positively regulated by Hic-5. Previously, Hic-5 was found to promote TGF β -induced signaling by binding to and inactivating the inhibitory Smads, Smad3 [68] and Smad7 [69] leading to enhanced TGF- β / Smad2 signaling required for EMT. Also, Hic-5 may bind to Smads 1, 5 and 8, for repressing bone morphogenetic protein

(BMP) signaling [70]. In addition, Hic-5 may serve as a scaffold protein that specifically activates the MAPK cascade. For example, in a model for abdominal aortic aneurysm (AAA), Hic-5 interacted specifically with JNK and its upstream kinase MAPKK4 to trigger the downstream signaling [60].

Differential impacts of Hic-5 and paxillin on cellular phenotypes

Given the aforementioned discrepancies of Hic-5 and paxillin in biochemical properties, it is anticipated to find the divergence of both adaptors in regulation of cellular phenotypes such as cytoskeletal organization, cell adhesion, cell migration and cell growth. Whereas paxillin is well known to be required for cell adhesion, Hic-5 may suppress excess changes in cytoskeleton structure by antagonizing paxillin [71, 72]. The regulation of cell growth by paxillin and Hic-5 is also antagonistic. In general, paxillin is responsible for transducing both adhesion-dependent and independent growth signaling [73-75]. On the contrary, Hic-5 serves as one of the fail-safe system for the adhesion dependence of cell growth by negatively regulating the cell cycle positive regulator, cyclin D1 [41].

The role of Hic-5 and paxillin in tumor progression

Distinct but complementary roles of Hic-5 and paxillin in tumor progression

As described in previous section, paxillin is well known to be involved in tumor progression, ascribed to its crucial role in mediating focal adhesion signaling [73]. Recently, the role of Hic-5 tumor progression was also emerging [33, 77, 78]. Both paxillin and Hic-5 expressed in a variety of invasive/metastatic cancers, including

breast, lung, and prostate tumors and are regarded as potential prognostic markers and therapeutic targets [33]. However, due to the differential biochemical and biological properties, the mechanisms for the two adaptors in triggering tumor progression are also rather divergent. Whereas tyrosine phosphorylation of paxillin is essential for mediating EMT and cell migration [79, 80], gene expression of Hic-5 was required for these processes [65, 81]. In the cell culture systems, Hic-5 is highly detectable in mesenchymal cell lines but absent in epithelial cell lines. Moreover, Hic-5 expression can be induced by TGF β leading to EMT, invadopodia formation, cell migration, and invasion [67]. This is ascribed to that Hic-5 can bind and inactivate the inhibitory Smads to enhance TGF- β receptor signaling as described above [68-70]. In addition, the induction of Hic-5 expression by TGF β was dependent on RhoA/ROCK1 [79]. Furthermore, ectopic expression of Hic-5 is sufficient to promote normal mammary cells to undergo EMT in the absence of TGF- β [67] and stimulate cell migration of NMuMG cells [34]. In contrast, overexpression of paxillin is unable to induce the transition to mesenchymal phenotype [82]. In spite of these discrepancies, it appears that both adaptors may contribute metastatic change in a concerted manner. This notion was supported by the evidence that Hic-5 may cooperate with paxillin to regulate metastasis of breast cancer

[78]. Also, the FA structure protein vinculin is able to interact with paxillin and Hic-5 via Rac1 and RhoA, respectively, required for FA turnover and cell migration [83].

The potential role of Hic-5 in HCC progression

The involvement of paxillin in HCC progression is evident as described in the **Introduction section**. Recently, the role of Hic-5 is also emerging. One report demonstrated that the expression and phosphorylation of Hic-5 were upregulated in HCCs overexpressing proline-rich tyrosine kinase 2 (Pyk2) [35], a member of the FAK family known to be involved in HCC metastasis [24, 84, 85]. Also, TGF- β , which can induce Hic-5 for malignant transformation [67], is recently highlighted to be one of the critical metastatic factors triggering HCC progression [86-88]. In our recent report, we found Hic-5 mediated ROS-JNK signaling and serve as a potential therapeutic target for prevention of HCC progression [89]. In this study, Hic-5 positively cross-talks with ROS to trigger sustained ERK (MAPK) signaling for HCC progression induced by hepatocyte growth factor (HGF) (see Scheme in Fig.1). Notably, ROS is also a well-known mediator of tumor progression including HCC [89-91]. Together, these studies implicated that Hic-5 is one of the key factors in HCC progression.

Fig. 1 Proposed mechanism for Hic-5 to regulate ROS-mediated sustained signaling in a positive feedback circuit

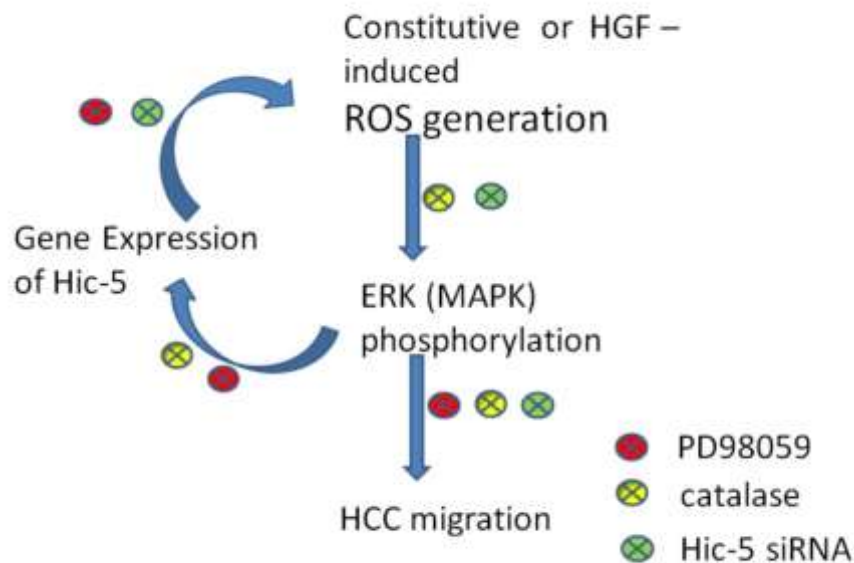


Fig. 1 Constitutive or HGF-induced ROS generation triggers ERK(MAPK) activation, leading to gene expression of Hic-5, which then contribute to sustained ROS generation and signaling transduction required for cell migration of HCC. This signal circuit was delineated by using inhibitors or siRNA against the indicated signal components. Circles with red, yellow and green represent inhibitors of ERK (PD98059), ROS scavenger (catalase) and Hic-5 siRNA, respectively. In the relevant experiments, activities (in case of ERK and ROS) or expression (in case of Hic-5) of each component in the circuit can be suppressed by inhibitors of the two upstream components. Cell migration is prevented by all inhibitors. HGF: hepatocyte growth factor.

Conclusions and perspectives

Since Hic-5 is the most homologous to paxillin, it can be expected that both adaptors share similar biological properties. However, a lot of the regulatory mechanism and molecular function of Hic-5 are not shared by paxillin (summarized in Table 1). Importantly, both

adaptors play distinct but complementary role in tumor progression. Due to the more limited distribution of Hic-5, it can be a more promising therapeutic target than paxillin for preventing HCC progression.

Table 1. Comparison of Pathophysiological characteristics of paxillin and Hic-5

Pathophysiological Characteristics	Hic-5	paxillin
Tissue distribution	limited	Ubiquitous
Stimulators	ROS, TGF β , HGF and others	Multiple growth factors/cytokines /integrin engagement
Regulatory mechanisms	Gene expression (major) /phosphorylation (rare)	Phosphorylation (major)/Gene expression (rare)
Effect on Cell growth	Safe guarding adhesion dependence of cell growth (-)	Anchorage- dependent and independent cell growth (+)
Effects on FA phenotype	EMT/ migration (+) Cell spreading (-) Cytoskeletal change (-)	EMT/migration (+) Cell spreading (+) Cytoskeletal change (+)
Involvement in tumor progression	metastasis of breast cancer (+) HCC progression (+)	metastasis of breast cancer (+) HCC progression (+)

(+), and (-) represent positive and negative regulation, respectively, of the indicated phenotypes by Hic-5 or paxillin; ROS:reactive oxygen species; HCC hepatocellular carcinoma

Reference

1. Bosch, F.X., Ribes, J., Diaz, M., and Cleries, R. (2004). Primary liver cancer: worldwide incidence and trends. *Gastroenterology* *127*, S5-S16.
2. El-Serag, H.B., Davila, J.A., Petersen, N.J., and McGlynn, K.A. (2003). The continuing increase in the incidence of hepatocellular carcinoma in the United States: an update. *Ann Intern Med* *139*, 817-823.
3. Tang, Z.Y., Ye, S.L., Liu, Y.K., Qin, L.X., Sun, H.C., Ye, Q.H., Wang, L., Zhou, J., Qiu, S.J., Li, Y., Ji, X.N., Liu, H., Xia, J.L., Wu, Z.Q., Fan, J., Ma, Z.C., Zhou, X.D., Lin, Z.Y., and Liu, K.D. (2004). A decade's studies on metastasis of hepatocellular carcinoma. *J Cancer Res Clin Oncol* *130*, 187-196.
4. Fornaro, L., Vivaldi, C., Caparello, C., Sacco, R., Rotella, V., Masettini, G., Luchi, S., Baldini, E.E., Falcone, A., and Masi, G. (2014). Dissecting signaling pathways in hepatocellular carcinoma: new perspectives in medical therapy. *Future Oncol* *10*, 285-304.
5. Moeini, A., Cornella, H., and Villanueva, A. (2012). Emerging signaling pathways in hepatocellular carcinoma. *Liver Cancer* *1*, 83-93.
6. Mazzocca, A., Antonaci, S., and Giannelli, G. (2012). The TGF-beta signaling pathway as a pharmacological target in a hepatocellular carcinoma. *Curr Pharm Des* *18*, 4148-4154.
7. Goyal, L., Muzumdar, M.D., and Zhu, A.X. (2013). Targeting the HGF/c-MET pathway in hepatocellular carcinoma. *Clin Cancer Res* *19*, 2310-2318.
8. Giordano, S., and Columbano, A. (2013). Met as a therapeutic target in HCC: facts and hopes. *J Hepatol* *60*, 442-452.
9. Bronte, F., Bronte, G., Cusenza, S., Fiorentino, E., Rolfo, C., Cicero, G., Bronte, E., Di Marco, V., Firenze, A., Angarano, G., Fontana, T., and Russo, A. (2014). Targeted therapies in hepatocellular carcinoma. *Curr Med Chem* *21*, 966-974.
10. Zhang, X., Cheng, S.L., Bian, K., Wang, L., Zhang, X., Yan, B., Jia, L.T., Zhao, J., Gammoh, N., Yang, A.G., and Zhang, R. (2014). MicroRNA-26a promotes anoikis in human hepatocellular carcinoma cells by targeting alpha5 integrin. *Oncotarget*.
11. Fu, Y., Feng, M.X., Yu, J., Ma, M.Z., Liu, X.J., Li, J., Yang, X.M., Wang, Y.H., Zhang, Y.L., Ao, J.P., Xue, F., Qin, W., Gu, J., Xia, Q., and Zhang, Z.G. (2014). DNA methylation-mediated silencing of matricellular protein dermatopontin promotes hepatocellular carcinoma metastasis by alpha3beta1

- integrin-Rho GTPase signaling. *Oncotarget* 5, 6701-6715.
12. Patman, G. (2014). Liver: loss of integrin beta1 impairs liver regeneration and HCC progression. *Nat Rev Gastroenterol Hepatol* 11, 392.
 13. Bogorad, R.L., Yin, H., Zeigerer, A., Nonaka, H., Ruda, V.M., Zerial, M., Anderson, D.G., and Kotliansky, V. (2014). Nanoparticle-formulated siRNA targeting integrins inhibits hepatocellular carcinoma progression in mice. *Nat Commun* 5, 3869.
 14. Hu, C.T., Wu, J.R., Cheng, C.C., Wang, S., Wang, H.T., Lee, M.C., Wang, L.J., Pan, S.M., Chang, T.Y., and Wu, W.S. (2011). Reactive oxygen species-mediated PKC and integrin signaling promotes tumor progression of human hepatoma HepG2. *Clin Exp Metastasis* 28, 851-863.
 15. Wu, Y., Qiao, X., Qiao, S., and Yu, L. (2011). Targeting integrins in hepatocellular carcinoma. *Expert Opin Ther Targets* 15, 421-437.
 16. Wang, S.M., Ooi, L.L., and Hui, K.M. (2011). Upregulation of Rac GTPase-activating protein 1 is significantly associated with the early recurrence of human hepatocellular carcinoma. *Clin Cancer Res* 17, 6040-6051.
 17. Lee, T.K., Poon, R.T., Yuen, A.P., Man, K., Yang, Z.F., Guan, X.Y., and Fan, S.T. (2006). Rac activation is associated with hepatocellular carcinoma metastasis by up-regulation of vascular endothelial growth factor expression. *Clin Cancer Res* 12, 5082-5089.
 18. Fujii, T., Koshikawa, K., Nomoto, S., Okochi, O., Kaneko, T., Inoue, S., Yatabe, Y., Takeda, S., and Nakao, A. (2004). Focal adhesion kinase is overexpressed in hepatocellular carcinoma and can be served as an independent prognostic factor. *J Hepatol* 41, 104-111.
 19. Yao, W.L., Ko, B.S., Liu, T.A., Liang, S.M., Liu, C.C., Lu, Y.J., Tzean, S.S., Shen, T.L., and Liou, J.Y. (2014). Cordycepin suppresses integrin/FAK signaling and epithelial-mesenchymal transition in hepatocellular carcinoma. *Anticancer Agents Med Chem* 14, 29-34.
 20. Ding, J., Huang, S., Wu, S., Zhao, Y., Liang, L., Yan, M., Ge, C., Yao, J., Chen, T., Wan, D., Wang, H., Gu, J., Yao, M., Li, J., Tu, H., and He, X. (2010). Gain of miR-151 on chromosome 8q24.3 facilitates tumour cell migration and spreading through downregulating RhoGDI. *Nat Cell Biol* 12, 390-399.
 21. Huang, J., Zheng, D.L., Qin, F.S., Cheng, N., Chen, H., Wan, B.B., Wang, Y.P.,

- Xiao, H.S., and Han, Z.G. (2010). Genetic and epigenetic silencing of SCARA5 may contribute to human hepatocellular carcinoma by activating FAK signaling. *J Clin Invest* 120, 223-241.
22. Jia, Y.L., Shi, L., Zhou, J.N., Fu, C.J., Chen, L., Yuan, H.F., Wang, Y.F., Yan, X.L., Xu, Y.C., Zeng, Q., Yue, W., and Pei, X.T. (2011). Epimorphin promotes human hepatocellular carcinoma invasion and metastasis through activation of focal adhesion kinase/extracellular signal-regulated kinase/matrix metalloproteinase-9 axis. *Hepatology* 54, 1808-1818.
23. Cortese, R., Almendros, I., Wang, Y., and Gozal, D. (2014). Tumor circulating DNA profiling in xenografted mice exposed to intermittent hypoxia. *Oncotarget*.
24. Cao, J., Chen, Y., Fu, J., Qian, Y.W., Ren, Y.B., Su, B., Luo, T., Dai, R.Y., Huang, L., Yan, J.J., Wu, M.C., Yan, Y.Q., and Wang, H.Y. (2013). High expression of proline-rich tyrosine kinase 2 is associated with poor survival of hepatocellular carcinoma via regulating phosphatidylinositol 3-kinase/AKT pathway. *Ann Surg Oncol* 20 Suppl 3, S312-323.
25. Ng, L., Tung-Ping Poon, R., Yau, S., Chow, A., Lam, C., Li, H.S., Chung-Cheung Yau, T., Law, W.L., and Pang, R. (2013). Suppression of actopaxin impairs hepatocellular carcinoma metastasis through modulation of cell migration and invasion. *Hepatology* 58, 667-679.
26. Mitra, S.K., Hanson, D.A., and Schlaepfer, D.D. (2005). Focal adhesion kinase: in command and control of cell motility. *Nat Rev Mol Cell Biol* 6, 56-68.
27. Deakin, N.O., and Turner, C.E. (2008). Paxillin comes of age. *J Cell Sci* 121, 2435-2444.
28. Bi, Y., Han, Y., Bi, H., Gao, F., and Wang, X. (2014). miR-137 impairs the proliferative and migratory capacity of human non-small cell lung cancer cells by targeting paxillin. *Hum Cell* 27, 95-102.
29. Li, H.G., Xie, D.R., Shen, X.M., Li, H.H., Zeng, H., and Zeng, Y.J. (2005). Clinicopathological significance of expression of paxillin, syndecan-1 and EMMPRIN in hepatocellular carcinoma. *World J Gastroenterol* 11, 1445-1451.
30. Hu, C.T., Cheng, C.C., Pan, S.M., Wu, J.R., and Wu, W.S. (2013). PKC mediates fluctuant ERK-paxillin signaling for hepatocyte growth factor-induced migration of hepatoma cell HepG2. *Cell Signal* 25, 1457-1467.

31. Ching, Y.P., Leong, V.Y., Lee, M.F., Xu, H.T., Jin, D.Y., and Ng, I.O. (2007). P21-activated protein kinase is overexpressed in hepatocellular carcinoma and enhances cancer metastasis involving c-Jun NH2-terminal kinase activation and paxillin phosphorylation. *Cancer Res* 67, 3601-3608.
32. Shibamura, M., Mashimo, J., Kuroki, T., and Nose, K. (1994). Characterization of the TGF beta 1-inducible hic-5 gene that encodes a putative novel zinc finger protein and its possible involvement in cellular senescence. *J Biol Chem* 269, 26767-26774.
33. Deakin, N.O., Pignatelli, J., and Turner, C.E. (2012). Diverse roles for the paxillin family of proteins in cancer. *Genes Cancer* 3, 362-370.
34. Tumbarello, D.A., Brown, M.C., Hetey, S.E., and Turner, C.E. (2005). Regulation of paxillin family members during epithelial-mesenchymal transformation: a putative role for paxillin delta. *J Cell Sci* 118, 4849-4863.
35. Sun, C.K., Ng, K.T., Lim, Z.X., Cheng, Q., Lo, C.M., Poon, R.T., Man, K., Wong, N., and Fan, S.T. (2011). Proline-rich tyrosine kinase 2 (Pyk2) promotes cell motility of hepatocellular carcinoma through induction of epithelial to mesenchymal transition. *PLoS One* 6, e18878.
36. Brown, M.C., and Turner, C.E. (2004). Paxillin: adapting to change. *Physiol Rev* 84, 1315-1339.
37. Nishiya, N., Shirai, T., Suzuki, W., and Nose, K. (2002). Hic-5 interacts with GIT1 with a different binding mode from paxillin. *J Biochem* 132, 279-289.
38. Nishiya, N., Iwabuchi, Y., Shibamura, M., Cote, J.F., Tremblay, M.L., and Nose, K. (1999). Hic-5, a paxillin homologue, binds to the protein-tyrosine phosphatase PEST (PTP-PEST) through its LIM 3 domain. *J Biol Chem* 274, 9847-9853.
39. Thomas, S.M., Hagel, M., and Turner, C.E. (1999). Characterization of a focal adhesion protein, Hic-5, that shares extensive homology with paxillin. *J Cell Sci* 112 (Pt 2), 181-190.
40. Liu, S., Thomas, S.M., Woodside, D.G., Rose, D.M., Kiosses, W.B., Pfaff, M., and Ginsberg, M.H. (1999). Binding of paxillin to alpha4 integrins modifies integrin-dependent biological responses. *Nature* 402, 676-681.
41. Shibamura, M., Mori, K., and Nose, K. (2012). HIC-5: A Mobile Molecular Scaffold Regulating the Anchorage Dependence of Cell Growth. *Int J Cell Biol* 2012, 426138.

42. Brugnera, E., Haney, L., Grimsley, C., Lu, M., Walk, S.F., Tosello-Trampont, A.C., Macara, I.G., Madhani, H., Fink, G.R., and Ravichandran, K.S. (2002). Unconventional Rac-GEF activity is mediated through the Dock180-ELMO complex. *Nat Cell Biol* 4, 574-582.
43. Liu, Z.X., Yu, C.F., Nickel, C., Thomas, S., and Cantley, L.G. (2002). Hepatocyte growth factor induces ERK-dependent paxillin phosphorylation and regulates paxillin-focal adhesion kinase association. *J Biol Chem* 277, 10452-10458.
44. Turner, C.E., Brown, M.C., Perrotta, J.A., Riedy, M.C., Nikolopoulos, S.N., McDonald, A.R., Bagrodia, S., Thomas, S., and Leventhal, P.S. (1999). Paxillin LD4 motif binds PAK and PIX through a novel 95-kD ankyrin repeat, ARF-GAP protein: A role in cytoskeletal remodeling. *J Cell Biol* 145, 851-863.
45. West, K.A., Zhang, H., Brown, M.C., Nikolopoulos, S.N., Riedy, M.C., Horwitz, A.F., and Turner, C.E. (2001). The LD4 motif of paxillin regulates cell spreading and motility through an interaction with paxillin kinase linker (PKL). *J Cell Biol* 154, 161-176.
46. Zhao, Z.S., Manser, E., Loo, T.H., and Lim, L. (2000). Coupling of PAK-interacting exchange factor PIX to GIT1 promotes focal complex disassembly. *Mol Cell Biol* 20, 6354-6363.
47. Matsuya, M., Sasaki, H., Aoto, H., Mitaka, T., Nagura, K., Ohba, T., Ishino, M., Takahashi, S., Suzuki, R., and Sasaki, T. (1998). Cell adhesion kinase beta forms a complex with a new member, Hic-5, of proteins localized at focal adhesions. *J Biol Chem* 273, 1003-1014.
48. Osada, M., Ohmori, T., Yatomi, Y., Satoh, K., Hosogaya, S., and Ozaki, Y. (2001). Involvement of Hic-5 in platelet activation: integrin alphaIIb beta3-dependent tyrosine phosphorylation and association with proline-rich tyrosine kinase 2. *Biochem J* 355, 691-697.
49. Wang, Y., and Gilmore, T.D. (2003). Zyxin and paxillin proteins: focal adhesion plaque LIM domain proteins go nuclear. *Biochim Biophys Acta* 1593, 115-120.
50. Woods, A.J., Roberts, M.S., Choudhary, J., Barry, S.T., Mazaki, Y., Sabe, H., Morley, S.J., Critchley, D.R., and Norman, J.C. (2002). Paxillin associates with poly(A)-binding protein 1 at the dense endoplasmic reticulum and the leading edge of migrating cells. *J Biol Chem* 277, 6428-6437.
51. Shibamura, M., Kim-Kaneyama, J.R., Ishino, K., Sakamoto, N., Hishiki, T., Yamaguchi, K., Mori, K., Mashimo, J.,

- and Nose, K. (2003). Hic-5 communicates between focal adhesions and the nucleus through oxidant-sensitive nuclear export signal. *Mol Biol Cell* 14, 1158-1171.
52. Shibamura, M., Mori, K., Kim-Kaneyama, J.R., and Nose, K. (2005). Involvement of FAK and PTP-PEST in the regulation of redox-sensitive nuclear-cytoplasmic shuttling of a LIM protein, Hic-5. *Antioxid Redox Signal* 7, 335-347.
53. Kasai, M., Guerrero-Santoro, J., Friedman, R., Leman, E.S., Getzenberg, R.H., and DeFranco, D.B. (2003). The Group 3 LIM domain protein paxillin potentiates androgen receptor transactivation in prostate cancer cell lines. *Cancer Res* 63, 4927-4935.
54. Yang, L., Guerrero, J., Hong, H., DeFranco, D.B., and Stallcup, M.R. (2000). Interaction of the tau2 transcriptional activation domain of glucocorticoid receptor with a novel steroid receptor coactivator, Hic-5, which localizes to both focal adhesions and the nuclear matrix. *Mol Biol Cell* 11, 2007-2018.
55. Chodankar, R., Wu, D.Y., Schiller, B.J., Yamamoto, K.R., and Stallcup, M.R. (2014). Hic-5 is a transcription coregulator that acts before and/or after glucocorticoid receptor genome occupancy in a gene-selective manner. *Proc Natl Acad Sci U S A* 111, 4007-4012.
56. Shibamura, M., Kim-Kaneyama, J.R., Sato, S., and Nose, K. (2004). A LIM protein, Hic-5, functions as a potential coactivator for Sp1. *J Cell Biochem* 91, 633-645.
57. Ghogomu, S.M., van Venrooy, S., Ritthaler, M., Wedlich, D., and Gradl, D. (2006). HIC-5 is a novel repressor of lymphoid enhancer factor/T-cell factor-driven transcription. *J Biol Chem* 281, 1755-1764.
58. Kim, K., Jr., Shibamura, M., and Nose, K. (2002). Transcriptional activation of the c-fos gene by a LIM protein, Hic-5. *Biochem Biophys Res Commun* 299, 360-365.
59. Nose, K. (2002). [Regulation of gene expression by active oxygen species]. *Yakugaku Zasshi* 122, 773-780.
60. Lei, X.F., Kim-Kaneyama, J.R., Arita-Okubo, S., Offermanns, S., Itabe, H., Miyazaki, T., and Miyazaki, A. (2014). Identification of Hic-5 as a novel scaffold for the MKK4/p54 JNK pathway in the development of abdominal aortic aneurysms. *J Am Heart Assoc* 3, e000747.
61. Usuki, F., Fujita, E., and Sasagawa, N. (2008). Methylmercury activates

- ASK1/JNK signaling pathways, leading to apoptosis due to both mitochondria- and endoplasmic reticulum (ER)-generated processes in myogenic cell lines. *Neurotoxicology* 29, 22-30.
62. Funahashi, Y., Wang, Z., O'Malley, K.J., Tyagi, P., DeFranco, D.B., Gingrich, J.R., Takahashi, R., Majima, T., Gotoh, M., and Yoshimura, N. (2015). Influence of *E. coli*-induced prostatic inflammation on expression of androgen-responsive genes and transforming growth factor beta 1 cascade genes in rats. *Prostate* 75, 381-389.
63. Li, X., Martinez-Ferrer, M., Botta, V., Uwamariya, C., Banerjee, J., and Bhowmick, N.A. (2011). Epithelial Hic-5/ARA55 expression contributes to prostate tumorigenesis and castrate responsiveness. *Oncogene* 30, 167-177.
64. Akiyama, N., Matsuo, Y., Sai, H., Noda, M., and Kizaka-Kondoh, S. (2000). Identification of a series of transforming growth factor beta-responsive genes by retrovirus-mediated gene trap screening. *Mol Cell Biol* 20, 3266-3273.
65. Wu, R.F., Xu, Y.C., Ma, Z., Nwariaku, F.E., Sarosi, G.A., Jr., and Terada, L.S. (2005). Subcellular targeting of oxidants during endothelial cell migration. *J Cell Biol* 171, 893-904.
66. Desai, L.P., Zhou, Y., Estrada, A.V., Ding, Q., Cheng, G., Collawn, J.F., and Thannickal, V.J. (2014). Negative regulation of NADPH oxidase 4 by hydrogen peroxide-inducible clone 5 (Hic-5) protein. *J Biol Chem* 289, 18270-18278.
67. Pignatelli, J., Tumbarello, D.A., Schmidt, R.P., and Turner, C.E. (2012). Hic-5 promotes invadopodia formation and invasion during TGF-beta-induced epithelial-mesenchymal transition. *J Cell Biol* 197, 421-437.
68. Wang, H., Song, K., Sponseller, T.L., and Danielpour, D. (2005). Novel function of androgen receptor-associated protein 55/Hic-5 as a negative regulator of Smad3 signaling. *J Biol Chem* 280, 5154-5162.
69. Wang, H., Song, K., Krebs, T.L., Yang, J., and Danielpour, D. (2008). Smad7 is inactivated through a direct physical interaction with the LIM protein Hic-5/ARA55. *Oncogene* 27, 6791-6805.
70. Shola, D.T., Wang, H., Wahdan-Alaswad, R., and Danielpour, D. (2012). Hic-5 controls BMP4 responses in prostate cancer cells through interacting with Smads 1, 5 and 8. *Oncogene* 31, 2480-2490.
71. Kim-Kaneyama, J.R., Suzuki, W., Ichikawa, K., Ohki, T., Kohno, Y., Sata,

- M., Nose, K., and Shibnuma, M. (2005). Uni-axial stretching regulates intracellular localization of Hic-5 expressed in smooth-muscle cells in vivo. *J Cell Sci* 118, 937-949.
72. Nishiya, N., Tachibana, K., Shibnuma, M., Mashimo, J.I., and Nose, K. (2001). Hic-5-reduced cell spreading on fibronectin: competitive effects between paxillin and Hic-5 through interaction with focal adhesion kinase. *Mol Cell Biol* 21, 5332-5345.
73. Mitra, S.K., and Schlaepfer, D.D. (2006). Integrin-regulated FAK-Src signaling in normal and cancer cells. *Curr Opin Cell Biol* 18, 516-523.
74. Ninio-Many, L., Grossman, H., Shomron, N., Chuderland, D., and Shalgi, R. (2013). microRNA-125a-3p reduces cell proliferation and migration by targeting Fyn. *J Cell Sci* 126, 2867-2876.
75. Wade, R., Brimer, N., Lyons, C., and Vande Pol, S. (2011). Paxillin enables attachment-independent tyrosine phosphorylation of focal adhesion kinase and transformation by RAS. *J Biol Chem* 286, 37932-37944.
76. Mori, K., Hirao, E., Toya, Y., Oshima, Y., Ishikawa, F., Nose, K., and Shibnuma, M. (2009). Competitive nuclear export of cyclin D1 and Hic-5 regulates anchorage dependence of cell growth and survival. *Mol Biol Cell* 20, 218-232.
77. Noguchi, F., Inui, S., Nakajima, T., and Itami, S. (2012). Hic-5 affects proliferation, migration and invasion of B16 murine melanoma cells. *Pigment Cell Melanoma Res* 25, 773-782.
78. Deakin, N.O., and Turner, C.E. (2011). Distinct roles for paxillin and Hic-5 in regulating breast cancer cell morphology, invasion, and metastasis. *Mol Biol Cell* 22, 327-341.
79. Aissaoui, H., Prevost, C., Boucharaba, A., Sanhadji, K., Bordet, J.C., Negrier, C., and Boukerche, H. (2014). MDA-9/Syntenin is essential for Factor VIIa-induced Signaling, Migration, and metastasis in melanoma Cells. *J Biol Chem*.
80. Kratimenos, P., Koutroulis, I., Marconi, D., Syriopoulou, V., Delivoria-Papadopoulos, M., Chrousos, G.P., and Theocharis, S. (2014). Multi-targeted molecular therapeutic approach in aggressive neuroblastoma: the effect of Focal Adhesion Kinase-Src-Paxillin system. *Expert Opin Ther Targets* 18, 1395-1406.
81. Avraamides, C., Bromberg, M.E., Gaughan, J.P., Thomas, S.M., Tsygankov, A.Y., and Panetti, T.S. (2007). Hic-5 promotes endothelial cell

- migration to lysophosphatidic acid. *Am J Physiol Heart Circ Physiol* 293, H193-203.
82. Nakamura, K., Yano, H., Uchida, H., Hashimoto, S., Schaefer, E., and Sabe, H. (2000). Tyrosine phosphorylation of paxillin alpha is involved in temporospatial regulation of paxillin-containing focal adhesion formation and F-actin organization in motile cells. *J Biol Chem* 275, 27155-27164.
83. Deakin, N.O., Ballestrem, C., and Turner, C.E. (2012). Paxillin and Hic-5 interaction with vinculin is differentially regulated by Rac1 and RhoA. *PLoS One* 7, e37990.
84. Liu, R.F., Xu, X., Huang, J., Fei, Q.L., Chen, F., Li, Y.D., and Han, Z.G. (2013). Down-regulation of miR-517a and miR-517c promotes proliferation of hepatocellular carcinoma cells via targeting Pyk2. *Cancer Lett* 329, 164-173.
85. Geng, W., Ng, K.T., Sun, C.K., Yau, W.L., Liu, X.B., Cheng, Q., Poon, R.T., Lo, C.M., Man, K., and Fan, S.T. (2011). The role of proline rich tyrosine kinase 2 (Pyk2) on cisplatin resistance in hepatocellular carcinoma. *PLoS One* 6, e27362.
86. Li, Q., Liu, G., Shao, D., Wang, J., Yuan, H., Chen, T., Zhai, R., Ni, W., and Tai, G. (2015). Mucin1 mediates autocrine transforming growth factor beta signaling through activating the c-Jun N-terminal kinase/activator protein 1 pathway in human hepatocellular carcinoma cells. *Int J Biochem Cell Biol* 59, 116-125.
87. Dhanasekaran, R., Nakamura, I., Hu, C., Chen, G., Oseini, A.M., Seven, E.S., Miamen, A.G., Moser, C.D., Zhou, W., vanKuppevelt, T.H., vanDeursen, J., Mounajjed, T., Fernandez-Zapico, M.E., and Roberts, L.R. (2014). Activation of the TGFbeta/SMAD transcriptional pathway underlies a novel tumor promoting role of sulfatase1 in hepatocellular carcinoma. *Hepatology*.
88. Reichl, P., Dengler, M., van Zijl, F., Huber, H., Fuhrlinger, G., Reichel, C., Sieghart, W., Peck-Radosavljevic, M., Grubinger, M., and Mikulits, W. (2014). Signaling of Axl via 14-3-3zeta activates autocrine transforming growth factor-beta signaling in hepatocellular carcinoma. *Hepatology*.
89. Wu J.R., Hu C.T., You R.I., Pan S.M., Cheng C.C., Lee M.C., Wu C.C., Chang Y.J., Lin S.C., Chen C.S., Lin T.Y., Wu W.S. (2015) Hydrogen peroxide inducible clone-5 mediates reactive oxygen species signaling for hepatocellular carcinoma progression. *Oncotarget* 6, 32526-32544.

90. Ren T., Zhang H., Wang J., Zhu J., Jin M., Wu Y., Guo X., Ji L., Huang Q, Zhang H., Yang H., Xing J. (2017) MCU-dependent mitochondrial Ca²⁺inhibits NAD/SIRT3/SOD2 pathway to promote ROS production and metastasis of HCC cells. *Oncogene* 36, 5897-5909.
91. Hu C.T., Wu J.R., Cheng C.C., Wu W.S. (2017) The Therapeutic Targeting of HGF/c-Met Signaling in Hepatocellular Carcinoma: Alternative Approaches. *Cancers (Basel)*, 9(6) pii: E58.