

REVIEW ARTICLE

Continuous Monitoring and Modeling Contractility of Skeletal Muscles in Motion

Authors

Xi WANG

Xiaoming Tao*

Affiliation:

Institute of Textiles and Clothing, The Hong Kong Polytechnic University, Hong Kong, China

Correspondence:

Xiaoming Tao, Institute of Textiles and Clothing, The Hong Kong Polytechnic University, Hong Kong, China. Email: xiao-ming.tao@polyu.edu.hk

Abstract

Continuous monitoring and analysis of skeletal muscles' contractility have been extensively associated with sensing and bio-signal processing technologies and increasingly demanded by applications in the fields of sports, control and interaction, rehabilitation and medical care. While most existing approaches are confined in isometric studies in clinics or laboratories, researchers have been devoted in recent years towards continuous monitoring and analysis of skeletal muscles' contractility in motion. This paper aims to provide an overview of current status of non-invasive sensing technologies for monitoring skeletal muscles' activation, up-to-date findings on observing and characterizing the force-length and force-velocity relationships, and various existing activation-contractility models. In addition, this paper evaluates various sensing technologies for muscle activation, indicates challenges for bio-mechanical modeling on activation-contractility, and makes recommendations on future developments in continuous monitoring and analysis of skeletal muscles' contractility in-motion.

1 Introduction

As the motors of the human motions and stabilizers of joint positions, skeletal muscles have attracted great interests of researchers on their contractile properties and characteristics. The bio-mechanical analysis of the contractility has been constantly required and extensively incorporated in the areas of rehabilitation and medical care, control and interactions, as well

as sports training ^[1-5]. However, direct measurements of muscle contractility are impossible, and inverse dynamics analysis only provides a net output of moment by all surrounding skeletal muscles around a joint. In order to determine the tension of individual muscles, using a variety of sensing technologies to link the muscle activation to contractility based on bio-mechanical models (Figure 1) remains the only feasible choice.

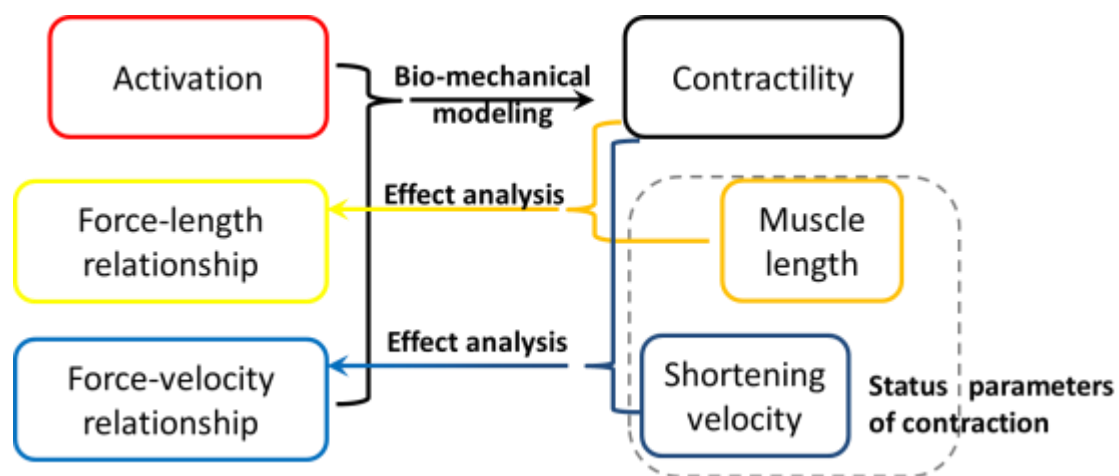


Figure 1. Typical Hill-type modeling process for activation-contraction relationship

Hence, this review delivers a conflation of up-to-date researches on monitoring the activation of skeletal muscles as well as the determination of force-length and force-velocity relationships, both are essential for the bio-mechanical modeling of activation-to-contraction, and presents the merits and challenges for in-motion monitoring of muscle contractility. Since studies of contraction in impaired muscles, such as neuromuscular diseases and fatiguing contraction, were reviewed somewhere else ^[6-13]. In this paper we mainly focus on non-fatigue muscle contractions of healthy subjects.

2 Monitoring Muscle Activation

2.1 Classical approaches

Skeletal muscles' contraction is generally accompanied by three typical kinds of bio-physical behavior, namely the polarization-depolarization of sarcolemma, vibration of tensioned muscle fiber, shortening in length while expansion in cross-sectional area of the muscle fiber. Accordingly three types of non-invasive approaches for monitoring muscle contraction include electrical (surface electromyography), mechanical (mechanomyogram), and morphological

(computed tomography, magnetic resonance imaging, ultrasound, etc.)^[1, 14-16]. Over the past decades, efforts have been made to track muscle contraction continuously by correlating bioelectricity of superficial skeletal muscles^[3, 17-21], low-frequency oscillation of firing muscle fibers^[21-23], as well as changing in specified morphology parameters of muscles during contraction^[24-28], with muscular contraction status (mostly indexed by contractile force). One thing should be noted is that, among all these attempts, the majority of quantitative researches are merely limited to isometric contractions. According to Hill's theory of muscle contraction, contractility is affected by activation of skeletal muscle and the status of contraction, apart from the maximum voluntary contraction of skeletal muscle(s). Isometric contractions are convenient because with fixed muscle length or joint position, the so-called 'monitoring of muscle contraction' is in essence the monitoring of indexes of muscle activation, which have been realized by the aforementioned three types of sensing technologies. However, as a successful index of activation for motion, the following aspects of each technology need to be considered: (1) the nature of the activation–force relationship, (2) stability of acquisition for all contraction modes. Further for activation-contraction modeling, elaborations of the influence factors on dynamic muscle contractions shall also be observed.

2.1.1 Surface electromyography (sEMG)

sEMG provides information on individual muscle's bio-physical activity, which has been correlated with contractile force. sEMG collects the action potential through electrodes attached to the skin surface at the middle of the muscle belly^[29]. As a composition of action potentials

of multiple motor units, sEMG signal has been widely applied for both clinical and research purposes^[30]. In particular, sEMG has been extensively used to study the muscular functions and muscles' coordination. The research scope of this area mainly consists of the following four aspects, where the 1st, 3rd and 4th aspects are related to the contractility of skeletal muscles:

1. isometric contraction, maximal voluntary contraction (MVC) and the relationship between sEMG and contractile force^[10, 21, 31-34];
2. healthy skeletal muscles' anatomical function during assigned movements or sports, jointly with other synchronized bio-signals^[11, 35-38];
3. muscle fatigue^[39];
4. computational and clinical studies of sEMG on assisting occupational rehabilitation, including sEMG-to-motion modeling^[3, 35, 40-43].

In an oscillation wave-shape, the sEMG signals can be analyzed and interpreted in frequency domain and time domain, the absolute amplitudes ('envelop' or the outline) of the later is related to the intensity of contraction. For a simple comparison analysis, the magnitudes of sEMG representing the gross innervation, with which the skeletal muscle works, avoiding the influences of non-reproducible components of sEMG at different time-intervals^[44, 45]. For continuous monitoring purpose, it is preferable to normalize the in-motion amplitude with Maximum Voluntary Contraction (MVC) to index the level of muscle activation^[3, 35, 40]. High correlation between the

normalized sEMG and force generation has been reported constantly in both linear and nonlinear patterns. A typical relation is monotonic and curvilinear, a higher sEMG is needed for a further unit increase in contractile force^[21, 46, 47]. For static tests, when both sEMG and force are normalized to their respective maximum values, some muscles tend to show an apparent linear EMG-force relationship^[48, 49]. The investigation of such relationships is important since sEMG has been used for assisting EMG-to-motion applications based on biomechanical models^[40, 42, 50]. However, there are two pertinent problems, especially for in-motion application. The first is that sEMG signals are sensitive to the size, number and firing rate of muscle units, which undermines the role of sEMG as representative index for the intensity of selected muscles' contraction, not to mention that sEMG signals are strongly influenced by conditions of detection, such as the skin humidity, contact resistances, both are difficult to maintain constant in fierce motions. The dependence on the instrumentation and acquisition procedures may have influences, which have been extensively discussed elsewhere^[51-53]. Secondly, a recent work shows that the sEMG normalization is limited and inaccurate especially for high-velocity dynamic tasks, which implies that the optimal normalization methods should be muscle and task-dependent^[46]. Because concentric contractions shorten the muscle length, the location between muscle and sEMG electrodes may be changed. Moreover, the status of muscle contraction, such as length and shortening velocity, also affects the sEMG signal^[54, 55]. Up to now, the quantitative studies have been limited on isometric contractions for limited time duration. Hence, although sEMG is now the most common noninvasive approach

to monitor skeletal muscles' activation, it still confronts several challenges as an index of muscle activation in motion.

2.1.2 Mechanomyography (MMG)

MMG signals represent the vibrations of active muscle fibers during contraction, which can be detected by piezoelectric sensors^[9, 56], microphones^[57, 58], accelerometers^[47, 59, 60] and laser distance sensors^[61, 62]. The oscillations, reported as prominently influenced by the global firing rate of motor units^[63-65], reflect the mechanical counterpart of the electrical activity of the motor unit as measured by sEMG^[19]. Compared with sEMG, MMG signals cover a wider physiological range of motor units, even the underlying muscles, only that the waves oscillate as discrete bursts rather than continuous tones^[66]. Furthermore, the placement of MMG sensors is not required to be precise or specific^[67], and MMG signals are not influenced by changes in the skin impedance and sweating^[68].

MMG studies on muscle activities are numerous, including characterization of neuromuscular disorders^[6, 69, 70], development of prosthesis and/or switch control^[64, 65, 71, 72], activity of motor units^[73-76], examination of mechanical properties during exercises^[77, 78], and rehabilitation systems^[79]. Temporal and spectral components of MMG signals with difference levels of contraction have been employed to determine muscle strength and stiffness. Studies of MMG versus isometric torque of human elbow indicate that the relationship between MMG amplitude and isometric torque is linear for lower-strength subjects and cubic for higher-strength subjects^[80-82]. It was reported that the RMS of MMG decreases at high levels of force due to

mechanical fusion of MU activity while the mean power frequency of MMG increases^[83, 84].

As an alternative promising monitoring approach for muscle contraction, however, MMG has not been developed fully as it cannot determine the activation level of muscle to reflect contractility. Existing studies on MMG are confined to a small sample size of healthy population^[64, 85]. Recently, A recent study combining sEMG and MMG has facilitated new understanding of the electromechanical coupling of skeletal muscles^[21, 22, 75, 86]. Noise contamination is also a major barrier for MMG. Low-frequency (5-100 Hz) MMG is not perfect for in-motion monitoring of muscle contractions, due to the fact that the low-frequency components of MMG are easily mixed with human movements. On the other hand, the high-frequency components of MMG are contaminated by nearby vibrating muscle fibers or environmental noises^[68].

2.1.3 Tomographic imaging methods

Skeletal muscle's architecture alters with contraction^[87], as muscle fibers shorten and simultaneously change the morphology of the entire skeletal muscle. Imaging methods such as the ultrasound (ultrasonography), computerized tomography (CT), and magnetic resonance imaging (MRI) have all been implemented to detect morphological deformation of skeletal muscles in real-time. Through on-line or post imaging processing, architectural parameters of skeletal muscles can be quantitatively identified, such as the changes in cross-section area^[24-26, 88] and muscle volume^[27, 28], muscle thickness and fiber pennation^[20, 21, 89], through image processing of cross-sectional area of skeletal muscles. Those architectural parameters have been reported as

index of contractile force during isometric contraction, and correlations between the morphological parameters and generated joint torque were achieved^[15, 20, 21, 90]. Being low-cost, non-ionizing, stable, and available for deep muscles, ultrasonography has been widely applied in researches covering:

1. Correlations among change of CSAs (cross-sectional areas), expansion of muscle size and contractile force have been reported frequently^[28, 29, 88, 91-98];
2. Shortening fascicle length and increasing pennation angle were also reported to be highly correlated to skeletal muscle's contraction^[99-103];
3. Muscular movement or displacement was detected with in-vivo ultrasound^[104, 105];
4. Potential of muscle fatigue evaluation and prosthetic control using the ultrasound has been discussed^[106], providing more comprehensive information than using sEMG only.
5. Muscle thickness extracted from CSA^[107] have been jointly studied and found correlated with other indexes of muscle contraction^[21, 91, 101, 108-110]. Corresponding image tracking algorithms have been developed for in-motion detecting. For relaxed skeletal muscle, muscle thickness was also observed in close relationship with joint angle due to the shortening of muscle length.

The 1st, 2nd, and 5th items are related to monitoring of muscle contraction. In particular,

for the biceps brachii (major components of elbow flexors), Hodges ^[111] found that the muscle thickness has a negative exponential relationship with sEMG signal, i.e., the muscle thickness increases with sEMG almost linearly in low-contraction (<30% MVC) while much slower in high-contraction condition. This observation was confirmed by Akagi ^[112], Abe ^[113], and Zheng's group ^[20, 21], who developed a system to record and analyze ultrasound images, force, joint angle and sEMG simultaneously. However, due to the difficult fixation of ultrasound sensor in dynamic conditions, almost all the studies were restrained in static (isometric) and quasi-static conditions.

Attempts have been made to establish the relationship between the measured muscle size (eg, thickness and/or cross-sectional area) and the level and timing of muscle activation ^[114]. The level of muscle activation was determined by comparing the size of a contracted muscle to its size during rest. Using measured muscle size from static ultrasound images as an indication

of muscle activation, however, has limitations. The level of muscle activation depends not just on a muscle's size, but on initial muscle (fascicle) length, amount of tendon stretch, type of contraction (isometric, concentric, or eccentric), muscle fiber pennation angle, and forces from surrounding tissues ^{[115] [116, 117]}. Up to date, there have not been sufficient studies on examining the reliability and validity of ultrasound for quantifying muscle activation during research and clinical practice ^[118].

2.2 New technologies for monitoring muscle contraction

New sensors and sensing technologies have illustrated great promises for monitoring and analyzing skeletal muscles' contraction. From recent papers published between 2014 and 2018, as shown in Table 1 and Figure 2, tensiomyography, optic sensors, novel ultrasound sensors, piezoelectric sensors and large-deformation strain sensors have been reported.

Table 1 Summary of new sensing technologies for monitoring muscle contractions

References	Type of sensor used	Principles	Remarks
[119-123]	Tensiomyography	Radial muscle belly displacement under electrical stimulus is analyzed for neuro-muscular function of muscle	For evaluating contraction only; Superficial muscle only; Isometric mode only;
[124-128]	LED & photo detector	Light absorption and reflection by muscle fibers vary during contraction; Tissue oxygenation decreases in contraction	Qualitative monitoring of muscle contraction

[129, 130]	Wearable ultrasound piezoelectric PVDF sensor	Muscle thickness increases during contraction	Qualitative monitoring of muscle contraction; Isometric mode only;
[13, 131]	Piezoelectric sensor	Modulus of muscle increases in tension	For analysis and evaluation of contraction only; Lack of dynamic study;
[132-135]	Large-deformation strain sensor	Expansion of CSA of muscle fibers during contraction	Model for activation-contraction; Involving both isometric and kinetic modes.

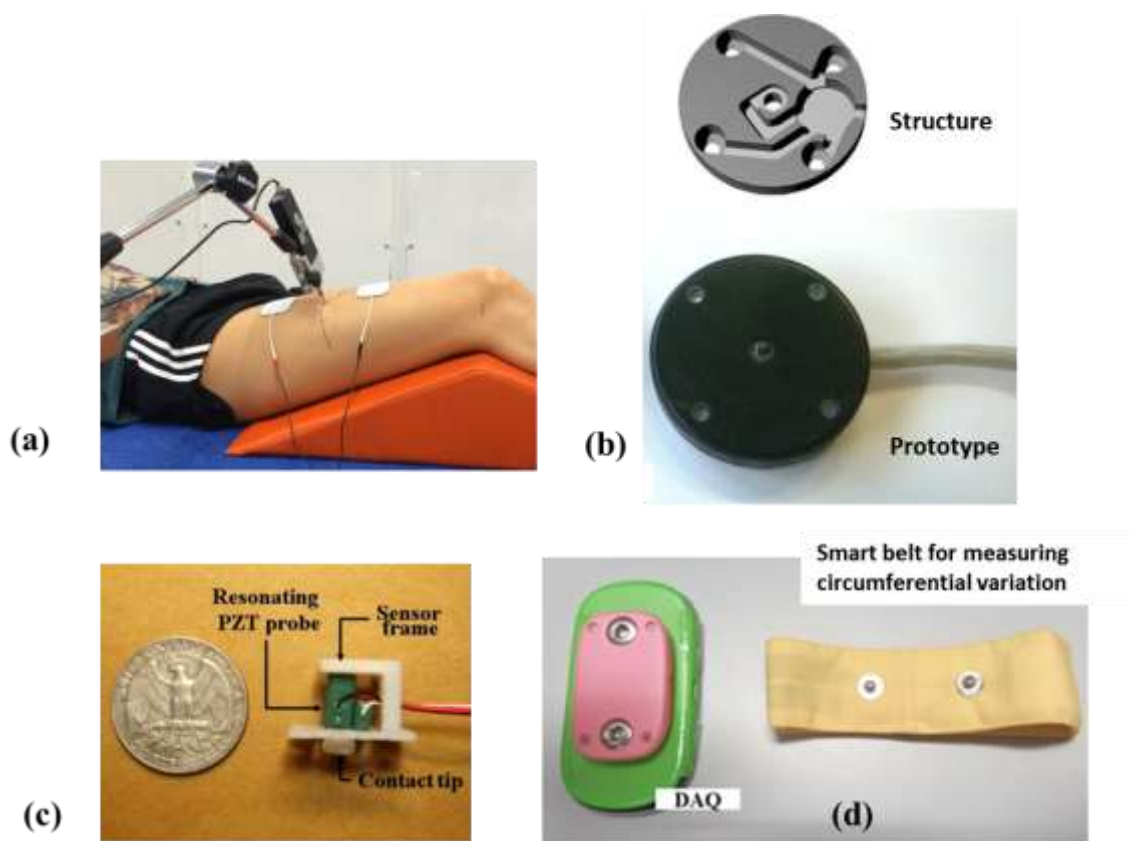


Figure 2. (a) Tensiomyography sensor ^[121]; (b) electro-optical muscle sensor ^[125]; (c) piezoelectric sensor ^[131] and (d) upgraded soft strain sensor ^[134]

Tensiomyography

Tensiomyography (TMG, sometimes the MC sensors) is a portable non-invasive method to assess in vivo passive muscle contractile properties^[136]. Inspired by MMG, tensiomyography uses a high-precision digital transducer placed on muscle surface to capture waveforms integrating parameters such as maximum radial displacement of the muscle belly and contraction time^[137, 138]. Compared to MMG techniques, TMG signals are not affected by slight muscle pretension, and thus have a higher signal-to-noise ratio.

TMG has been used for evaluation of muscular fatigue^[12], impairment^[139], as well as muscular changes/adaptations^[140]. Variations of TMG-derived parameters show significant correlation with changes in MVC^[123]. TMG has been appreciated by strength and conditioning coaches, physiotherapists, and sport scientists, who preferentially seek accurate and practical assessment methods which do not disturb their professional routines^[141, 142]. However, TMG has a number of shortcomings. First, studies of TMG have been confined in isometric mode, no dynamic application is yet possible. Secondly, TMG is not able to assess deep muscles. Finally, congestion due to training may affect the accuracy of TMG data. In summary, due to the lack of in-motion monitoring ability and stability, TMG can serve as an evaluation tool of contractile properties such as muscle fatigue and capability, other than monitoring muscle contraction.

Optical sensors

Optical sensing devices for muscle contraction consist of light emitting diode (LED) and photo detector (photodiodes)^[124-126]. The working principle is to measure the change in intensity of back scattered light from skeletal muscle

tissue, which is caused by myosin proteins' crystalline properties during contraction. Recent work^[124] shows that photodiodes can also derive the tissue oxyhemoglobin absorption from the measured light intensity, i.e., the steady decrease in the tissue oxygenation during ischemia. However, studies on optical sensors are still in initial stage, especially as a tool for assessing muscle contractility.

New wearable piezoelectric ultrasound sensors

Inspired by conventional ultrasound methods, simple and wearable sensing devices have been designed for monitoring muscle contraction by quantifying muscle thickness and active muscle stiffness, respectively. Examples include disk-shaped ultrasound device^[109] and piezoelectric sensor-based reasoning device^[103]. The latter is more of an evaluation tool of contractile properties while the wearable ultrasound sensor needs to prove its stability of signal in dynamic conditions.

Anthropometric measurement devices

Involving measurements of various dimensional descriptors of human body, anthropometry has been widely used in industrial and clothing design, ergonomics and human fitness evaluation^[143]. Muscle size from anthropometric measurement (e.g. limb circumferences) has been found to correspond to the muscularity^[108, 144-149], i.e., higher muscular strength is associated with greater limb circumference and vice versa. Since the thickness, identified from the muscle cross-sectional area, has been frequently reported as architectural index for skeletal muscle's contraction^[21, 91, 101, 108-110], the change in limb circumference induced by expansion of cross sectional area may be another index.

Correlations between the limb circumference and torque at MVC have been reported, moreover, most of which were obtained at static or quasi-static conditions ^[132, 145, 150, 151].

The anthropometric measurement renders a number of wearable sensing devices that facilitate long-term continuous monitoring in dynamic conditions. A mechanical armband with steel wires was demonstrated ^[152] for measuring the circumference of human forearm in-motion, and found an apparent linear relationship between forearm circumferences and grip force, which is in agreement with the strength-size research findings ^[153, 154]. By using a muscle circumference sensor (with metal wires), Kim^[132] proposed a preliminary Hill-based model for human upper arm, elbow torque was predicted from the measured mid-upper-arm circumference with significant estimation error. There was no in-depth studies published supporting the arbitrary replacement of sEMG with measured mid-upper-arm circumference as a new 'activation level'. Other factors should be further studied, such as the joint position and speed of flexion. Nevertheless, these works have inspired developments of wearable monitoring systems for deformation of muscle during contraction. However, the measurement devices were rigid, interfering with muscle activity. A new type of strain sensors has been commercially available for large repeated deformation up to 60%, high sensitivity and good accuracy ^[14, 155-160]. They were made from elastic fabrics coated with elastomer/carbon nano-particles composite. Wang ^[133] used a measurement device with these fabric strain sensors and studied the relationship between upper arm circumferential strain and elbow flexion, in isometric, isokinetic and isotonic flexions. He has obtained empirical relationships between the

circumferential strain and contraction torque ^[133], in addition, a biomechanical model for kinetic flexions was proposed based on the observations of force-length and force-velocity relationship ^[134]. The model was validated for isokinetic contractions for moderate speeds. As the derivation of force-velocity relationship was based on slow or median speed, lack of experimental evidence on contraction at high-shortening speeds (>10 m/s or 450°/s), there is still a question unanswered, that is, how can the circumferential strain be linked to the muscle activation, especially in high-speed dynamic conditions.

It is noted that these anthropometric studies have focused on the activation based on measured contraction-induced circumference strains and the activation-contraction model even in very early stage ^[132, 134].

In general, studies of muscle activation using conventional technologies (EMG, MMG and tomographic imaging) are still restricted in static/isometric contractions instead of in-motion, due to aforementioned drawbacks impairing the signal stability in dynamic conditions. The new technologies have not been tested yet for long thus no sufficient cases have been reported. TMG has not been used for in-vivo monitoring of contraction. The optical sensors are immune to electric and electronic disturbance but affiliated bulky and heavy modem and wires prevent them from in-field applications. The piezoelectric ultrasound sensors need to prove their signal stability in dynamic conditions. The anthropometric measurement devices is convenient for sports, extracting the strains of individual muscles could be a challenge, however. Meanwhile, how the circumferential strain

indexes muscle activation in kinetic contractions should be further studied.

3 Biomechanical Models of Activation-Contractility

Although over 80 years have passed since Hill^[161] first revealed his biomechanical insights into muscle contraction, quantitative modeling of contractility has been mainly limited by phenomenological implementation of various Hill-based models. Huxley^[162] further introduced the dynamics of cross-bridge cycling into the contractile element (instead of a black box in Hill's) and successfully reproduced fast-twitch muscles, which could not be explained by a classical Hill model^[163, 164]. However, Huxley's consideration was claimed too computation-time-consuming for use in musculoskeletal modeling^[165, 166], due to the complex mathematical formulation. Meanwhile, recent works show that muscle tensions predicted by both Hill and Huxley models are within the same range^[165, 167]. Hence, though Huxley's model provides more realistic patterns of muscle contraction, it is economical and reasonable to use a simpler numeric implementation based on Hill type models.

Apart from the activation talked above, the influences of status parameters on the contractility shall be determined, i.e., the force-length and force-velocity relationships. Moreover, the acquisition of the status parameters is addressed, by reviewing sensing technologies on joint angle measurement. All above are essential to complete a Hill-type biomechanical modeling of activation-contractility in motion.

3.1 Hill-type biomechanical models

Once the muscle activation is determined experimentally, the next step shall be to build an activation-to-contraction conversion linkage, commonly known as the biomechanical modeling. As first released in Hill's macroscopic studies of skeletal muscles' contraction^[168, 169], the muscle tension has a hyperbolic relationship with shortening velocity^[161, 170, 171]. This finding was then extended and expressed by Zajac^[172] and Winters^[173, 174] in a four-element one-dimensional Hill-type model:

$$F_{CE} = \alpha(t) \cdot f(v) \cdot [f(l) \cdot F_{Max}]$$

The contraction force of muscle fiber, F_{CE} , is a function of MVC at current muscle length, scale factor for the activation level, $\alpha(t)$, and normalized factor of shortening velocity, $f(v)$ ^[172, 175-177]. The functions $\alpha(t)$, $f(v)$ and $f(l)$ can be in diverse forms with particular parameters, to be specific to different skeletal muscles^[174, 175, 178, 179].

These models have been used extensively for assessing skeletal muscle characteristics^[43, 49, 178, 180], contractions and movements^[41, 42, 175, 181, 182], analyzing neuromuscular-related diseases and rehabilitation^[183-190]. In particular, with sEMG derived activation levels, Hill-type models have been proven effective for emulating muscular behavior^[35, 191]. However, the accuracy and reliability of in-vivo muscle forces predicted by these models remains unknown, due to the lack of suitable implanted transducers.

Microscopic muscle fiber models are established with bio-physical tuning parameters that predict muscle characteristics and

contractions quite well ^[192-196]. Though relying on intrinsic properties likewise, namely the construction of activation from bio-signals, the force-length and force-velocity relationships, the macroscopic muscle models, however, are commonly based on experiments and phenomenology ^[197-200]. The adoption of microscopic sarcomere model to whole muscle or straight application of existing macroscopic model for one kind of skeletal muscle to another, have been reported to induce huge errors in prediction of real contractility, especially for movements at the low or high ends of speed ^[201]. The reason lies in the fact that the real structure of skeletal muscles and recruitment patterns of slower and faster motor units in muscle make the contractile properties largely indescribable ^[173, 201-205]. This naturally requires task-dependent observations on the determination of parameters in the models ^[206-208]. Moreover, since muscles are normally surrounded by other tissues, the impedance caused by connective tissues and bones ^[209, 210] bring some difficulties in modeling.

In summary, to construct a Hill-type biomechanical model for in-motion monitoring of contractility, it is better to determine the core elements of a Hill type muscle model through isolated designed experiments with selected conditions. Hence, the next subsections will cover the research status on determining the two core elements, that is, the force-length and force-velocity relationships.

3.2 Force-length relationship

In the human musculoskeletal system, the tension and status of contraction (i.e., the length and shortening speed) of skeletal muscles are alternatively represented by corresponding joint torque, joint position (or

joint angle) and joint angular velocity, respectively ^[211, 212].

Experimental studies have been conducted on sarcomere, fibers and whole muscles from various animals, mostly cat, frog, rabbit and rat, and in isometric contraction mode ^[213, 214]. In the isometric mode, the contraction force of muscle was length dependent while velocity was zero and maximum contraction incurred, the same did maximum activation. For muscular-skeletal systems, the force-length relationship is replaced by torque-angle relationship, which differs from the biological force-length relationship since it incorporates the effect of moment arm of skeletal muscles to the joint. There is definitely a difference in optimum muscle length for concentric and eccentric contractions, respectively, as reported by Melo et.al ^[215] in knee flexion and extension studies. Moreover, the force-length relationship was found to be activation-dependent ^[216], which is in consistence with Hill's theory.

The first qualitative descriptions of force-length relationship were associated with theoretical considerations on the interaction of actin and myosin, known as sliding filaments theory ^[192, 217]. Since then, a variety of force-length relationship has been proposed. The force-length relationship for the whole muscle is the easiest to obtained from experiments and thus discussed frequently ^[115, 173, 209]. However, most derived relationships were empirical, based on best data fitting without underlining biomechanical or physiological analysis. A theoretical force-length relationship is more difficult because it should incorporate a complex combination of properties of sarcomere, tendon, and muscle unit, as well as architectural particularities and history of contraction ^[213, 218-220], not to mention the

controversy over variation of muscle length after strength training. Until recently, only one purely physiological model was presented [221], where the force-length relationship was parameterized by using the geometry of internal muscle structures.

3.3 Force-velocity relationship

The force-velocity relationship was generally obtained from various activities at maximum activation, such as cycling [222-224], vertical jumps [225-229], treadmill training [230, 231], leg press [232-234], arm and upper body movements [223, 224, 227, 235]. The standardization and observation of force-velocity relationship are essential not just for routine tests but also for biomechanical modeling. One thing should be pointed out is that the ‘force’ used by these researchers is either an index of load/resistance or contractility deducted inversely from devices, while the ‘velocity’ is not exactly the shortening velocity of muscle but a speed of the motion, somehow linked to the skeletal muscles’ shortening velocity. For example, recent researches use mean force exerted onto the ground and the mean velocity of the mass center to establish the force-velocity profile in the vertical squat jump [236, 237]. In the human muscular-skeletal system, the joint moments and angular velocity appears to possess a hyperbolic relationship, similar to the original force-velocity relationship for single muscle fibers [238], referred as the joint torque-angular velocity properties. For individual movements, the force-velocity relationship can be determined in isokinetic training mode, by dynamometers, such as Biodex™.

While the force-velocity relationship of isolated muscles has been known to be hyperbolic [161], multi-joint functional tasks typically reveal

strong and approximately linear patterns [225, 239, 240]. A linear force-velocity relationship has been observed in the squat [241], leg press [240], free and loaded vertical jumps [225, 236], cycling [222, 230], treadmill running [230, 231], arm cranking [223], in both bench presses and bench press throws [235, 242], and during rowing [243]. Despite the experimental facts, the mechanisms of non-hyperbolic relationship in multi-joint movements are still not clear. As a consequence, no physiological model has been proposed. In the classical Hill-type biomechanical models, the force-velocity and force-length relationships are independent to each other, however, the interaction item between them has been reported [244], which needs investigations in future study.

According to Hill’s theory, the best condition to identify and determine the force-velocity relationship is to maintain the maximum activation level, for kinetic modes, ‘maximum’ means to move as rapidly as possible regardless of the restraints/resistance, which, however, cannot be fully satisfied by maximum movements/tasks in practice. Behm [245] claimed after studying ankle dorsiflexion isokinetically that the “intent to move quickly” is the only important factor for producing accurate velocity-strength relationship. Although numerous isometric studies have constantly shown correlations between the contraction force and activation indicators, activation itself actually represents the level of output power in Hill’s model. Recent studies have shown difference in the force-velocity relationships obtained by using different external loads [246, 247]. Furthermore, different shortening velocity also varies the observed force-length relationships [248, 249]. There is always a challenge that although it’s long believed that activation, force-length and force-

velocity properties are mutually independent, a crossed instead of separated consideration of activation, length and shortening velocity shall be done during Hill type modeling, where combined effect of the three mentioned factors on scaling the output contractility require s further research investigations.

3.4 In-motion angle measurement technologies

As elaborated above, the observation of the key status indicators of muscle contraction, i.e., the length of muscle fiber and the shortening velocity, rely on the detection of joint angles. A brief review of in-motion determination of joint angles base on various technologies is to be presented in this section. More detailed reviews can be found from reference [250-252].

Inertial measurement units (IMU)

For posture measurement, most conventional and common solution is using the inertial measurement units (IMU), which has been implemented either as a stand-alone sensing device or integrated in smart phones. The IMUs can measure angular velocity, acceleration and the magnetic field vector in their own 3D local coordinate system (Figure 3.a). Strap-down integration [250, 256] of angular velocity is used as a preliminary estimate of the displacement, the drifts of which are corrected based on a number of Kalman-filtering algorithms [257, 258]. The combination of multiple IMUs placed on body segments around a joint provides the joint angles [253, 259-261]. However, the measured acceleration and magnetic field vector are disturbed by impact on ligaments and presence of magnetic objects, lowering the accuracy of displacement or orientation. Furthermore, for accurate measurement of joint angles, a multi-IMU system is needed, which often undermines the freedom of movement in motions.

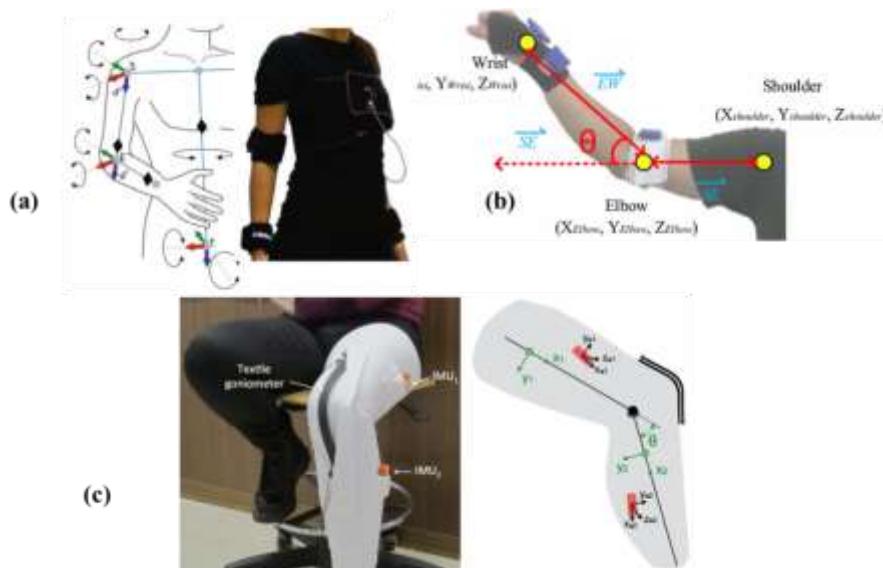


Figure 3. Reported technologies on joint angle measurement/posture recognition: a) IMUs^[253] b) optoelectronics sensors^[254] c) fabric-based (KPF)^[255]

Multi-camera motion capture system

The motion-capture system based on optoelectronic sensors has been used for visual assessment in interaction, physical therapy or rehabilitation [251, 252]. With or without markers on major limbs, optoelectronic sensors work with cameras and 3D post-processing vision system (either contrast based or depth based) to track joint angles through limb orientation and motion of body segments (Figure 3.b), only for major limbs, however. Marker-based vision-capture systems are accurate and reliable, conventionally referred as benchmark [254, 262, 263]. However, they are costly, need professional calibration and strict-conditioned circumstance. Environmental noises in captured images due to occlusion, self-occlusion, and unconventional body postures can induce wrong limb-identifications [264].

Soft goniometers

Recently, soft goniometers such as textile-based wearable sensors, have been working with or without the aforementioned other three types of sensors [265, 266]. Conductive elastomer coated fabrics [267, 268] and knitted piezoresistive fabric [255, 269-271] have been studied for movements/ postures recognition [268, 269, 271] and joint angle measurement [272], due to the merits of high compatibility with in-field activities (Figure 3.c). Correlations between knee angle and resistance change were observed and characteristics of gait cycle can be accurately identified, with a mean error of <3% [273, 274], comparable to that of commercial IMUs. For simpler on-and-off applications, thresholds have been set to evaluate the range of motion, for instance, whether the target range was achieved [275]. For upper-limbs

applications, gloves, sleeves and shirt have been developed based on those soft sensors. With an angle measurement error equal or less than 8%, those garments give reliable identification of static posture of hand, arm and shoulder [276]. However, these prototypes perform poorly in transient measurements, due to the drift in angle-resistance curves affected by stretching speed of the sensing area, as well as in the recovery time prior to the second use [277]. Up-to-date, effort has been reported in optimization of device design and arrangement of sensors, and in employment of IMUs in order to improve the accuracy of measurements [272]. Although some angle-sensing gloves and shirts have been demonstrated for providing feedback for people with central nervous system lesion in therapeutic exercises [7, 267, 269, 278, 279], further research is required to enhance their accuracy, reliability as the angle measurement, diagnosis and rehabilitation tools.

4 Applications

Up-to-date, sEMG has been the only widely used instrumental tool to construct activation of skeletal muscle contraction due to its biophysical nature. Generally, the sEMG-based activation-contraction models have continually been incorporated in studies of prosthetic/supporting robotics, many times merely based on EMG interpretation and pattern learning [280-283], thus will not be elaborated here. By studying how much muscle force is being produced or to be produced for rehabilitation and medical intervention evaluation purposes, therapists can set safe limits in their therapies, meanwhile patients can learn to adjust force production to fulfill designated actions. However, these studies are still confined in static condition/isometric mode of contractions (Figure 4.a). Manal [284]

successfully predicted the ankle moments in isometric plantar flexion and dorsiflexion with a tuned sEMG-driven Hill model. A system for quasi-dynamic monitoring of ankle moments and achilles tendon force was also preliminarily presented, consisting of electro-goniometers and EMG sensors (Figure 4.c). Shao [50] from the same group applied the EMG-driven Hill model for four stroke patients and predicted ankle moment during stance with an acceptable RMS error of between 9.7%~14.7%, conforming the model’s consistency and effectiveness as rehabilitation therapies to assess intervention. Apart from isometric

contractions, Koo [285] and Pau [286] tested their sEMG-driven Hill models in the other way, by comparing elbow joint trajectory predicted with externally measured during isotonic elbow flexions (Figure 4.b). It was claimed that the discrepancy between which was due to muscle activation constructed from sEMG signals in dynamic conditions. More work related to the above two perspectives was reviewed by Biewener [191], suggesting better sensing technologies to further improve the accuracy of Hill-based activation-contractility models among multiple tasks.

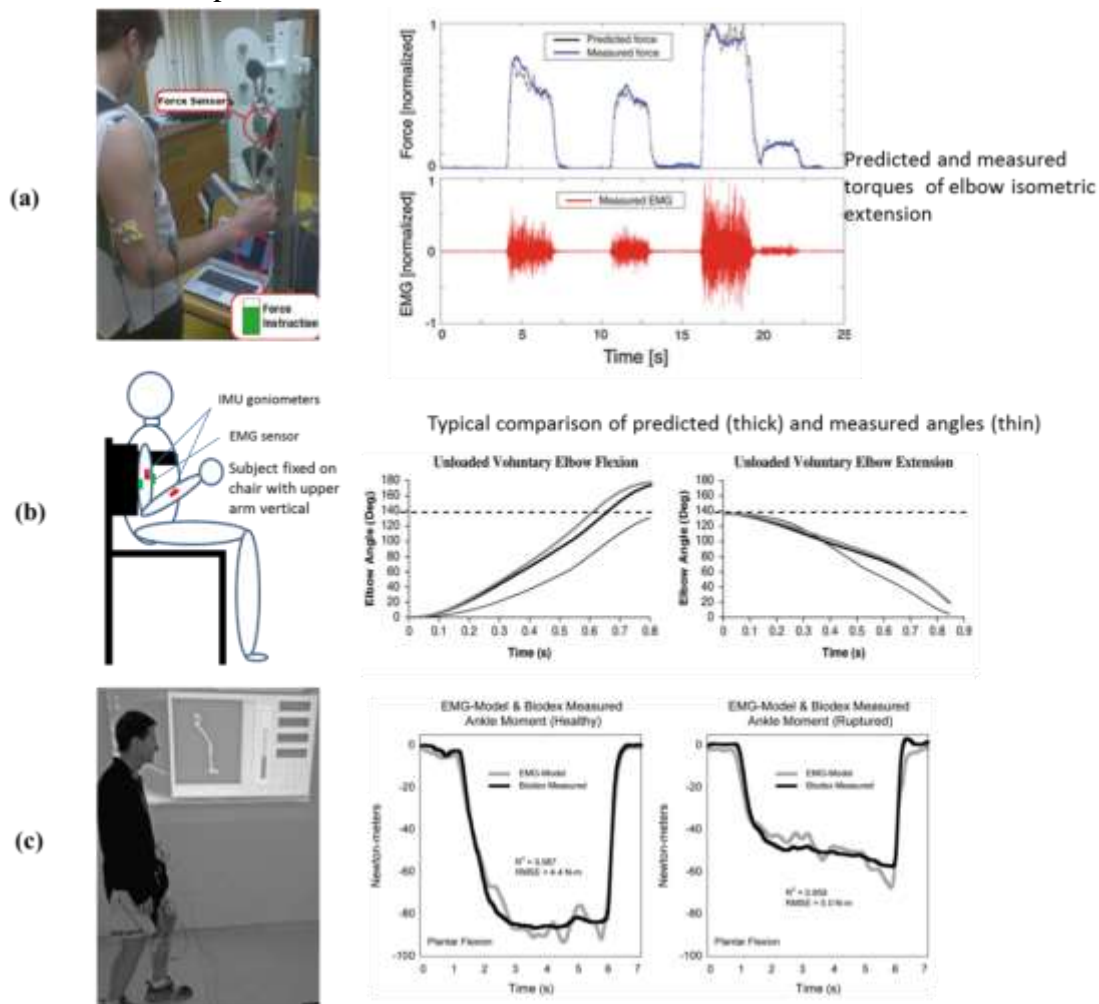


Figure 4. (a) Prediction of torque based on sEMG-force Hill model during isometric contraction^[287]; (b) Elbow joint position predicted based on Hill type EMG-force model in isotonic contractions^[285]; (c) A proposed system with sEMG and position measurements for in-motion estimation of ankle moment^[284]

Due to the limitation of current sensing technologies of activation in dynamic circumstances, no published research has been able to track the muscular tensions in sports and field training, even in very simple specific or chosen tasks. Ligament orientation and joint positions were captured by cameras, Louis^[288] studied the a common clinical routine of reach-grasp movements, only able to compare muscle tension determined between different EMG-force models. In a most frequently cited work, Lloyd^[41] illustrated general Hill-type modeling, used a modified EMG-driven model to calculate knee moments in crossover cuts and straight runs, and validated the model by comparing the calculated torque with that obtained from inverse dynamics. In this work, joint position was accessed by electrogoniometer. Langenderfer^[42] used tuned EMG-driven Hill model in isometric flexions for determining contractility of individual skeletal muscles in fore and upper arms, but only able to compare net moment around elbow joint between measured and predicted. A most recent dynamic application of the EMG-driven Hill model was revealed by Lee^[289] on predicting tension of goat gastrocnemius muscles during walking and running, with the RMS error observed as high as 32%. In this work, muscle length and its time rate of change were obtained by lab-made ultrasound sensors. The reviewed applications above adequately and representatively reflect the current research gaps in monitoring muscle contractility based on activation-contractility modeling: First, although sEMG has been proven frequently effective to assess muscle activation during isometric contractions, it is far way from being reliable in dynamic conditions. As such, additional or alternative descriptions of activation based on other sensing technologies

are required; Secondly, simultaneous measurements of activation and contraction status (muscle length and shortening velocity) have been proposed in order to achieve a complete activation-contractility modeling for dynamic contractions, which, however, have not been truly realized until now. Therefore and thirdly, inconsistency of predictions from a model was reported for a diverse range of tasks now and then. Finally, there is still lack of direct or indirect sensing approaches for verification of the muscle tensions determined from the activation-contractility models.

5 Conclusions and Recommendations

Due to the lack of direct implanted transducers, indirect monitoring of contractility based on activation-contractility modeling are only feasible solution, involving an index of activation, the observation of force-length and force-relationship and the activation-contractility modeling. Hence, efforts have been given on the real-time monitoring on the intensity of skeletal muscles' contraction (which is actually the monitoring of indexes for activation of muscle contraction), especially on the non-invasive approaches. A brief overview of both classical and up-to-date new technologies has been presented.

Up to date, the classic sEMG and MMG have not been applied in quantitative studies in dynamic motions, although they are used as an index of muscle activation in isometric or static condition. sEMG and MMG encountered significant noises problems especially in field activities. Imaging methods such as ultrasound scanning, despite their inconvenience for out-laboratory or in-field applications, have inspired a variety of other novel technologies on monitoring muscle contraction, among

which continuous anthropometric measurement based on soft sensors appears to be effective and have been repeatedly understood as another index of activation and introduced in strain-contraction modeling. Anthropometric measurement based on fabric strain gauges also show great potentials for dynamic conditions such as in-field training. Applying such anthropometric techniques requires better understanding of strain-activation mechanism. Moreover, status parameters of contraction status, i.e., muscle length and shortening velocity, need to be obtained experimentally for independent monitoring of contractility,

In activation-contraction modeling, the effects of status parameters on muscle contraction, the force-length and force-velocity properties should be determined. For the force-length relationship, it is not recommended to scale an ideal biophysical parameterized force-length relationship for sarcomere to fit the target skeletal muscle, due to too many influencing factors difficult to reflect. To obtain the force-length relationship experimentally, there is a challenge for maintaining the 'intention of muscle contraction' (activation). Meanwhile, the difference between the torque-joint angle relationship and the real force-length relationship of skeletal muscle obtained in isometric tests shall also be noted. With regard to the force-velocity relationship, one should be aware is that the 'force' is either an index of load/resistance or contractility deduced inversely from devices such as force plates while the 'velocity' is not exactly the shortening velocity but a speed of the motion, only serves as representative of skeletal muscles' shortening velocity only. Literatures show that both the activation and muscle length impact the observed force-velocity relationship. While the force-velocity relationship of isolated

muscles has been known to be hyperbolic, the multi-joint functional tasks typically reveal strong and approximately a linear force-velocity relationship, though biological mechanisms remain elusive.

The accuracy of a Hill type model on predicting muscle characteristics and contraction relies on the bio-signals as indexes of activations, the force-length and force-velocity relationship. To construct a Hill-type biomechanical model for in-motion monitoring of contraction, it is desirable to determine the core elements of a Hill type muscle through isolated designed tasks and selected conditions. However, since the model of a single sarcomere and that of a muscle may reveal huge difference due to the assumption of averaged shortening of sarcomeres and the recruitment patterns of slower and faster motor units, as well as impedance caused by connective tissue and bones, it is recommended to give comprehensive consideration of the effect of specified length and velocity while focusing on the activation-contraction correlation, instead of product of the obtained factors as in the classical Hill's model.

Furthermore, the status of contraction, i.e., tension, muscle length and shortening speed are represented by joint torque, joint position (or joint angle) and joint angular velocity, respectively in the human musculoskeletal system, based on angle-to-length and angular velocity-to-shortening speed transform functions either with previously reported anatomical parameters or inversely derived from experiments. Hence, direct measurement of joint angles is not only essential for determination of force-length and force-velocity relationships but also for independent monitoring of contraction in-motion. A review

of joint angle measurement technologies is presented, indicating that for in-motion conditions, soft goniometers are potentially better choices compared to camera-based motion capture system that is limited in laboratory and IMUS hindering normal movements. Further research is required to enhance accuracy, reliability of soft goniometers. If successful, an activation-contraction solution for motions can be completed and utilized as the effective monitoring, diagnosis and rehabilitation tools.

Acknowledgements

The authors acknowledge the funding supports from Innovation and Technology Commission, Hong Kong SAR Government (grant no. ITT/035/14TP, ITT/011/11TT and ITP/039/16TP).

References

1. R. Hale and S. Mookerjee, *EMG Amplitude-to-torque Ratios In Males And Females During Isokinetic Exercise*. *Medicine and Science in Sports and Exercise*, 2014. **46**(5): p. 189-190. DOI: 10.1249/01.mss.0000493748.38383.b4
2. R.C.H. So, J.K.F. Ng, R.W.K. Lam, C.K.K. Lo, and G.Y.F. Ng, *EMG Wavelet Analysis of Quadriceps Muscle during Repeated Knee Extension Movement*. *Medicine and Science in Sports and Exercise*, 2009. **41**(4): p. 788-796. DOI: 10.1249/MSS.0b013e31818cb4d0
3. R.M. Campy, A.J. Coelho, and D.M. Pincivero, *EMG-torque relationship and reliability of the medial and lateral hamstring muscles*. *Med Sci Sports Exerc*, 2009. **41**(11): p. 2064-71. DOI: 10.1249/MSS.0b013e3181a8c4cb
4. O.M. Blake, Y. Champoux, and J.M. Wakeling, *Muscle coordination patterns for efficient cycling*. *Med Sci Sports Exerc*, 2012. **44**(5): p. 926-38. DOI: 10.1249/MSS.0b013e3182404d4b
5. G. Wei, F. Tian, G. Tang, and C. Wang, *A Wavelet-Based Method to Predict Muscle Forces From Surface Electromyography Signals in Weightlifting*. *Journal of Bionic Engineering*, 2012. **9**(1): p. 48-58. DOI: 10.1016/s1672-6529(11)60096-6
6. J. Marusiak, A. Jaskólska, K. Kisiel-Sajewicz, G.H. Yue, and A. Jaskólski, *EMG and MMG activities of agonist and antagonist muscles in Parkinson's disease patients during absolute submaximal load holding*. *Journal of electromyography and kinesiology*, 2009. **19**(5): p. 903-914. DOI: 10.1016/j.jelekin.2008.03.003
7. R.P. Hubble, G.A. Naughton, P.A. Silburn, and M.H. Cole, *Wearable Sensor Use for Assessing Standing Balance and Walking Stability in People with Parkinson's Disease: A Systematic Review*. *Plos One*, 2015. **10**(4). DOI: 10.1371/journal.pone.0123705
8. J. Shi, Y.P. Zheng, X. Chen, and Q.H. Huang, *Assessment of muscle fatigue using sonomyography: Muscle thickness change detected from ultrasound images*. *Medical Engineering & Physics*, 2007. **29**(4): p. 472-479. DOI: 10.1016/j.medengphy.2006.07.004
9. K.T. Ebersole and D.M. Malek, *Fatigue and the electromechanical efficiency of the vastus medialis and vastus lateralis muscles*. *J Athl Train*, 2008. **43**(2): p. 152-6. DOI: 10.4085/1062-6050-43.2.152
10. C.R. Hendrix, T.J. Housh, G.O. Johnson, M. Mielke, C.L. Camic, J.M. Zuniga, and R.J. Schmidt, *Comparison of critical force to EMG fatigue thresholds during isometric leg extension*. *Med Sci Sports Exerc*, 2009. **41**(4): p. 956-64. DOI: 10.1249/MSS.0b013e318190bdf7
11. M. Song, D.B. Segala, J.B. Dingwell, and D. Chelidze, *Slow-time changes in human EMG muscle fatigue states are fully represented in movement kinematics*. *J Biomech Eng*, 2009. **131**(2): p. 021004. DOI: 10.1115/1.3005177
12. J.M. Garcia-Manso, D. Rodriguez-Ruiz, D. Rodriguez-Matoso, Y. de Saa, S. Sarmiento, and M. Quiroga, *Assessment of muscle fatigue after an ultra-*

- endurance triathlon using tensiomyography (TMG)*. *Journal of Sports Sciences*, 2011. **29**(6): p. 619-625. DOI: 10.1080/02640414.2010.548822
13. H. Han, S. Jo, and J. Kim, *Comparative study of a muscle stiffness sensor and electromyography and mechanomyography under fatigue conditions*. *Medical & Biological Engineering & Computing*, 2015. **53**(7): p. 577-588. DOI: 10.1007/s11517-015-1271-1
 14. W. Zeng, L. Shu, Q. Li, S. Chen, F. Wang, and X.M. Tao, *Fiber-based wearable electronics: a review of materials, fabrication, devices, and applications*. *Adv Mater*, 2014. **26**(31): p. 5310-36. DOI: 10.1002/adma.201400633
 15. P.W. Hodges, L.H.M. Pengel, R.D. Herbert, and S.C. Gandevia, *Measurement of muscle contraction with ultrasound imaging*. *Muscle & Nerve*, 2003. **27**(6): p. 682-692. DOI: 10.1002/Mus.10375
 16. E.D. Ryan, T.W. Beck, T.J. Herda, M.J. Hartman, J.R. Stout, T.J. Housh, and J.T. Cramer, *Mechanomyographic amplitude and mean power frequency responses during isometric ramp vs. step muscle actions*. *Journal of Neuroscience Methods*, 2008. **168**(2): p. 293-305. DOI: 10.1016/j.jneumeth.2007.10.010
 17. H.S. Milner-Brown and R.B. Stein, *The relation between the surface electromyogram and muscular force*. *The Journal of Physiology*, 1975. **246**(3): p. 549-569. DOI: 10.1113/jphysiol.1975.sp010904
 18. J.H. Lawrence and C.J. De Luca, *Myoelectric Signal Versus Force Relationship In Different Human Muscles*. *Journal Of Applied Physiology*, 1983. **54**(6): p. 1653-1659. DOI: 10.1152/jappl.1983.54.6.1653
 19. E.A. Clancy, O. Bida, and D. Rancourt, *Influence of advanced electromyogram (EMG) amplitude processors on EMG-to-torque estimation during constant-posture, force-varying contractions*. *Journal Of Biomechanics*, 2006. **39**(14): p. 2690-2698. DOI: 10.1016/j.jbiomechs.2005.08.007
 20. J. Shi, Y.P. Zheng, Q.H. Huang, and X. Chen, *Continuous monitoring of sonomyography, electromyography and torque generated by normal upper arm muscles during isometric contraction: Sonomyography assessment for arm muscles*. *Ieee Transactions on Biomedical Engineering*, 2008. **55**(3): p. 1191-1198. DOI: 10.1109/Tbme.2007.909538
 21. J.Y. Guo, Y.P. Zheng, H.B. Xie, and X. Chen, *Continuous monitoring of electromyography (EMG), mechanomyography (MMG), sonomyography (SMG) and torque output during ramp and step isometric contractions*. *Medical Engineering & Physics*, 2010. **32**(9): p. 1032-1042. DOI: 10.1016/j.medengphy.2010.07.004
 22. N. Shima, C.J. McNeil, and C.L. Rice, *Mechanomyographic and electromyographic responses to stimulated and voluntary contractions in the dorsiflexors of young and old men*. *Muscle & Nerve*, 2007. **35**(3): p. 371-378. DOI: 10.1002/Mus.20704
 23. S.R. Perry-Rana, T.J. Housh, G.O.

- Johnson, A.J. Bull, and J.T. Cramer, *MMG and EMG responses during 25 maximal, eccentric, isokinetic muscle actions*. *Medicine and Science in Sports and Exercise*, 2003. **35**(12): p. 2048-2054. DOI: 10.1249/01.Mss.0000099090.73560.77
24. E.J. Jones, P.A. Bishop, A.K. Woods, and F.M. Green, *Cross-Sectional Area and Muscular Strength A Brief Review*. *Sports Medicine*, 2008. **38**(12): p. 987-994. DOI: 10.2165/00007256-200838120-00003
25. J. Davies, D.F. Parker, O.M. Rutherford, and D.A. Jones, *Changes In Strength And Cross-Sectional Area Of the Elbow Flexors as a Result Of Isometric Strength Training*. *European Journal Of Applied Physiology And Occupational Physiology*, 1988. **57**(6): p. 667-670. DOI: 10.1007/Bf01075986
26. M.V. Narici, G.S. Roi, L. Landoni, A.E. Minetti, and P. Cerretelli, *Changes In Force, Cross-Sectional Area And Neural Activation during Strength Training And Detraining Of the Human Quadriceps*. *European Journal Of Applied Physiology And Occupational Physiology*, 1989. **59**(4): p. 310-319. DOI: 10.1007/Bf02388334
27. T. Fukunaga, M. Miyatani, M. Tachi, M. Kouzaki, Y. Kawakami, and H. Kanehisa, *Muscle volume is a major determinant of joint torque in humans*. *Acta Physiologica Scandinavica*, 2001. **172**(4): p. 249-255. DOI: 10.1046/j.1365-201x.2001.00867.x
28. R. Akagi, Y. Takai, M. Ohta, H. Kanehisa, Y. Kawakami, and T. Fukunaga, *Muscle volume compared to cross-sectional area is more appropriate for evaluating muscle strength in young and elderly individuals*. *Age And Ageing*, 2009. **38**(5): p. 564-569. DOI: 10.1093/ageing/afp122
29. K. Hakkinen, M. Kallinen, M. Izquierdo, K. Jokelainen, H. Lassila, E. Malkia, W.J. Kraemer, R.U. Newton, and M. Alen, *Changes in agonist-antagonist EMG, muscle CSA, and force during strength training in middle-aged and older people*. *J Appl Physiol* (1985), 1998. **84**(4): p. 1341-9. DOI: 10.1152/jappl.1998.84.4.1341
30. M.B. Raez, M.S. Hussain, and F. Mohd-Yasin, *Techniques of EMG signal analysis: detection, processing, classification and applications*. *Biol Proced Online*, 2006. **8**: p. 11-35. DOI: 10.1251/bpo115
31. J.L. Dantas, T.V. Camata, M.A. Brunetto, A.C. Moraes, T. Abrao, and L.R. Altimari, *Fourier and wavelet spectral analysis of EMG signals in isometric and dynamic maximal effort exercise*. *Conf Proc IEEE Eng Med Biol Soc*, 2010. **2010**: p. 5979-82. DOI: 10.1109/IEMBS.2010.5627579
32. J.Y. Hogrel, *Use of surface EMG for studying motor unit recruitment during isometric linear force ramp*. *Journal of Electromyography and Kinesiology*, 2003. **13**(5): p. 417-423. DOI: 10.1016/s1050-6411(03)00026-9
33. P. Liu, L. Liu, F. Martel, D. Rancourt, and E.A. Clancy, *Influence of joint angle on EMG-torque model during constant-posture, quasi-constant-torque contractions*. *J Electromyogr Kinesiol*, 2013. **23**(5): p. 1020-8. DOI: 10.1016/j.jelekin.2013.06.011
34. E.A. Clancy, L. Liu, P. Liu, and D.V.

- Moyer, *Identification of constant-posture EMG-torque relationship about the elbow using nonlinear dynamic models*. IEEE Trans Biomed Eng, 2012. **59**(1): p. 205-12. DOI: 10.1109/TBME.2011.2170423
35. H. Cao, S. Boudaoud, F. Marin, and C. Marque, *Surface EMG-force modelling for the biceps brachii and its experimental evaluation during isometric isotonic contractions*. Comput Methods Biomech Biomed Engin, 2015. **18**(9): p. 1014-1023. DOI: 10.1080/10255842.2013.867952
36. T.I. Suvinen and P. Kempainen, *Review of clinical EMG studies related to muscle and occlusal factors in healthy and TMD subjects*. J Oral Rehabil, 2007. **34**(9): p. 631-44. DOI: 10.1111/j.1365-2842.2007.01769.x
37. R.C. So, J.K. Ng, R.W. Lam, C.K. Lo, and G.Y. Ng, *EMG wavelet analysis of quadriceps muscle during repeated knee extension movement*. Med Sci Sports Exerc, 2009. **41**(4): p. 788-96. DOI: 10.1249/MSS.0b013e31818cb4d0
38. E. Fujita, H. Kanehisa, Y. Yoshitake, T. Fukunaga, and H. Nishizono, *Association between knee extensor strength and EMG activities during squat movement*. Med Sci Sports Exerc, 2011. **43**(12): p. 2328-34. DOI: 10.1249/MSS.0b013e3182207ed8
39. T.W. Beck, M.S. Stock, and J.M. Defreitas, *Shifts in EMG spectral power during fatiguing dynamic contractions*. Muscle Nerve, 2014. **50**(1): p. 95-102. DOI: 10.1002/mus.24098
40. M. Sartori, M. Reggiani, D. Farina, and D.G. Lloyd, *EMG-driven forward-dynamic estimation of muscle force and joint moment about multiple degrees of freedom in the human lower extremity*. PLoS One, 2012. **7**(12): p. e52618. DOI: 10.1371/journal.pone.0052618
41. D.G. Lloyd and T.F. Besier, *An EMG-driven musculoskeletal model to estimate muscle forces and knee joint moments in vivo*. Journal of Biomechanics, 2003. **36**(6): p. 765-776. DOI: 10.1016/s0021-9290(03)00010-1
42. J. Langenderfer, S. LaScalza, A. Mell, J.E. Carpenter, J.E. Kuhn, and R.E. Hughes, *An EMG-driven model of the upper extremity and estimation of long head biceps force*. Comput Biol Med, 2005. **35**(1): p. 25-39. DOI: 10.1016/j.compbiomed.2003.12.002
43. D. Shin, J. Kim, and Y. Koike, *A myokinetic arm model for estimating joint torque and stiffness from EMG signals during maintained posture*. J Neurophysiol, 2009. **101**(1): p. 387-401. DOI: 10.1152/jn.00584.2007
44. S. Karlsson and B. Gerdle, *Mean frequency and signal amplitude of the surface EMG of the quadriceps muscles increase with increasing torque--a study using the continuous wavelet transform*. J Electromyogr Kinesiol, 2001. **11**(2): p. 131-40. DOI: 10.1016/S1050-6411(00)00046-8
45. N.J. Cronin, S. Kumpulainen, T. Joutjarvi, T. Finni, and H. Piitulainen, *Spatial variability of muscle activity during human walking: the effects of different EMG normalization approaches*. Neuroscience, 2015. **300**: p. 19-28. DOI: 10.1016/j.neuroscience.2015.05.003
46. N. Ball and J. Scurr, *Electromyography normalization methods for high-velocity*

- muscle actions: review and recommendations.* J Appl Biomech, 2013. **29**(5): p. 600-8. DOI: 10.1123/jab.29.5.600
47. W. Youn and J. Kim, *Estimation of elbow flexion force during isometric muscle contraction from mechanomyography and electromyography.* Med Biol Eng Comput, 2010. **48**(11): p. 1149-57. DOI: 10.1007/s11517-010-0641-y
48. D. Farina and R. Merletti, *Comparison of algorithms for estimation of EMG variables during voluntary isometric contractions.* J Electromyogr Kinesiol, 2000. **10**(5): p. 337-49. DOI: 10.1016/S1050-6411(00)00025-0
49. K. Brzostowski and J. Swiatek, *Different Approaches to Model Relationship Between EMG Signals and Force Moments in Human Skeletal Muscle. Analysis for Diagnosis of Neuronmuscular Disorders.* Fundamenta Informaticae, 2009. **96**(4): p. 465-475. DOI: 10.3233/Fi-2009-188
50. Q. Shao, D.N. Bassett, K. Manal, and T.S. Buchanan, *An EMG-driven model to estimate muscle forces and joint moments in stroke patients.* Comput Biol Med, 2009. **39**(12): p. 1083-8. DOI: 10.1016/j.compbimed.2009.09.002
51. E.A. Clancy, E.L. Morin, and R. Merletti, *Sampling, noise-reduction and amplitude estimation issues in surface electromyography.* Journal of Electromyography and Kinesiology, 2002. **12**(1): p. 1-16. DOI: 10.1016/S1050-6411(01)00033-5
52. E. Huigen, A. Peper, and C.A. Grimbergen, *Investigation into the origin of the noise of surface electrodes.* Medical & Biological Engineering & Computing, 2002. **40**(3): p. 332-338. DOI: 10.1007/Bf02344216
53. S. Nishimura, Y. Tomita, and T. Horiuchi, *Clinical-Application of an Active Electrode Using an Operational-Amplifier.* Ieee Transactions on Biomedical Engineering, 1992. **39**(10): p. 1096-1099. DOI: 10.1109/10.161342
54. A. Hof, *The relationship between electromyogram and muscle force.* Sportverletzung· Sportschaden, 1997. **11**(03): p. 79-86. DOI: 10.1055/s-2007-993372
55. A.L. Hof, *Muscle mechanics and neuromuscular control.* Journal of Biomechanics, 2003. **36**(7): p. 1031-1038. DOI: 10.1016/S0021-9290(03)00036-8
56. M. Tanaka, T. Okuyama, and K. Saito, *Study on evaluation of muscle conditions using a mechanomyogram sensor.* 2011 Ieee International Conference on Systems, Man, and Cybernetics (Smc), 2011: p. 741-745. URL: <https://www.thieme-connect.com/products/ejournals/abstract/10.1055/s-2007-993372>
57. S. Kawakami, N. Kodama, N. Maeda, S. Sakamoto, K. Oki, Y. Yanagi, J.I. Asaumi, T. Maeda, and S. Minagi, *Mechanomyographic activity in the human lateral pterygoid muscle during mandibular movement.* Journal of Neuroscience Methods, 2012. **203**(1): p. 157-162. DOI: 10.1016/j.jneumeth.2011.09.026
58. L.P. Qi, J.M. Wakeling, A. Green, K. Lambrecht, and M. Ferguson-Pell, *Spectral properties of*

- electromyographic and mechanomyographic signals during isometric ramp and step contractions in biceps brachii.* Journal of Electromyography and Kinesiology, 2011. **21**(1): p. 128-135. DOI: 10.1016/j.jelekin.2010.09.006
59. K.F. Lei, W.W. Tsai, W.Y. Lin, and M.Y. Lee, *MMG-Torque Estimation under Dynamic Contractions.* 2011 Ieee International Conference on Systems, Man, and Cybernetics (Smc), 2011: p. 585-590. URL: <http://ieeexplore.ieee.org/abstract/document/6083774/>
60. E.M. Scheeren, E. Krueger-Beck, G. Nogueira-Neto, P. Nohama, and V.L.d.S.N. Button, *Wrist Movement Characterization by Mechanomyography.* Journal of Medical and Biological Engineering, 2010. **30**(6): p. 373-380. DOI: 10.5405/jmbe.757
61. C. Orizio, M. Solomonow, B. Diemont, and M. Gobbo, *Muscle-joint unit transfer function derived from torque and surface mechanomyogram in humans using different stimulation protocols.* Journal of Neuroscience Methods, 2008. **173**(1): p. 59-66. DOI: 10.1016/j.jneumeth.2008.05.012
62. T.W. Beck, M.A. Dillon, J.M. DeFreitas, and M.S. Stock, *Cross-correlation analysis of mechanomyographic signals detected in two axes.* Physiological Measurement, 2009. **30**(12): p. 1465-1471. DOI: 10.1088/0967-3334/30/12/012
63. M.O. Ibitoye, N.A. Hamzaid, J.M. Zuniga, N. Hasnan, and A.K. Wahab, *Mechanomyographic parameter extraction methods: an appraisal for clinical applications.* Sensors (Basel), 2014. **14**(12): p. 22940-70. DOI: 10.3390/s141222940
64. T.W. Beck, T.J. Housh, J.T. Cramer, J.P. Weir, G.O. Johnson, J.W. Coburn, M.H. Malek, and M. Mielke, *Mechanomyographic amplitude and frequency responses during dynamic muscle actions: a comprehensive review.* Biomed Eng Online, 2005. **4**: p. 67. DOI: 10.1186/1475-925X-4-67
65. Y. Yoshitake, M. Shinohara, H. Ue, and T. Moritani, *Characteristics of surface mechanomyogram are dependent on development of fusion of motor units in humans.* J Appl Physiol (1985), 2002. **93**(5): p. 1744-52. DOI: 10.1152/jappphysiol.00008.2002
66. F.V. Brozovich and G.H. Pollack, *Muscle-Contraction Generates Discrete Sound Bursts.* Biophysical Journal, 1983. **41**(1): p. 35-40. DOI: 10.1016/S0006-3495(83)84403-8
67. N. Alves and T. Chau, *Stationarity distributions of mechanomyogram signals from isometric contractions of extrinsic hand muscles during functional grasping.* Journal of Electromyography and Kinesiology, 2008. **18**(3): p. 509-515. DOI: 10.1016/j.jelekin.2006.11.010
68. H.B. Xie, Y.P. Zheng, and J.Y. Guo, *Classification of the mechanomyogram signal using a wavelet packet transform and singular value decomposition for multifunction prosthesis control.* Physiological Measurement, 2009. **30**(5): p. 441-457. DOI: 10.1088/0967-3334/30/5/002
69. P.E. Taylor, G.J. Almeida, T. Kanade,

- and J.K. Hodgins, *Classifying human motion quality for knee osteoarthritis using accelerometers*. Conf Proc IEEE Eng Med Biol Soc, 2010. **2010**: p. 339-43. DOI: 10.1109/IEMBS.2010.5627665
70. S.L. Tian, Y. Liu, L. Li, W.J. Fu, and C.H. Peng, *Mechanomyography is more sensitive than EMG in detecting age-related sarcopenia*. Journal of Biomechanics, 2010. **43**(3): p. 551-556. DOI: 10.1016/j.jbiomech.2009.09.034
71. J. Silva, W. Heim, and T. Chau, *MMG-based classification of muscle activity for prosthesis control*. Conf Proc IEEE Eng Med Biol Soc, 2004. **2**: p. 968-71. DOI: 10.1109/IEMBS.2004.1403322
72. N. Alves, T.H. Falk, and T. Chau, *A novel integrated mechanomyogram-vocalization access solution*. Med Eng Phys, 2010. **32**(8): p. 940-4. DOI: 10.1016/j.medengphy.2010.06.003
73. C. Orizio, M. Gobbo, B. Diemont, F. Esposito, and A. Veicsteinas, *The surface mechanomyogram as a tool to describe the influence of fatigue on biceps brachii motor unit activation strategy. Historical basis and novel evidence*. Eur J Appl Physiol, 2003. **90**(3-4): p. 326-36. DOI: 10.1007/s00421-003-0924-1
74. E. Bichler, *Mechanomyograms recorded during evoked contractions of single motor units in the rat medial gastrocnemius muscle*. Eur J Appl Physiol, 2000. **83**(4 -5): p. 310-9. DOI: 10.1007/s004210000261
75. M.T. Tarata, *Mechanomyography versus electromyography, in monitoring the muscular fatigue*. Biomed Eng Online, 2003. **2**: p. 3. DOI: 10.1186/1475-925X-2-3
76. A. Archer and K.G. Sabra, *Two dimensional spatial coherence of the natural vibrations of the biceps brachii muscle generated during voluntary contractions*. Conf Proc IEEE Eng Med Biol Soc, 2010. **2010**: p. 170-3. DOI: 10.1109/IEMBS.2010.5627271
77. F. Esposito, E. Limonta, and E. Ce, *Time course of stretching-induced changes in mechanomyogram and force characteristics*. J Electromyogr Kinesiol, 2011. **21**(5): p. 795-802. DOI: 10.1016/j.jelekin.2011.07.012
78. J.T. Cramer, T.W. Beck, T.J. Housh, L.L. Massey, S.M. Marek, S. Danglemeier, S. Purkayastha, J.Y. Culbertson, K.A. Fitz, and A.D. Egan, *Acute effects of static stretching on characteristics of the isokinetic angle - torque relationship, surface electromyography, and mechanomyography*. J Sports Sci, 2007. **25**(6): p. 687-98. DOI: 10.1080/02640410600818416
79. G. Trager, G. Michaud, S. Deschamps, and T.M. Hernmerling, *Comparison of phonomyography, kinemyography and mechanomyography for neuromuscular monitoring*. Canadian Journal of Anaesthesia-Journal Canadien D Anesthesie, 2006. **53**(2): p. 130-135. DOI: 10.1007/Bf03021816
80. S.M. Marek, J.T. Cramer, A.L. Fincher, L.L. Massey, S.M. Dangelmaier, S. Purkayastha, K.A. Fitz, and J.Y. Culbertson, *Acute Effects of Static and Proprioceptive Neuromuscular Facilitation Stretching on Muscle Strength and Power Output*. J Athl Train, 2005. **40**(2): p. 94-103. DOI:

- 10.1016/S0162-0908(08)70360-X
81. E.D. Ryan, J.T. Cramer, T.J. Housh, T.W. Beck, T.J. Herda, and M.J. Hartman, *Inter-individual variability in the torque-related patterns of responses for mechanomyographic amplitude and mean power frequency*. *J Neurosci Methods*, 2007. **161**(2): p. 212-9. DOI: 10.1016/j.jneumeth.2006.11.007
82. J.T. Cramer, T.J. Housh, G.O. Johnson, K.T. Ebersole, S.R. Perry, and A.J. Bull, *Mechanomyographic amplitude and mean power output during maximal, concentric, isokinetic muscle actions*. *Muscle Nerve*, 2000. **23**(12): p. 1826-31.
83. K. Akataki, K. Mita, M. Watakabe, and K. Itoh, *Mechanomyographic responses during voluntary ramp contractions of the human first dorsal interosseous muscle*. *Eur J Appl Physiol*, 2003. **89**(6): p. 520-5. DOI: 10.1007/s00421-003-0835-1
84. J.W. Coburn, T.J. Housh, J.T. Cramer, J.P. Weir, J.M. Miller, T.W. Beck, M.H. Malek, and G.O. Johnson, *Mechanomyographic time and frequency domain responses of the vastus medialis muscle during submaximal to maximal isometric and isokinetic muscle actions*. *Electromyogr Clin Neurophysiol*, 2004. **44**(4): p. 247-55. URL: <https://www.ncbi.nlm.nih.gov/pubmed/15224821>
85. M.A. Islam, K. Sundaraj, R.B. Ahmad, and N.U. Ahamed, *Mechanomyogram for muscle function assessment: a review*. *PLoS One*, 2013. **8**(3): p. e58902. DOI: 10.1371/journal.pone.0058902
86. T.W. Beck, T.J. Housh, G.O. Johnson, J.P. Weir, J.T. Cramer, J.W. Coburn, and M.H. Malek, *Comparison of Fourier and wavelet transform procedures for examining the mechanomyographic and electromyographic frequency domain responses during fatiguing isokinetic muscle actions of the biceps brachii*. *J Electromyogr Kinesiol*, 2005. **15**(2): p. 190-9. DOI: 10.1016/j.jelekin.2004.08.007
87. R.L. Lieber and J. Friden, *Functional and clinical significance of skeletal muscle architecture*. *Muscle Nerve*, 2000. **23**(11): p. 1647-66. DOI: 10.1002/1097-4598(200011)23:11<1647::AID-MUS1>3.3.CO;2-D
88. R. Akagi, S. Iwanuma, S. Hashizume, H. Kanehisa, T. Fukunaga, and Y. Kawakami, *Determination of Contraction-Induced Changes in Elbow Flexor Cross-Sectional Area for Evaluating Muscle Size-Strength Relationship during Contraction*. *Journal of Strength and Conditioning Research*, 2015. **29**(6): p. 1741-1747. DOI: 10.1519/JSC.0000000000000793
89. D.B. Starkey, M.L. Pollock, Y. Ishida, M.A. Welsch, W.F. Brechue, J.E. Graves, and M.S. Feigenbaum, *Effect of resistance training volume on strength and muscle thickness*. *Medicine and Science in Sports and Exercise*, 1996. **28**(10): p. 1311-1320. DOI: 10.1097/00005768-199610000-00016
90. R. Akagi, S. Iwanuma, M. Fukuoka, H. Kanehisa, T. Fukunaga, and Y. Kawakami, *Methodological Issues Related to Thickness-Based Muscle Size Evaluation*. *Journal Of Physiological*

- Anthropology, 2011. **30**(4): p. 169-174. DOI: 10.2114/Jpa2.30.169
91. M. Miyatani, H. Kanehisa, M. Ito, Y. Kawakami, and T. Fukunaga, *The accuracy of volume estimates using ultrasound muscle thickness measurements in different muscle groups*. *Eur J Appl Physiol*, 2004. **91**(2-3): p. 264-72. DOI: 10.1007/s00421-003-0974-4
92. P.K. Commean, L.J. Tuttle, M.K. Hastings, M.J. Strube, and M.J. Mueller, *Magnetic resonance imaging measurement reproducibility for calf muscle and adipose tissue volume*. *J Magn Reson Imaging*, 2011. **34**(6): p. 1285-94. DOI: 10.1002/jmri.22791
93. A.R. Seo, H.Y. Jang, W.S. Kim, C.S. Han, and J.S. Han, *Development and verification of a volume sensor for measuring human behavior*. *International Journal of Precision Engineering and Manufacturing*, 2012. **13**(6): p. 899-904. DOI: 10.1007/s12541-012-0117-0
94. A. Macaluso, M.A. Nimmo, J.E. Foster, M. Cockburn, N.C. McMillan, and G. De Vito, *Contractile muscle volume and agonist-antagonist coactivation account for differences in torque between young and older women*. *Muscle Nerve*, 2002. **25**(6): p. 858-63. DOI: 10.1002/mus.10113
95. G. Chi-Fishman, J.E. Hicks, H.M. Cintas, B.C. Sonies, and L.H. Gerber, *Ultrasound imaging distinguishes between normal and weak muscle*. *Archives of Physical Medicine and Rehabilitation*, 2004. **85**(6): p. 980-986. DOI: 10.1016/j.apmr.2003.07.008
96. M. Ikai and T. Fukunaga, *A Study on Training Effect on Strength Per Unit Cross-Sectional Area of Muscle by Means of Ultrasonic Measurement*. *Internationale Zetischrift Fur Angewandte Physiologie Einschliesslich Arbeitsphysiologie*, 1970. **28**(3): p. 173-180. DOI: 10.1007/BF00696025
97. A. Wilson, J.A. Hides, L. Blizzard, M. Callisaya, A. Cooper, V.K. Srikanth, and T. Winzenberg, *Measuring ultrasound images of abdominal and lumbar multifidus muscles in older adults: A reliability study*. *Man Ther*, 2016. **23**: p. 114-9. DOI: 10.1016/j.math.2016.01.004
98. M.V. Franchi, P.J. Atherton, N.D. Reeves, M. Fluck, J. Williams, W.K. Mitchell, A. Selby, R.M. Beltran Valls, and M.V. Narici, *Architectural, functional and molecular responses to concentric and eccentric loading in human skeletal muscle*. *Acta Physiol (Oxf)*, 2014. **210**(3): p. 642-54. DOI: 10.1111/apha.12225
99. Y. Qi, C.B. Soh, E. Gunawan, K.S. Low, and R. Thomas, *Lower Extremity Joint Angle Tracking with Wireless Ultrasonic Sensors during a Squat Exercise*. *Sensors (Basel)*, 2015. **15**(5): p. 9610-27. DOI: 10.3390/s150509610
100. L.K. Kwah, R.Z. Pinto, J. Diong, and R.D. Herbert, *Reliability and validity of ultrasound measurements of muscle fascicle length and pennation in humans: a systematic review*. *J Appl Physiol (1985)*, 2013. **114**(6): p. 761-9. DOI: 10.1152/jappphysiol.01430.2011
101. E.M. Strasser, T. Draskovits, M. Praschak, M. Quittan, and A. Graf, *Association between ultrasound*

- measurements of muscle thickness, pennation angle, echogenicity and skeletal muscle strength in the elderly.* Age, 2013. **35**(6): p. 2377-2388. DOI: 10.1007/s11357-013-9517-z
102. J.G. Gillett, R.S. Barrett, and G.A. Lichtwark, *Reliability and accuracy of an automated tracking algorithm to measure controlled passive and active muscle fascicle length changes from ultrasound.* Computer Methods in Biomechanics and Biomedical Engineering, 2013. **16**(6): p. 678-687. DOI: 10.1080/10255842.2011.633516
103. A. Cuesta-Vargas and M. Gonzalez-Sanchez, *Correlation between architectural variables and torque in the erector spinae muscle during maximal isometric contraction.* J Sports Sci, 2014. **32**(19): p. 1797-804. DOI: 10.1080/02640414.2014.924054
104. R.E. Stafford, J.A. Ashton-Miller, C.E. Constantinou, and P.W. Hodges, *A New Method to Quantify Male Pelvic Floor Displacement From 2D Transperineal Ultrasound Images.* Urology, 2013. **81**(3): p. 685-689. DOI: 10.1016/j.urology.2012.11.034
105. M. Leitner, H. Moser, J. Taeymans, A. Kuhn, and L. Radlinger, *Pelvic floor muscle displacement during voluntary and involuntary activation in continent and incontinent women: a systematic review.* International Urogynecology Journal, 2015. **26**(11): p. 1587-1598. DOI: 10.1007/s00192-015-2700-2
106. Y.P. Zheng, M.M.F. Chan, J. Shi, X. Chen, and Q.H. Huang, *Sonomyography: Monitoring morphological changes of forearm muscles in actions with the feasibility for the control of powered prosthesis.* Medical Engineering & Physics, 2006. **28**(5): p. 405-415. DOI: 10.1016/j.medengphy.2005.07.012
107. D. Guo, S.B. Bai, and Q. Wang, *A novel halogen-free flame retardant poly (vinyl alcohol) foam with intrinsic flame retardant characteristics prepared through continuous extrusion.* Journal of Cellular Plastics, 2015. **51**(2): p. 145-163. DOI: 10.1177/0021955X14529296
108. P. Silitertpisan, U. Pirunsan, A. Puangmali, J. Ratanapinunchai, S. Kiatwattanacharoen, H. Neamin, and J.J. Laskin, *Comparison of lateral abdominal muscle thickness between weightlifters and matched controls.* Phys Ther Sport, 2011. **12**(4): p. 171-4. DOI: 10.1016/j.ptsp.2011.02.002
109. R.G. Timmins, A.J. Shield, M.D. Williams, C. Lorenzen, and D.A. Opar, *Architectural adaptations of muscle to training and injury: a narrative review outlining the contributions by fascicle length, pennation angle and muscle thickness.* Br J Sports Med, 2016. DOI: 10.1136/bjsports-2015-094881
110. J.-Y. Guo, *Dynamic monitoring of forearm muscles using one-dimensional sonomyography system.* The Journal of Rehabilitation Research and Development, 2008. **45**(1): p. 187-196. DOI: 10.1682/jrrd.2007.02.0026
111. P.W. Hodges, L.H. Pengel, R.D. Herbert, and S.C. Gandevia, *Measurement of muscle contraction with ultrasound imaging.* Muscle Nerve, 2003. **27**(6): p. 682-92. DOI: 10.1002/mus.10375
112. R. Akagi, S. Iwanuma, S. Hashizume, H. Kanehisa, T. Yanai, and Y.

- Kawakami, *Association Between Contraction-Induced Increases in Elbow Flexor Muscle Thickness and Distal Biceps Brachii Tendon Moment Arm Depends on the Muscle Thickness Measurement Site*. *Journal of Applied Biomechanics*, 2014. **30**(1): p. 134-139. DOI: 10.1123/jab.2012-0145
113. T. Abe, J.P. Loenneke, and R.S. Thiebaud, *Morphological and functional relationships with ultrasound measured muscle thickness of the lower extremity: a brief review*. *Ultrasound*, 2015. **23**(3): p. 166-173. DOI: 10.1177/1742271X15587599
114. P.W. Hodges, *Ultrasound imaging in rehabilitation: Just a fad?* *Journal of Orthopaedic & Sports Physical Therapy*, 2005. **35**(6): p. 333-337. DOI: 10.2519/jospt.2005.0106
115. R.D. Herbert, A.M. Moseley, J.E. Butler, and S.C. Gandevia, *Change in length of relaxed muscle fascicles and tendons with knee and ankle movement in humans*. *Journal of Physiology-London*, 2002. **539**(2): p. 637-645. DOI: 10.1013/jphysiol.2001.012756
116. L. Ito, Y. Kawakami, Y. Ichinose, S. Fukashiro, and T. Fukunaga, *Nonisometric behavior of fascicles during isometric contractions of a human muscle*. *Journal of Applied Physiology*, 1998. **85**(4): p. 1230-1235. DOI: 10.1152/jappl.1998.85.4.1230
117. M.V. Narici, T. Binzoni, E. Hiltbrand, J. Fasel, F. Terrier, and P. Cerretelli, *In vivo human gastrocnemius architecture with changing joint angle at rest and during graded isometric contraction*. *Journal of Physiology-London*, 1996. **496**(1): p. 287-297. DOI: 10.1113/jphysiol.1996.sp021685
118. L.F. de Oliveira and L.L. Menegaldo, *Individual-specific muscle maximum force estimation using ultrasound for ankle joint torque prediction using an EMG-driven Hill-type model*. *Journal of Biomechanics*, 2010. **43**(14): p. 2816-2821. DOI: 10.1016/j.jbiomech.2010.05.035
119. J.M. Garcia, B. Calvo, L. Monteiro, L. Massuca, J. Portillo, and J. Abian-Vicen, *Impact of hydration on muscle contraction properties of elite competitive wrestlers*. *Archives of Budo*, 2016. **12**: p. 25-34. URL: <http://hdl.handle.net/10578/8157>
120. I. Loturco, L.A. Pereira, R. Kobal, K. Kitamura, R. Ramirez-Campillo, V. Zanetti, C.C.C. Abad, and F.Y. Nakamura, *Muscle Contraction Velocity: A Suitable Approach to Analyze the Functional Adaptations in Elite Soccer Players*. *Journal of Sports Science and Medicine*, 2016. **15**(3): p. 483-491. URL: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4974861/>
121. H.V. Wilson, M.I. Johnson, and P. Francis, *Repeated stimulation, inter-stimulus interval and inter-electrode distance alters muscle contractile properties as measured by Tensiomyography*. *Plos One*, 2018. **13**(2). DOI: ARTN e0191965 10.1371/journal.pone.0191965
122. P. Alvarez-Diaz, E. Alentorn-Geli, S. Ramon, M. Marin, G. Steinbacher, M. Rius, R. Seijas, J. Ballester, and R. Cugat, *Comparison of tensiomyographic neuromuscular characteristics between muscles of the*

- dominant and non-dominant lower extremity in male soccer players.* *Knee Surgery Sports Traumatology Arthroscopy*, 2016. **24**(7): p. 2259-2263. DOI: 10.1007/s00167-014-3298-5
123. R.A.D. Simola, N. Harms, C. Raeder, M. Kellmann, T. Meyer, M. Pfeiffer, and A. Ferrauti, *Assessment of Neuromuscular Function after Different Strength Training Protocols Using Tensiomyography.* *Journal of Strength and Conditioning Research*, 2015. **29**(5): p. 1339-1348. DOI: 10.1519/JSC.0000000000000768
124. A.K. Bansal, S.B. Hou, O. Kulyk, E.M. Bowman, and I.D.W. Samuel, *Wearable Organic Optoelectronic Sensors for Medicine.* *Advanced Materials*, 2015. **27**(46): p. 7638-+. DOI: 10.1002/adma.201403560
125. A. Chianura and M.E. Giardini, *An electrooptical muscle contraction sensor.* *Medical & Biological Engineering & Computing*, 2010. **48**(7): p. 731-734. DOI: 10.1007/s11517-010-0626-x
126. L. Cen, H. Han, and J. Kim, *Optical muscle activation sensors for estimating upper limb force level.* 2011 *Ieee International Instrumentation and Measurement Technology Conference (I2mtc)*, 2011: p. 1657-1660. URL: <http://ieeexplore.ieee.org/abstract/document/5944228/>
127. M. Belau, M. Ninck, G. Hering, L. Spinelli, D. Contini, A. Torricelli, and T. Gisler, *Noninvasive observation of skeletal muscle contraction using near-infrared time-resolved reflectance and diffusing-wave spectroscopy.* *Journal of Biomedical Optics*, 2010. **15**(5). DOI: Artn 057007 10.1117/1.3503398
128. M. Ferrari, M. Muthalib, and V. Quaresima, *The use of near-infrared spectroscopy in understanding skeletal muscle physiology: recent developments.* *Philosophical Transactions of the Royal Society a-Mathematical Physical and Engineering Sciences*, 2011. **369**(1955): p. 4577-4590. DOI: 10.1098/rsta.2011.0230
129. I. AlMohimeed, H. Turkistani, and Y. Ono, *Development of Wearable and Flexible Ultrasonic Sensor for Skeletal Muscle Monitoring.* 2013 *Ieee International Ultrasonics Symposium (Ius)*, 2013: p. 1129-1132. DOI: 10.1109/Ultsym.2013.0291
130. N. Hettiarachchi, Z.J. Ju, and H.H. Liu, *A New Wearable Ultrasound Muscle Activity Sensing System for Dexterous Prosthetic Control.* 2015 *Ieee International Conference on Systems, Man, and Cybernetics (Smc 2015): Big Data Analytics for Human-Centric Systems*, 2015: p. 1415-1420. DOI: 10.1109/Smc.2015.251
131. H. Han and J. Kim, *Active muscle stiffness sensor based on piezoelectric resonance for muscle contraction estimation.* *Sensors and Actuators a-Physical*, 2013. **194**: p. 212-219. DOI: 10.1016/j.sna.2013.01.054
132. W.S. Kim, H.D. Lee, D.H. Lim, J.S. Han, K.S. Shin, and C.S. Han, *Development of a muscle circumference sensor to estimate torque of the human elbow joint.* *Sensors and Actuators a-Physical*, 2014. **208**: p. 95-103. DOI: 10.1016/j.sna.2013.12.036
133. X. Wang, X.M. Tao, R.C.H. So, L. Shu, B. Yang, and Y. Li, *Monitoring elbow*

- isometric contraction by novel wearable fabric sensing device*. *Smart Materials and Structures*, 2016. **25**(12). DOI: Artn 125022 10.1088/0964-1726/25/12/125022
134. X. Wang, X.M. Tao, and R.C.H. So, *A Bio-mechanical Model for Elbow Isokinetic and Isotonic Flexions*. *Scientific Reports*, 2017. **7**. DOI: Artn 8919 10.1038/S41598-017-09071-X
135. P. Bifulco, D. Esposito, G.D. Gargiulo, S. Savino, V. Niola, L. Iuppariello, and M. Cesarelli. *A stretchable, conductive rubber sensor to detect muscle contraction for prosthetic hand control*. in *2017 E-Health and Bioengineering Conference (EHB)*. 2017. DOI: 10.1109/EHB.2017.7995389
136. D. Zhang, Y. Matsuoka, W. Kong, U. Imtiaz, L. Bartolomeo, S. Cosentino, M. Zecca, S. Sessa, H. Ishii, and A. Takanishi, *Development of new muscle contraction sensor to replace sEMG for using in muscles analysis fields*. 2014 36th Annual International Conference of the Ieee Engineering in Medicine and Biology Society (Embc), 2014: p. 6945-6948. URL: <http://ieeexplore.ieee.org/abstract/document/6945225/>
137. M. Zhang, Y.P. Zheng, and A.F. Mak, *Estimating the effective Young's modulus of soft tissues from indentation tests--nonlinear finite element analysis of effects of friction and large deformation*. *Med Eng Phys*, 1997. **19**(6): p. 512-7. DOI: 10.1016/S1350-4533(97)00017-9
138. D.R. Ruiz, M.E.Q. Escudero, D.R. Matoso, S.S. Montesdeoca, J.L. Reyna, Y.D. Guerra, G.P. Bautista, and J.M.G. Manso, *The Tensiomyography Used for Evaluating High Level Beach Volleyball Players*. *Revista Brasileira De Medicina Do Esporte*, 2012. **18**(2): p. 95-99. DOI: 10.1590/S1517-86922012000200006
139. A.M. Hunter, S.D.R. Galloway, I.J. Smith, J. Tallent, M. Ditroilo, M.M. Fairweather, and G. Howatson, *Assessment of eccentric exercise-induced muscle damage of the elbow flexors by tensiomyography*. *Journal of Electromyography and Kinesiology*, 2012. **22**(3): p. 334-341. DOI: 10.1016/j.jelekin.2012.01.009
140. K. Kersevan, V. Valencic, S. Djordjevic, and B. Simunic, *The muscle adaptation process as a result of pathological changes or specific training procedures*. *Cellular & Molecular Biology Letters*, 2002. **7**(2): p. 367-369. URL: <http://europepmc.org/abstract/med/12097988>
141. E. Alentorn-Geli, P. Alvarez-Diaz, S. Ramon, M. Marin, G. Steinbacher, M. Rius, R. Seijas, O. Ares, and R. Cugat, *Assessment of gastrocnemius tensiomyographic neuromuscular characteristics as risk factors for anterior cruciate ligament injury in male soccer players*. *Knee Surgery Sports Traumatology Arthroscopy*, 2015. **23**(9): p. 2502-2507. DOI: 10.1007/s00167-014-3007-4
142. S. Martin-Rodriguez, E. Alentorn-Geli, J. Tous-Fajardo, K. Samuelsson, M. Marin, P. Alvarez-Diaz, and R. Cugat, *Is tensiomyography a useful assessment tool in sports medicine?* *Knee Surgery Sports Traumatology Arthroscopy*, 2017. **25**(12): p. 3980-3981. DOI:

- 10.1007/s00167-017-4600-0
143. C.f.D. Control and Prevention, *Anthropometry procedures manual*. 2007, GA Atlanta. DOI:
144. N. Cameron, *Essential anthropometry: Baseline anthropometric methods for human biologists in laboratory and field situations*. *Am J Hum Biol*, 2013. **25**(3): p. 291-9. DOI: 10.1002/ajhb.22388
145. J. Tresignie, A. Scafoglieri, E. Cattrysse, and J.P. Clarys, *Cross-sectional content analysis of clinically applied circumferences*. *European Journal of Clinical Investigation*, 2012. **42**(9): p. 961-966. DOI: 10.1111/j.1365-2362.2012.02683.x
146. N. Miyatake, M. Miyachi, I. Tabata, N. Sakano, T. Hirao, and T. Numata, *Relationship between muscle strength and anthropometric, body composition parameters in Japanese adolescents*. *Health*, 2012. **04**(01): p. 1-5. DOI: 10.4236/health.2012.41001
147. K. Bouillard, A. Nordez, P.W. Hodges, C. Cornu, and F. Hug, *Evidence of changes in load sharing during isometric elbow flexion with ramped torque*. *J Biomech*, 2012. **45**(8): p. 1424-9. DOI: 10.1016/j.jbiomech.2012.02.020
148. L.A. Green and D.A. Gabriel, *Anthropometrics and electromyography as predictors for maximal voluntary isometric arm strength*. *Journal of Sport and Health Science*, 2012. **1**(2): p. 107-113. DOI: 10.1016/j.jshs.2012.05.004
149. F. Hussain, M.R. Abdul Kadir, A.H. Zulkifly, A. Sa'at, A.A. Aziz, G. Hossain, T. Kamarul, and A. Syahrom, *Anthropometric measurements of the human distal femur: a study of the adult Malay population*. *Biomed Res Int*, 2013. **2013**: p. 175056. DOI: 10.1155/2013/175056
150. B. Knechtle, P. Knechtle, I. Schulze, and G. Kohler, *Upper arm circumference is associated with race performance in ultra-endurance runners*. *Br J Sports Med*, 2008. **42**(4): p. 295-9; discussion 299. DOI: 10.1136/bjism.2007.038570
151. F. Lemma and P. Shetty, *Seasonal variations in the relationship between mid-upper arm circumference and maximum voluntary contraction among Ethiopian farmers*. *Eur J Clin Nutr*, 2009. **63**(4): p. 513-20. DOI: 10.1038/sj.ejcn.1602966
152. J.A.R. Cannan and H.S. Hu, *Automatic Circumference Measurement for Aiding in the Estimation of Maximum Voluntary Contraction (MVC) in EMG Systems*. *Intelligent Robotics and Applications, Pt I*, 2011. **7101**: p. 202-211. DOI: 10.1007/978-3-642-25486-4_21
153. K. Li, D.J. Hewson, J. Duchene, and J.Y. Hogrel, *Predicting maximal grip strength using hand circumference*. *Manual Therapy*, 2010. **15**(6): p. 579-585. DOI: 10.1016/j.math.2010.06.010
154. O. Heimbürger, A.R. Qureshi, W.S. Blaner, L. Berglund, and P. Stenvinkel, *Hand-grip muscle strength, lean body mass, and plasma proteins as markers of nutritional status in patients with chronic renal failure close to start of dialysis therapy*. *American Journal of Kidney Diseases*, 2000. **36**(6): p. 1213-1225. DOI: 10.1053/ajkd.2000.19837
155. L. Shu, X.M. Tao, and D.D. Feng, *A Wearable, Wireless Electronic Interface for Textile Sensors*. 2010 Ieee

- International Symposium on Circuits and Systems, 2010: p. 3104-3107. DOI: 10.1109/ISCAS.2010.5537973
156. L. Shu, T. Hua, Y.Y. Wang, Q.A. Li, D.D. Feng, and X.M. Tao, *In-Shoe Plantar Pressure Measurement and Analysis System Based on Fabric Pressure Sensing Array*. *Ieee Transactions on Information Technology in Biomedicine*, 2010. **14**(3): p. 767-775. DOI: 10.1109/Titb.2009.2038904
157. Y.Y. Wang, T. Hua, B. Zhu, Q. Li, W.J. Yi, and X.M. Tao, *Novel fabric pressure sensors: design, fabrication, and characterization*. *Smart Materials and Structures*, 2011. **20**(6): p. 065015. DOI: 10.1088/0964-1726/20/6/065015
158. W.J. Yi, Y.Y. Wang, G.F. Wang, and X.M. Tao, *Investigation of carbon black/silicone elastomer/dimethylsilicone oil composites for flexible strain sensors*. *Polymer Testing*, 2012. **31**(5): p. 677-684. DOI: 10.1016/j.polymertesting.2012.03.006
159. Y. Li, X.Y. Cheng, M.Y. Leung, J. Tsang, X.M. Tao, and C.W.M. Yuen, *A flexible strain sensor from polypyrrole-coated fabrics*. *Synthetic Metals*, 2005. **155**(1): p. 89-94. DOI: 10.1016/j.synthmet.2005.06.008
160. W.J. Yi, X.M. Tao, G.F. Wang, and Y.Y. Wang, *Performance specifications and evaluation methods for fabric strain sensors*. *Proceedings of 2009 International Textile Science and Technology Forum*, 2010: p. 75-80.
161. A.V. Hill, *The Heat of Shortening and the Dynamic Constants of Muscle*. *Proceedings of the Royal Society B: Biological Sciences*, 1938. **126**(843): p. 136-195. DOI: 10.1098/rspb.1938.0050
162. A.F. Huxley, *Muscle structure and theories of contraction*. *Prog Biophys Biophys Chem*, 1957. **7**: p. 255-318.
163. U. Stoecker, I.A. Telley, E. Stussi, and J. Denoth, *A multisegmental cross-bridge kinetics model of the myofibril*. *Journal of Theoretical Biology*, 2009. **259**(4): p. 714-726. DOI: 10.1016/j.jtbi.2009.03.032
164. K.S. Campbell, *Interactions between Connected Half-Sarcomeres Produce Emergent Mechanical Behavior in a Mathematical Model of Muscle*. *Plos Computational Biology*, 2009. **5**(11). DOI: ARTN e1000560 10.1371/journal.pcbi.1000560
165. M. Cadova, M. Vilimek, and M. Daniel, *A comparative study of muscle force estimates using Huxley's and Hill's muscle model*. *Computer Methods in Biomechanics and Biomedical Engineering*, 2014. **17**(4): p. 311-317. DOI: 10.1080/10255842.2012.683426
166. W.O. Williams, *Huxley's Model of Muscle Contraction with Compliance*. *Journal of Elasticity*, 2011. **105**(1-2): p. 365-380. DOI: 10.1007/s10659-011-9304-y
167. K.K. Lemaire, G.C. Baan, R.T. Jaspers, and A.J. van Soest, *Comparison of the validity of Hill and Huxley muscle-tendon complex models using experimental data obtained from rat m. soleus in situ*. *Journal of Experimental Biology*, 2016. **219**(7): p. 977-987. DOI: 10.1242/jeb.128280
168. J.I. Mechanick, L. Sun, and M. Zaidi, *Introduction to Molecular and Integrative Physiology of the*

- Musculoskeletal System*. Ann N Y Acad Sci, 2010. **1211**: p. 1-2. DOI: 10.1111/j.1749-6632.2010.05815.x
169. V.H. Frankel, *Biomechanics of the musculoskeletal system. Introduction*. Arch Surg, 1973. **107**(3): p. 405.
170. A.V. Hill, *First and last experiments in muscle mechanics*. 1970: Cambridge University Press. URL: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1005788/>
171. H.S. Gasser and A.V. Hill, *The dynamics of muscular contraction*. Proceedings of the Royal Society of London Series B-Containing Papers of a Biological Character, 1924. **96**(678): p. 398-436. DOI: 10.1098/rspb.1924.0035
172. F.E. Zajac, *How Musculotendon Architecture and Joint Geometry Affect the Capacity of Muscles to Move and Exert Force on Objects - a Review with Application to Arm and Forearm Tendon Transfer Design*. Journal of Hand Surgery-American Volume, 1992. **17A**(5): p. 799-804. DOI: 10.1016/0363-5023(92)90445-U
173. T.M. Winters, M. Takahashi, R.L. Lieber, and S.R. Ward, *Whole muscle length-tension relationships are accurately modeled as scaled sarcomeres in rabbit hindlimb muscles*. Journal of Biomechanics, 2011. **44**(1): p. 109-115. DOI: 10.1016/j.jbiomech.2010.08.033
174. J.M. Winters, in *Multiple muscle systems*. 1990, Springer. p. 69-93.
175. T. Komura, Y. Shinagawa, and T.L. Kunii, *Creating and retargetting motion by the musculoskeletal human body model*. Visual Computer, 2000. **16**(5): p. 254-270. DOI: 10.1007/s003719900065
176. D.T. Chen and D. Zeltzer, *Pump It up - Computer Animation of a Biomechanically Based Model of Muscle Using the Finite-Element Method*. Siggraph 92 : Conference Proceedings, 1992. **26**: p. 89-98.
177. D.F. Haeufle, M. Gunther, A. Bayer, and S. Schmitt, *Hill-type muscle model with serial damping and eccentric force-velocity relation*. J Biomech, 2014. **47**(6): p. 1531-6. DOI: 10.1016/j.jbiomech.2014.02.009
178. J.W. Ramsay, B.V. Hunter, and R.V. Gonzalez, *Muscle moment arm and normalized moment contributions as reference data for musculoskeletal elbow and wrist joint models*. J Biomech, 2009. **42**(4): p. 463-73. DOI: 10.1016/j.jbiomech.2008.11.035
179. A. Rahikainen, J. Avela, and M. Virnava, *Modeling the Force-Velocity Relationship in Arm*. World Journal of Mechanics, 2012. **02**(02): p. 90-97. DOI: 10.4236/wjm.2012.22011
180. E. Pennestri, R. Stefanelli, P.P. Valentini, and L. Vita, *Virtual musculoskeletal model for the biomechanical analysis of the upper limb*. J Biomech, 2007. **40**(6): p. 1350-61. DOI: 10.1016/j.jbiomech.2006.05.013
181. A. Erdemir, S. McLean, W. Herzog, and A.J. van den Bogert, *Model-based estimation of muscle forces exerted during movements*. Clin Biomech (Bristol, Avon), 2007. **22**(2): p. 131-54. DOI: 10.1016/j.clinbiomech.2006.09.005
182. P. Gerus, G. Rao, and E. Berton, *Subject-specific tendon-aponeurosis definition in Hill-type model predicts higher muscle forces in dynamic tasks*.

- PLoS One, 2012. **7**(8): p. e44406. DOI: 10.1371/journal.pone.0044406
183. F.C. Anderson and M.G. Pandy, *Individual muscle contributions to support in normal walking*. *Gait Posture*, 2003. **17**(2): p. 159-69. DOI: 10.1016/S0966-6362(02)00073-5
184. D.G. Thelen, F.C. Anderson, and S.L. Delp, *Generating dynamic simulations of movement using computed muscle control*. *Journal of Biomechanics*, 2003. **36**(3): p. 321-328. DOI: 10.1016/S0021-9290(02)00432-3
185. M.D. Fox, J.A. Reinbolt, S. Ounpuu, and S.L. Delp, *Mechanisms of improved knee flexion after rectus femoris transfer surgery*. *Journal of Biomechanics*, 2009. **42**(5): p. 614-619. DOI: 10.1016/j.jbiomech.2008.12.007
186. S.R. Hamner, A. Seth, and S.L. Delp, *Muscle contributions to propulsion and support during running*. *Journal of Biomechanics*, 2010. **43**(14): p. 2709-2716. DOI: 10.1016/j.jbiomech.2010.06.025
187. S.L. Delp, F.C. Anderson, A.S. Arnold, P. Loan, A. Habib, C.T. John, E. Guendelman, and D.G. Thelen, *OpenSim: open-source software to create and analyze dynamic Simulations of movement*. *Ieee Transactions on Biomedical Engineering*, 2007. **54**(11): p. 1940-1950. DOI: 10.1109/Tbme.2007.901024
188. A.L. Hall, C.L. Peterson, S.A. Kautz, and R.R. Neptune, *Relationships between muscle contributions to walking subtasks and functional walking status in persons with post-stroke hemiparesis*. *Clinical Biomechanics*, 2011. **26**(5): p. 509-515. DOI: 10.1016/j.clinbiomech.2010.12.010
189. C.L. Peterson, S.A. Kautz, and R.R. Neptune, *Muscle work is increased in pre-swing during hemiparetic walking*. *Clinical Biomechanics*, 2011. **26**(8): p. 859-866. DOI: 10.1016/j.clinbiomech.2011.04.010
190. T.K. Rupp, W. Ehlers, N. Karajan, M. Gunther, and S. Schmitt, *A forward dynamics simulation of human lumbar spine flexion predicting the load sharing of intervertebral discs, ligaments, and muscles*. *Biomechanics and Modeling in Mechanobiology*, 2015. **14**(5): p. 1081-1105. DOI: 10.1007/s10237-015-0656-2
191. A.A. Biewener, J.M. Wakeling, S.S. Lee, and A.S. Arnold, *Validation of Hill-Type Muscle Models in Relation to Neuromuscular Recruitment and Force-Velocity Properties: Predicting Patterns of In Vivo Muscle Force*. *Integrative and Comparative Biology*, 2014. **54**(6): p. 1072-1083. DOI: 10.1093/icb/icu070
192. H.E. Huxley, *Fifty years of muscle and the sliding filament hypothesis*. *European Journal of Biochemistry*, 2004. **271**(8): p. 1403-1415. DOI: 10.1111/j.1432-1033.2004.04044.x
193. V. Lombardi, G. Piazzesi, M. Reconditi, M. Linari, L. Lucii, A. Stewart, Y.B. Sun, P. Boesecke, T. Narayanan, T. Irving, and M. Irving, *X-ray diffraction studies of the contractile mechanism in single muscle fibres*. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences*, 2004. **359**(1452): p. 1883-1893. DOI: 10.1098/rstb.2004.1557
194. I.A. Telley, J. Denoth, and K.W. Ranatunga, *Inter-sarcomere dynamics in*

- muscle fibres - A neglected subject ?* Molecular and Cellular Aspects of Muscle Contraction, 2003. **538**: p. 481-500. URL: <https://www.ncbi.nlm.nih.gov/pubmed/15098693>
195. L. Tskhovrebova and J. Trinick, *Role of titin in vertebrate striated muscle*. Philosophical Transactions of the Royal Society B-Biological Sciences, 2002. **357**(1418): p. 199-206. DOI: 10.1098/rstb.2001.1028
196. A.F. Huxley, *Mechanics and models of the myosin motor*. Philosophical Transactions of the Royal Society of London Series B-Biological Sciences, 2000. **355**(1396): p. 433-440. DOI: 10.1098/rstb.2000.0584
197. M. Gunther and S. Schmitt, *A macroscopic ansatz to deduce the Hill relation*. Journal of Theoretical Biology, 2010. **263**(4): p. 407-418. DOI: 10.1016/j.jtbi.2009.12.027
198. D.F.B. Haeufle, M. Gunther, R. Blickhan, and S. Schmitt, *Proof of concept of an artificial muscle: theoretical model, numerical model, and hardware experiment*. 2011 Ieee International Conference on Rehabilitation Robotics (Icorr), 2011.
199. D.F.B. Haeufle, M. Gunther, R. Blickhan, and S. Schmitt, *Proof of concept: Model based bionic muscle with hyperbolic force-velocity relation*. Applied Bionics and Biomechanics, 2012. **9**(3): p. 267-274. DOI: 10.3233/Abb-2011-0052
200. S. Schmitt, D.F.B. Haeufle, R. Blickhan, and M. Gunther, *Nature as an engineer: one simple concept of a bio-inspired functional artificial muscle*. Bioinspiration & Biomimetics, 2012. **7**(3). DOI: Artn 036022 10.1088/1748-3182/7/3/036022
201. A. Rajagopal, C.L. Dembia, M.S. DeMers, D.D. Delp, J.L. Hicks, and S.L. Delp, *Full-Body Musculoskeletal Model for Muscle-Driven Simulation of Human Gait*. Ieee Transactions on Biomedical Engineering, 2016. **63**(10): p. 2068-2079. DOI: 10.1109/Tbme.2016.2586891
202. E.J. Perreault, C.J. Heckman, and T.G. Sandercock, *Hill muscle model errors during movement are greatest within the physiologically relevant range of motor unit firing rates*. Journal of Biomechanics, 2003. **36**(2): p. 211-218. DOI: 10.1016/S0021-9290(02)00332-9
203. S.S. Blemker and S.L. Delp, *Rectus femoris and vastus intermedius fiber excursions predicted by three-dimensional muscle models*. Journal of Biomechanics, 2006. **39**(8): p. 1383-1391. DOI: 10.1016/j.jbiomech.2005.04.012
204. M. Millard, T. Uchida, A. Seth, and S.L. Delp, *Flexing Computational Muscle: Modeling and Simulation of Musculotendon Dynamics*. Journal of Biomechanical Engineering-Transactions of the Asme, 2013. **135**(2). DOI: Artn 021005 10.1115/1.4023390
205. G.G. Handsfield, C.H. Meyer, J.M. Hart, M.F. Abel, and S.S. Blemker, *Relationships of 35 lower limb muscles to height and body mass quantified using MRI*. Journal of Biomechanics, 2014. **47**(3): p. 631-638. DOI: 10.1016/j.jbiomech.2013.12.002
206. O.M. Blake and J.M. Wakeling, *Early deactivation of slower muscle fibres at*

- high movement frequencies. *Journal of Experimental Biology*, 2014. **217**(19): p. 3528-3534. DOI: 10.1242/jeb.108266
207. J.M. Wakeling and T. Horn, *Neuromechanics of Muscle Synergies During Cycling*. *Journal of Neurophysiology*, 2009. **101**(2): p. 843-854. DOI: 10.1152/jn.90679.2008
208. S.S.M. Lee, M.D. Miara, A.S. Arnold, A.A. Biewener, and J.M. Wakeling, *Recruitment of faster motor units is associated with greater rates of fascicle strain and rapid changes in muscle force during locomotion*. *Journal of Experimental Biology*, 2013. **216**(2): p. 198-207. DOI: 10.1242/jeb.072637
209. M. Bernabei, J.H. van Dieen, G.C. Baan, and H. Maas, *Significant mechanical interactions at physiological lengths and relative positions of rat plantar flexors*. *Journal of Applied Physiology*, 2015. **118**(4): p. 427-436. DOI: 10.1152/jappphysiol.00703.2014
210. L. Reinhardt, T. Siebert, K. Leichsenring, R. Blickhan, and M. Bol, *Intermuscular pressure between synergistic muscles correlates with muscle force*. *Journal of Experimental Biology*, 2016. **219**(15): p. 2311-2319. DOI: 10.1242/jeb.135566
211. J.J. Knapik, J.E. Wright, R.H. Mawdsley, and J. Braun, *Isometric, isotonic, and isokinetic torque variations in four muscle groups through a range of joint motion*. *Physical Therapy*, 1983. **63**(6): p. 938-947. DOI: 10.1093/ptj/63.6.938
212. M.A. Anderson, J.H. Gieck, D. Perrin, A. Weltman, R. Rutt, and C. Denegar, *The relationships among isometric, isotonic, and isokinetic concentric and eccentric quadriceps and hamstring force and three components of athletic performance*. *Journal of Orthopaedic & Sports Physical Therapy*, 1991. **14**(3): p. 114-120. DOI: 10.2519/jospt.1991.14.3.114
213. L.M. Alegre, A. Ferri-Morales, R. Rodriguez-Casares, and X. Aguado, *Effects of isometric training on the knee extensor moment-angle relationship and vastus lateralis muscle architecture*. *European Journal of Applied Physiology*, 2014. **114**(11): p. 2437-2446. DOI: 10.1007/s00421-014-2967-x
214. M. Noorkoiv, K. Nosaka, and A.J. Blazevich, *Effects of isometric quadriceps strength training at different muscle lengths on dynamic torque production*. *Journal of Sports Sciences*, 2015. **33**(18): p. 1952-1961. DOI: 10.1080/02640414.2015.1020843
215. R.C. Melo, A.C.M. Takahashi, R.J. Quiterio, T.F. Salvini, and A.M. Catai, *Eccentric Torque-Producing Capacity Is Influenced by Muscle Length in Older Healthy Adults*. *Journal of Strength and Conditioning Research*, 2016. **30**(1): p. 259-266. DOI: 10.1519/Jsc.0000000000001047
216. N.A. Tillin, M.T.G. Pain, and J.P. Folland, *Contraction type influences the human ability to use the available torque capacity of skeletal muscle during explosive efforts*. *Proceedings of the Royal Society B-Biological Sciences*, 2012. **279**(1736): p. 2106-2115. DOI: 10.1098/rspb.2011.2109
217. O.S. Andersen, *50-year anniversary of sliding filament*. *Journal of General Physiology*, 2004. **123**(6): p. 629-629.

- DOI: 10.1085/jgp.200409079
218. S. Nimphius, M.R. McGuigan, and R.U. Newton, *Changes in Muscle Architecture and Performance during a Competitive Season in Female Softball Players*. *Journal of Strength and Conditioning Research*, 2012. **26**(10): p. 2655-2666. DOI: 10.1519/JSC.0b013e318269f81e
219. B.M. Baroni, J.M. Geremia, R. Rodrigues, R.D. Franke, K. Karamanidis, and M.A. Vaz, *Muscle Architecture Adaptations to Knee Extensor Eccentric Training: Rectus Femoris Vs. Vastus Lateralis*. *Muscle & Nerve*, 2013. **48**(4): p. 498-506. DOI: 10.1002/mus.23785
220. T.C. Scanlon, M.S. Fragala, J.R. Stout, N.S. Emerson, K.S. Beyer, L.P. Oliveira, and J.R. Hoffman, *Muscle Architecture and Strength: Adaptations to Short- Term Resistance Training in Older Adults*. *Muscle & Nerve*, 2014. **49**(4): p. 584-592. DOI: 10.1002/mus.23969
221. C. Rode, T. Siebert, A. Tomalka, and R. Blickhan, *Myosin filament sliding through the Z-disc relates striated muscle fibre structure to function*. *Proceedings of the Royal Society B-Biological Sciences*, 2016. **283**(1826). DOI: Artn 20153030 10.1098/Rspb.2015.3030
222. T. Driss, H. Vandewalle, J.M. Le Chevalier, and H. Monod, *Force-velocity relationship on a cycle ergometer and knee-extensor strength indices*. *Can J Appl Physiol*, 2002. **27**(3): p. 250-62. DOI: 10.1139/h02-015
223. P.T. Nikolaidis, *Age- and Sex-Related Differences in Force-Velocity Characteristics of Upper and Lower Limbs of Competitive Adolescent Swimmers*. *Journal of Human Kinetics*, 2012. **32**: p. 87-95. DOI: 10.2478/v10078-012-0026-4
224. H. Jaafar, E. Attiogbe, M. Rouis, H. Vandewalle, and T. Driss, *Reliability of Force-Velocity Tests in Cycling and Cranking Exercises in Men and Women*. *Biomed Res Int*, 2015. **2015**: p. 954780. DOI: 10.1155/2015/954780
225. I. Cuk, M. Markovic, A. Nedeljkovic, D. Ugarkovic, M. Kukolj, and S. Jaric, *Force-velocity relationship of leg extensors obtained from loaded and unloaded vertical jumps*. *European Journal of Applied Physiology*, 2014. **114**(8): p. 1703-1714. DOI: 10.1007/s00421-014-2901-2
226. D. Feeney, S.J. Stanhope, T.W. Kaminski, A. Machi, and S. Jaric, *Loaded Vertical Jumping: Force-Velocity Relationship, Work, and Power*. *J Appl Biomech*, 2016. **32**(2): p. 120-7. DOI: 10.1123/jab.2015-0136
227. A. Garcia-Ramos, S. Jaric, P. Padijal, and B. Ferlic, *Force-Velocity Relationship of Upper Body Muscles: Traditional Versus Ballistic Bench Press*. *J Appl Biomech*, 2016. **32**(2): p. 178-85. DOI: 10.1123/jab.2015-0162
228. G. Rabita, S. Dorel, J. Slawinski, E. Saez-de-Villarreal, A. Couturier, P. Samozino, and J.B. Morin, *Sprint mechanics in world-class athletes: a new insight into the limits of human locomotion*. *Scand J Med Sci Sports*, 2015. **25**(5): p. 583-94. DOI: 10.1111/sms.12389
229. H. Vandewalle, G. Peres, J. Heller, J. Panel, and H. Monod, *Force-velocity*

- relationship and maximal power on a cycle ergometer. Correlation with the height of a vertical jump.* *Eur J Appl Physiol*, 1987. **56**(6): p. 650-6. URL: <https://link.springer.com/article/10.1007/BF00424805>
230. A. Jaskolska, P. Goossens, B. Veenstra, A. Jaskolski, and J.S. Skinner, *Comparison of treadmill and cycle ergometer measurements of force-velocity relationships and power output.* *International Journal of Sports Medicine*, 1999. **20**(3): p. 192-197. DOI: 10.1055/s-2007-971116
231. J.B. Morin, P. Samozino, R. Bonnefoy, P. Edouard, and A. Belli, *Direct measurement of power during one single sprint on treadmill.* *Journal of Biomechanics*, 2010. **43**(10): p. 1970-1975. DOI: 10.1016/j.jbiomech.2010.03.012
232. J. Yamauchi, C. Mishima, S. Nakayama, and N. Ishii, *Force-velocity, force-power relationships of bilateral and unilateral leg multi-joint movements in young and elderly women.* *Journal of Biomechanics*, 2009. **42**(13): p. 2151-7. DOI: 10.1016/j.jbiomech.2009.05.032
233. P. Samozino, E. Rejc, P.E. Di Prampero, A. Belli, and J.B. Morin, *Optimal force-velocity profile in ballistic movements--altius: citius or fortius?* *Medicine & Science in Sports & Exercise*, 2012. **44**(2): p. 313-22. DOI: 10.1249/MSS.0b013e31822d757a
234. P. Samozino, E. Rejc, P.E. di Prampero, A. Belli, and J.B. Morin, *Force-velocity properties' contribution to bilateral deficit during ballistic push-off.* *Medicine & Science in Sports & Exercise*, 2014. **46**(1): p. 107-14. DOI: 10.1249/MSS.0b013e3182a124fb
235. S. Sreckovic, I. Cuk, S. Djuric, A. Nedeljkovic, D. Mirkov, and S. Jaric, *Evaluation of force-velocity and power-velocity relationship of arm muscles.* *European Journal of Applied Physiology*, 2015. **115**(8): p. 1779-1787. DOI: 10.1007/s00421-015-3165-1
236. P. Samozino, P. Edouard, S. Sangnier, M. Brughelli, P. Gimenez, and J.B. Morin, *Force-Velocity Profile: Imbalance Determination and Effect on Lower Limb Ballistic Performance.* *International Journal of Sports Medicine*, 2014. **35**(6): p. 505-510. DOI: 10.1055/s-0033-1354382
237. P. Jimenez-Reyes, P. Samozino, V. Cuadrado-Penafiel, F. Conceicao, J.J. Gonzalez-Badillo, and J.B. Morin, *Effect of countermovement on power-force-velocity profile.* *European Journal of Applied Physiology*, 2014. **114**(11): p. 2281-2288. DOI: 10.1007/s00421-014-2947-1
238. H. Hauraix, S. Dorel, G. Rabita, G. Guilhem, and A. Nordez, *Muscle fascicle shortening behaviour of vastus lateralis during a maximal force-velocity test.* *European Journal of Applied Physiology*, 2017. **117**(2): p. 289-299. DOI: 10.1007/s00421-016-3518-4
239. S. Jaric, *Force-velocity Relationship of Muscles Performing Multi-joint Maximum Performance Tasks.* *International Journal of Sports Medicine*, 2015. **36**(9): p. 699-704. DOI: 10.1055/s-0035-1547283
240. M.F. Bobbert, *Why is the force-velocity relationship in leg press tasks quasi-linear rather than hyperbolic?* *Journal*

- of Applied Physiology, 2012. **112**(12): p. 1975-1983. DOI: 10.1152/jappphysiol.00787.2011
241. J.M. Sheppard, S. Cormack, K.L. Taylor, M.R. McGuigan, and R.U. Newton, *Assessing the Force-Velocity Characteristics of the Leg Extensors in Well-Trained Athletes: The Incremental Load Power Profile*. *Journal of Strength and Conditioning Research*, 2008. **22**(4): p. 1320-1326. DOI: 10.1519/JSC.0b013e31816d671b
242. A. García-Ramos, S. Jaric, P. Padial, and B. Feriche, *Force-velocity relationship of upper body muscles: traditional versus ballistic bench press*. *Journal of applied biomechanics*, 2016. **32**(2): p. 178-185. DOI: 10.1123/jab.2015-0162
243. R.C. Sprague, J.C. Martin, C.J. Davidson, and R.P. Farrar, *Force-velocity and power-velocity relationships during maximal short-term rowing ergometry*. *Medicine and Science in Sports and Exercise*, 2007. **39**(2): p. 358-364. DOI: 10.1249/01.mss.0000241653.37876.73
244. D. Hahn, W. Herzog, and A. Schwartz, *Interdependence of torque, joint angle, angular velocity and muscle action during human multi-joint leg extension*. *European Journal of Applied Physiology*, 2014. **114**(8): p. 1691-1702. DOI: 10.1007/s00421-014-2899-5
245. D.G. Behm and D.G. Sale, *Intended Rather Than Actual Movement Velocity Determines Velocity-Specific Training Response*. *Journal of Applied Physiology*, 1993. **74**(1): p. 359-368. DOI: 10.1152/jappl.1993.74.1.359
246. D.M. Frost, S. Bronson, J.B. Cronin, and R.U. Newton, *Changes in maximal strength, velocity, and power after 8 weeks of training with pneumatic or free weight resistance*. *The Journal of Strength & Conditioning Research*, 2016. **30**(4): p. 934-944. DOI: 10.1519/JSC.0000000000001179
247. M. Riviere, L. Louit, A. Strokosch, and L.B. Seitz, *Variable Resistance Training Promotes Greater Strength and Power Adaptations Than Traditional Resistance Training in Elite Youth Rugby League Players*. *Journal of Strength and Conditioning Research*, 2017. **31**(4): p. 947-955. DOI: 10.1519/Jsc.0000000000001574
248. R. Formon, L.E. Ford, and E.H. Sonnenblick, *Effect of muscle length on the force-velocity relationship of tetanized cardiac muscle*. *Circulation research*, 1972. **31**(2): p. 195-206. DOI: 10.1161/01.RES.31.2.195
249. C.d. Ruiter, W. Didden, D. Jones, and A.d. Haan, *The force-velocity relationship of human adductor pollicis muscle during stretch and the effects of fatigue*. *The Journal of Physiology*, 2000. **526**(3): p. 671-681. DOI: 10.1111/j.1469-7793.2000.00671.x
250. S. Sprager and M.B. Juric, *Inertial Sensor-Based Gait Recognition: A Review*. *Sensors (Basel)*, 2015. **15**(9): p. 22089-127. DOI: 10.3390/s150922089
251. N.U. Ahamed, K. Sundaraj, B. Ahmad, M. Rahman, M.A. Ali, M.A. Islam, and R. Palaniappan, *Rehabilitation systems for physically disabled patients: A brief review of sensor-based computerised signal-monitoring systems*. *Biomedical Research-India*, 2013. **24**(3): p. 370-376.

252. A. Ali, K. Sundaraj, B. Ahmad, N. Ahamed, and A. Islam, *Gait disorder rehabilitation using vision and non-vision based sensors: A systematic review*. *Bosnian Journal of Basic Medical Sciences*, 2012. **12**(3): p. 193-202. DOI: 10.17305/bjbms.2012.2484
253. A. Reiss, G. Hendeby, G. Bleser, and D. Stricker, *Activity Recognition Using Biomechanical Model Based Pose Estimation*. *Smart Sensing and Context*, 2010. **6446**: p. 42-55. DOI: 10.1007/978-3-642-16982-3_4
254. Y.-C. Du, C.-B. Shih, S.-C. Fan, H.-T. Lin, and P.-J. Chen, *An IMU-compensated skeletal tracking system using Kinect for the upper limb*. *Microsystem Technologies*, 2018. DOI: 10.1007/s00542-018-3769-6
255. A. Tognetti, F. Lorussi, N. Carbonaro, and D. de Rossi, *Wearable Goniometer and Accelerometer Sensory Fusion for Knee Joint Angle Measurement in Daily Life*. *Sensors*, 2015. **15**(11): p. 28435-28455. DOI: 10.3390/s151128435
256. C. Chen, R. Jafari, and N. Kehtarnavaz, *A survey of depth and inertial sensor fusion for human action recognition*. *Multimedia Tools and Applications*, 2017. **76**(3): p. 4405-4425. DOI: 10.1007/s11042-015-3177-1
257. D. Roetenberg, H.J. Luinge, C.T. Baten, and P.H. Veltink, *Compensation of magnetic disturbances improves inertial and magnetic sensing of human body segment orientation*. *IEEE Trans Neural Syst Rehabil Eng*, 2005. **13**(3): p. 395-405. DOI: 10.1109/TNSRE.2005.847353
258. A.M. Sabatini, *Estimating three-dimensional orientation of human body parts by inertial/magnetic sensing*. *Sensors (Basel)*, 2011. **11**(2): p. 1489-525. DOI: 10.3390/s110201489
259. H. Dejnabadi, B.M. Jolles, and K. Aminian, *A new approach to accurate measurement of uniaxial joint angles based on a combination of accelerometers and gyroscopes*. *IEEE Trans Biomed Eng*, 2005. **52**(8): p. 1478-84. DOI: 10.1109/TBME.2005.851475
260. D. Roetenberg, P.J. Slycke, and P.H. Veltink, *Ambulatory position and orientation tracking fusing magnetic and inertial sensing*. *IEEE Trans Biomed Eng*, 2007. **54**(5): p. 883-90. DOI: 10.1109/TBME.2006.889184
261. H.J. Luinge, P.H. Veltink, and C.T. Baten, *Ambulatory measurement of arm orientation*. *J Biomech*, 2007. **40**(1): p. 78-85. DOI: 10.1016/j.jbiomech.2005.11.011
262. M. Khademi, H.M. Hondori, L. Dodakian, S. Cramer, and C.V. Lopes, *Comparing "pick and place" task in spatial Augmented Reality versus non-immersive Virtual Reality for rehabilitation setting*. *Conf Proc IEEE Eng Med Biol Soc*, 2013. **2013**: p. 4613-6. DOI: 10.1109/EMBC.2013.6610575
263. Y.Q. Tao and H.S. Hu, *Colour based human motion tracking for home-based rehabilitation*. *2004 Ieee International Conference on Systems, Man & Cybernetics, Vols 1-7*, 2004: p. 773-778.
264. A. Mobini, S. Behzadipour, and M. Saadat Foumani, *Accuracy of Kinect's skeleton tracking for upper body rehabilitation applications*. *Disabil Rehabil Assist Technol*, 2014. **9**(4): p.

- 344-52. DOI: 10.3109/17483107.2013.805825
265. R. Paradiso and D. De Rossi. *Advances in textile sensing and actuation for e-textile applications*. in *Engineering in Medicine and Biology Society, 2008. EMBS 2008. 30th Annual International Conference of the IEEE*. 2008. IEEE. URL: <http://ieeexplore.ieee.org/abstract/document/4649993/>
266. M. Hamed, R. Forchheimer, and O. Inganas, *Towards woven logic from organic electronic fibres*. *Nature Materials*, 2007. **6**(5): p. 357-362. DOI: 10.1038/nmat1884
267. T. Giorgino, P. Tormene, F. Lorussi, D. De Rossi, and S. Quaglini, *Sensor evaluation for wearable strain gauges in neurological rehabilitation*. *IEEE Trans Neural Syst Rehabil Eng*, 2009. **17**(4): p. 409-15. DOI: 10.1109/TNSRE.2009.2019584
268. P. Tormene, M. Bartolo, A.M. De Nunzio, F. Fecchio, S. Quaglini, C. Tassorelli, and G. Sandrini, *Estimation of human trunk movements by wearable strain sensors and improvement of sensor's placement on intelligent biomedical clothes*. *Biomed Eng Online*, 2012. **11**: p. 95. DOI: 10.1186/1475-925X-11-95
269. A. Tognetti, F. Lorussi, G.D. Mura, N. Carbonaro, M. Pacelli, R. Paradiso, and D.D. Rossi, *New generation of wearable goniometers for motion capture systems*. *J Neuroeng Rehabil*, 2014. **11**: p. 56. DOI: 10.1186/1743-0003-11-56
270. L. Shu, T. Hua, Y. Wang, Q. Qiao Li, D.D. Feng, and X. Tao, *In-shoe plantar pressure measurement and analysis system based on fabric pressure sensing array*. *IEEE Trans Inf Technol Biomed*, 2010. **14**(3): p. 767-75. DOI: 10.1109/TITB.2009.2038904
271. N. Carbonaro, G. Dalle Mura, F. Lorussi, R. Paradiso, D. De Rossi, and A. Tognetti, *Exploiting wearable goniometer technology for motion sensing gloves*. *IEEE J Biomed Health Inform*, 2014. **18**(6): p. 1788-95. DOI: 10.1109/JBHI.2014.2324293
272. A. Tognetti, F. Lorussi, N. Carbonaro, and D. de Rossi, *Wearable Goniometer and Accelerometer Sensory Fusion for Knee Joint Angle Measurement in Daily Life*. *Sensors (Basel)*, 2015. **15**(11): p. 28435-55. DOI: 10.3390/s151128435
273. S.J. Preece, L.P.J. Kenney, M.J. Major, T. Dias, E. Lay, and B.T. Fernandes, *Automatic identification of gait events using an instrumented sock*. *Journal of Neuroengineering and Rehabilitation*, 2011. **8**. DOI: Artn 32 10.1186/1743-0003-8-32
274. T.W. Shyr, J.W. Shie, C.H. Jiang, and J.J. Li, *A textile-based wearable sensing device designed for monitoring the flexion angle of elbow and knee movements*. *Sensors (Basel)*, 2014. **14**(3): p. 4050-9. DOI: 10.3390/s140304050
275. B.J. Munro, T.E. Campbell, G.G. Wallace, and J.R. Steele, *The intelligent knee sleeve: A wearable biofeedback device*. *Sensors and Actuators B-Chemical*, 2008. **131**(2): p. 541-547. DOI: 10.1016/j.snb.2007.12.041
276. A. Tognetti, F. Lorussi, R. Bartalesi, S. Quaglini, M. Tesconi, G. Zupone, and D. De Rossi, *Wearable kinesthetic*

- system for capturing and classifying upper limb gesture in post-stroke rehabilitation.* J Neuroeng Rehabil, 2005. **2**(1): p. 8. DOI: 10.1186/1743-0003-2-8
277. T. Giorgino, P. Tormene, and S. Quaglini, *A multivariate time-warping based classifier for gesture recognition with wearable strain sensors.* 2007 Annual International Conference of the Ieee Engineering in Medicine and Biology Society, Vols 1-16, 2007: p. 4903-4906. DOI: 10.1109/Iembs.2007.4353439
278. R. McLaren, F. Joseph, C. Baguley, and D. Taylor, *A review of e-textiles in neurological rehabilitation: How close are we?* J Neuroeng Rehabil, 2016. **13**(1): p. 59. DOI: 10.1186/s12984-016-0167-0
279. D. Galeano, F. Brunetti, D. Torricelli, S. Piazza, and J.L. Pons, *A Tool for Balance Control Training Using Muscle Synergies and Multimodal Interfaces.* Biomed Research International, 2014. DOI: Artn 565370 10.1155/2014/565370
280. Y. Ma, S.Q. Xie, and Y.X. Zhang, *A Patient-Specific EMG-Driven Musculoskeletal Model for Improving the Effectiveness of Robotic Neurorehabilitation.* Intelligent Robotics and Applications, Icira 2014, Pt I, 2014. **8917**: p. 390-401.
281. Q.S. Ai, B. Ding, Q. Liu, and W. Meng, *A Subject-Specific EMG-Driven Musculoskeletal Model for Applications in Lower-Limb Rehabilitation Robotics.* International Journal of Humanoid Robotics, 2016. **13**(3). DOI: Artn 1650005 10.1142/S0219843616500055
282. L. Peng, Z.G. Hou, L.C. Luo, L. Peng, W.Q. Wang, and L. Cheng, *An sEMG-Driven Neuromusculoskeletal Model of Upper Limb for Rehabilitation Robot Control.* 2016 Ieee International Conference on Robotics and Biomimetics (Robio), 2016: p. 1486-1491.
283. L.M. Vaca Benitez, M. Tabie, N. Will, S. Schmidt, M. Jordan, and E.A. Kirchner, *Exoskeleton technology in rehabilitation: Towards an EMG-based orthosis system for upper limb neuromotor rehabilitation.* Journal of Robotics, 2013. **2013**. DOI: 10.1155/2013/610589
284. K. Manal, K. Gravare-Silbernagel, and T.S. Buchanan, *A real-time EMG-driven musculoskeletal model of the ankle.* Multibody System Dynamics, 2012. **28**(1-2): p. 169-180. DOI: 10.1007/s11044-011-9285-4
285. T.K.K. Koo and A.F.T. Mak, *Feasibility of using EMG driven neuromusculoskeletal model for prediction of dynamic movement of the elbow.* Journal of Electromyography and Kinesiology, 2005. **15**(1): p. 12-26. DOI: 10.1016/j.jelekin.2004.06.007
286. J.W.L. Pau, S.S.Q. Xie, and A.J. Pullan, *Neuromuscular Interfacing: Establishing an EMG-Driven Model for the Human Elbow Joint.* Ieee Transactions on Biomedical Engineering, 2012. **59**(9): p. 2586-2593. DOI: 10.1109/Tbme.2012.2206389
287. H. Wagner, K. Bostrom, and B. Rinke, *Predicting isometric force from muscular activation using a physiologically inspired model.* Biomechanics and Modeling in

- Mechanobiology, 2011. **10**(6): p. 955-961. DOI: 10.1007/s10237-011-0286-2
288. N. Louis and P. Gorce, *Upper limb muscle forces during a simple reach-to-grasp movement: a comparative study*. *Medical & Biological Engineering & Computing*, 2009. **47**(11): p. 1173-1179. DOI: 10.1007/s11517-009-0530-4
289. S.S.M. Lee, A.S. Arnold, M.D. Miara, A.A. Biewener, and J.M. Wakeling, *Accuracy of gastrocnemius muscles forces in walking and running goats predicted by one-element and two-element Hill-type models*. *Journal of Biomechanics*, 2013. **46**(13): p. 2288-2295. DOI: 10.1016/j.jbiomech.2013.06.001