

REVIEW ARTICLE

Angiogenic and lymphangiogenic factors in wound healing

Authors

Kazuhiko Shimizu and Fumi Sato

Affiliations

Department of Anatomy, School of Medicine, Toho University, Tokyo, Japan; 5-21-16 Omori-Nishi, Ota-ku, Tokyo 143-8540, Japan.

Telephone Number: +81-3-3762-4151, Fax Number: +81-3-5493-5411

Corresponding Author:

Kazuhiko Shimizu

Department of Anatomy, School of Medicine, Toho University,
5-21-16 Omori-Nishi, Ota-ku, Tokyo 143-8540, Japan.

Email: kazuhiko.shimizu@med.toho-u.ac.jp

Telephone Number: +81-3-3762-4151, Fax Number: +81-3-5493-5411

ABSTRACT

When any tissue is damaged, multiple cells and tissues work towards the repair of the wounded site. Blood and lymphatic vessels are particularly important for the regeneration and healing of tissues. Angiogenesis is the process by which new blood vessels are formed. Angiogenesis is induced by angiogenic factors such as vascular endothelial growth factor (VEGF)-A which plays an important role in the repair of the damaged site. VEGF-A is expressed by macrophages, but pericytes also promote vascularization by expressing VEGF-A. In addition to VEGF-A, wound-related macrophages express tumor necrosis factor- α , Platelet-derived growth factor-bb, Interleukin (IL)-1, IL-6, and transforming growth factor β , and act on other cells. Pericytes change properties depending on the stage of the wound. For lymphangiogenesis, the expression of VEGF-C or -D, which are lymphatic endothelial growth factors, is the most important. Lymphatic vessel endothelial hyaluronan receptor-1-positive macrophages, which appear in the stroma, are also actively involved in lymphangiogenesis. On the other hand, podoplanin-positive myofibroblasts are indirectly involved in wound healing by being affecting in leukocyte migration as an “extravascular pathway”.

Keywords:: angiogenesis, lymphangiogenesis, wound healing, LYVE-1, podoplanin
dothelial dysfunction, ambulatory blood pressure, central blood pressure, prehypertension

Introduction

Wound healing is a complex process that combines multiple processes and has an inflammatory phase, proliferative phase, and remodeling phase. During the inflammatory phase, platelets aggregate and block the wound, and various cytokines and cell growth factors are secreted, causing several cells to infiltrate the wound. During the growth phase, epidermal cells, fibroblasts, and vascular endothelial cells proliferate, forming granulation tissue. Scars form during the remodeling phase. Blood and lymphatic vessels play a vital role in the wound healing processes. In this review document, we will discuss angiogenesis and lymphangiogenesis, and the various factors involved in wound healing.

1. Angiogenic factors and their expression

Angiogenesis is a complex system that is highly controlled in the body, and requires various factors to act in conjunction to be effective. Factors pertaining to angiogenesis in wound-sites include vascular endothelial growth factor (VEGF), angiopoietin (ANGPT), fibroblast growth factor (FGF), and transforming growth factor β (TGF- β). Co-expression of these growth factors is essential for angiogenesis. Keratinocytes, macrophages, fibroblasts, and pericytes (PCs) are among the cells that express the angiogenic factor. The interaction of these cells with vascular endothelial cells makes the process of wound healing intricate.

1.1 VEGF-A

VEGF has been isolated as a substance with two properties; it is a growth factor for vascular endothelial cells and also acts as a vascular permeability factor, known as VEGF-A¹⁻³. VEGF-A has at least six known isoforms namely VEGF 121, 145, 165, 183, 189, and 206^{4,5}. VEGF-A is the most

important angiogenic factor involved in wound healing. VEGF-A is a downstream protein of Hypoxia-inducible factor 1-alpha, and its expression is induced by hypoxia in wounds⁶. VEGF binds to vascular endothelial growth factor receptor (VEGFR)-1 (encoded by the FLT-1 gene) and/or VEGFR-2 (encoded by the Flk-1 gene), expressed in vascular endothelial cells, thereby promoting endothelial cell migration and proliferation. This induces angiogenesis and ensures the survival of the cells.

1.2 ANGPT

ANGPT belongs to the VEGF family and mainly regulates the adhesion of PCs to vascular endothelial cells. Both the agonist ANGPT-1, and the antagonist ANGPT-2, bind to the endothelial cell receptor, TIE2. ANGPT-1 is produced by PCs, and ANGPT-2 is produced by endothelial cells. ANGPT-1 attaches PCs to vascular endothelial cells to form a mature blood vessels, whereas ANGPT-2 releases PCs⁷.

1.3 FGFs

Many types of FGFs have been reported (more than 23 homologs) of which FGF1 and FGF2 are particularly important for angiogenesis. FGF1 and FGF2 not only stimulate the proliferation of vascular endothelial cells, but also organize the vascular lumen structure⁸. Additionally, FGFs play an important role in the formation of granulation tissue in the process of wound healing.

1.4 TGF- β

TGF- β is a multifunctional cytokine with at least three isoforms (TGF- β 1, - β 2, and - β 3). TGF- β controls cell growth, proliferation, differentiation, and apoptosis for various cell types. TGF- β also has many signaling pathways and promotes angiogenesis during wound healing⁹.

1.5 Macrophages

Macrophages play an important role in the inflammatory phase of tissue repair¹⁰. Due

to their dynamic plasticity, macrophages can mediate both tissue destruction and repair¹¹. Macrophages have many functions, defined by an intricate subset of cell-derived cytokines. Wound associated macrophages play an important role in angiogenesis. They not only express VEGF, but also promote VEGF production in keratinocytes and fibroblasts via tumor necrosis factor (TNF)- α , Platelet-derived growth factor (PDGF)-bb, Interleukin (IL)-1, IL-6, and TGF- β ¹².

1.6 Keratinocytes

Increased amounts of VEGF in the wound area is as a result of migratory keratinocytes and macrophages in the granulation tissue¹³. Keratinocyte VEGF expression is indirectly promoted by macrophages expressing TNF- α and TGF- β ¹⁴.

1.7 Fibroblasts

Fibroblasts are stimulated by TGF- β to produce connective tissue growth factor (CTGF)¹⁵. CTGF induces connective tissue proliferation, vascular endothelial cell migration, and tube formation^{16,17}.

1.8 PCs

PCs, a type of wall cell in blood vessels, adhere to the outer circumference of capillaries and venules. Unlike vascular smooth muscle, PCs wrap around the basement membrane together with vascular endothelial cells and are in direct contact with endothelial cells. One of their functions is to stabilize blood vessels, but the mechanism by which this is achieved is still quite unclear¹⁸. It has been suggested that the function of PCs during angiogenesis is to suppress endothelial cell proliferation and to stabilize the vessel wall¹⁹⁻²¹. However, some studies have suggested that PCs can induce endothelial cell proliferation and sprouting during angiogenesis²²⁻³⁰. In addition, PCs exhibit heterogeneity at each stage of wound healing, and express VEGF-A to promote proliferation in the neovascular tip and surrounding regions of the endothelial cell³¹. The zebrafish model suggests that the

wound activates PCs and induces angiogenesis³².

2. Lymphangiogenic factors and their expression

Lymphatic vessels play an important role in tissue fluid collection and immunity transfer pathways. However, although the mechanism of lymphatic vessel formation and the *in vivo* regulatory factors affecting them have recently been studied, it still remains unclear.

The formation of lymphatic vessels during inflammation and wound healing, has also been reported and several influential factors identified. However, lymphatic vessels have not been studied as extensively as blood vessels. VEGF-C and -D are lymphatic endothelial growth factors, Prox-1 is a lymphatic endothelial cell master factor, and Foxc2 is important for lymphatic valve formation. Furthermore, lymphatic markers VEGFR3, LYVE-1, and podoplanin are also considered key. In particular, a large number of cells expressing lymphatic markers appear at the wound site, and the relationship between these cells and lymphangiogenesis may be important.

2.1 VEGF-C and -D

VEGF-C and -D, ligands for VEGFR3/Flt4, play an important role in the proliferation and migration of lymphatic endothelial cells³³. VEGF-C and -D bind to VEGFR2³⁴⁻³⁶, and VEGFR2 and VEGFR3 can also form heterodimers^{37,38}. In wound healing, coordinated signaling of VEGFR2 and VEGFR3 is thus a key factor³⁹.

2.2 Prospero homeobox protein 1 (PROX1)

PROX1 is an essential master transcription factor for the development and maintenance of lymphatic vessels⁴⁰⁻⁴². Binding of VEGFC, D, and VEGFR3 activates PROX1. In a corneal injury model, PROX1 was suppressed by microRNA miR-466, suggesting that lymphangiogenesis could be suppressed⁴³. The functionality of PROX1

in wound healing is expected to develop in the future.

2.3 Forkhead box protein (FOX)C2

FOXC2, is highly expressed in the fetus and adult lymphatic vessels^{44,45}. In the fetus, FOXC2 is involved in the formation of lymphatic vessels in coordination with

FOXC1⁴⁶. In mature lymphatic vessels, FOXC2 is downregulated, resulting in decreased expression levels of PROX1, VEGFR-3, and LYVE-1⁴⁷. However, the expression and function of FOXC2 during lymphangiogenesis in wounds is still unknown.

Figure 1: Double immunofluorostaining in the wound section of the skin in a mouse-model (C57BL / 6N, 8-week-old male).

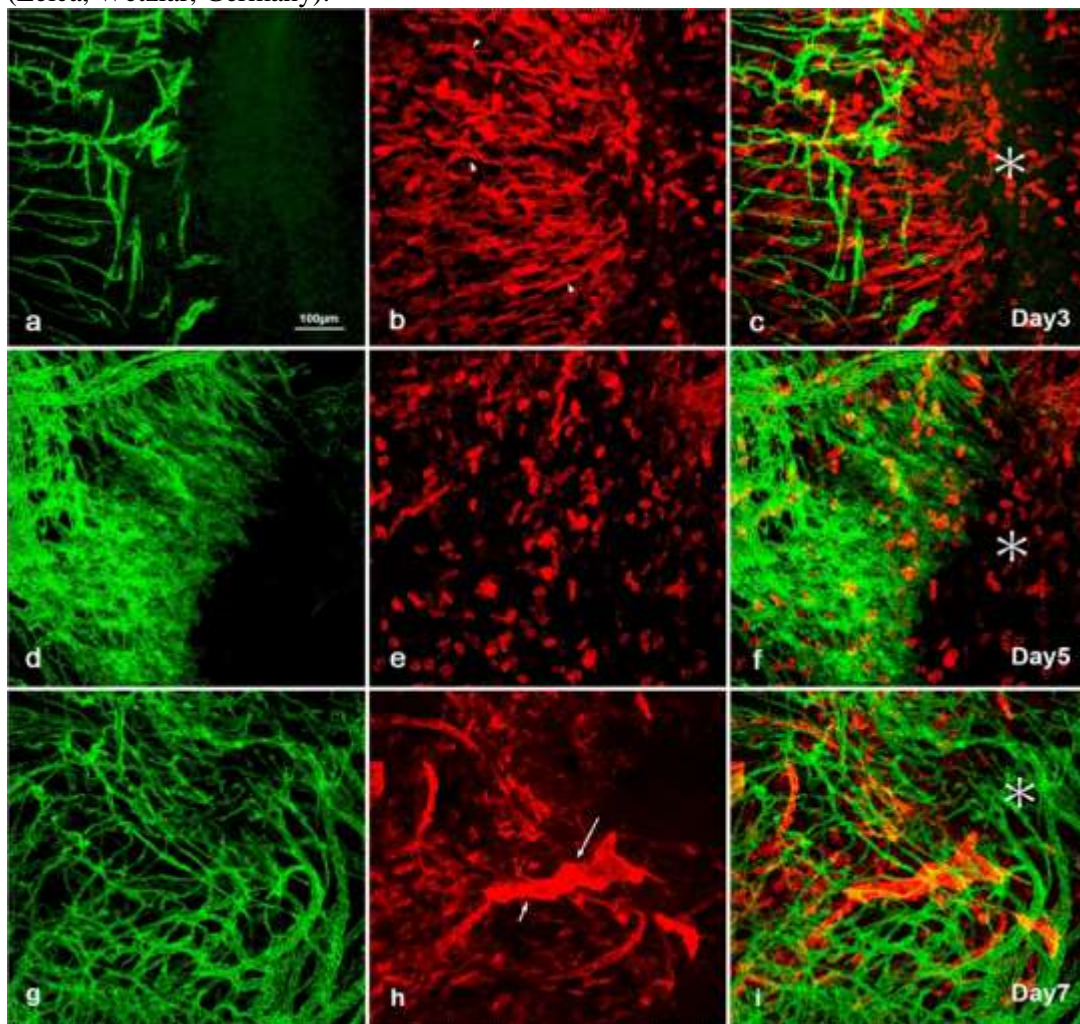
a-c: third day after injury, d-f: fifth day after injury, g-i: seventh day after injury. Green: CD31, red : LYVE-1, *: wound area, scale bar = 100µm.

On the third day after injury, many LYVE-1-positive cells were seen to appear. LYVE-1 positive cells (represented by the arrow markings) extended in row towards the center of the wound (a-c).

On the fifth day after injury, abundant new blood vessels were seen (d), but LYVE-1-positive cells did not line up as they had done on the third day.

Seven days after injury, blood vessels formed a network (g), and lymphatic-like structures could be observed (h, represented by arrows).

These fluorescent images were observed under a Leica TCS-SL confocal laser scanning microscope (Leica, Wetzlar, Germany).

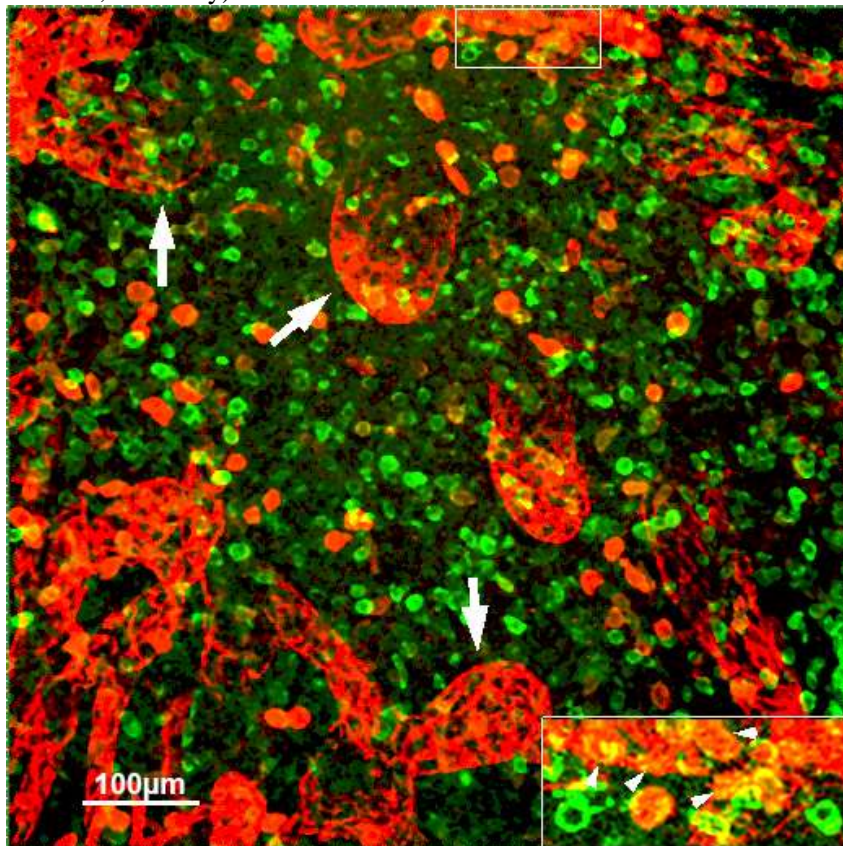


2.4 LYVE-1 / LYVE-1⁺ cells

LYVE-1, a lymphatic endothelial cell marker, is a hyaluronic acid receptor and a CD44 homolog⁴⁸. Moreover, LYVE-1 is expressed in sinusoids, macrophages, and several cells⁴⁹. In order to observe lymphangiogenesis during wound healing, a full-thickness defect was made in the skin of mice. It was stained and observed for

LYVE-1, CD31, and CD11b (Figure 1 and 2). On the third day after injury, LYVE-1 positive macrophages accumulated to form a lymphatic-like structure (Figure 1 a-c), but on the fifth day, this structure deteriorated (Figure 1 d-f). On the 7th day after injury, normal lymphatic vessels were observed (Figure 1g-i).

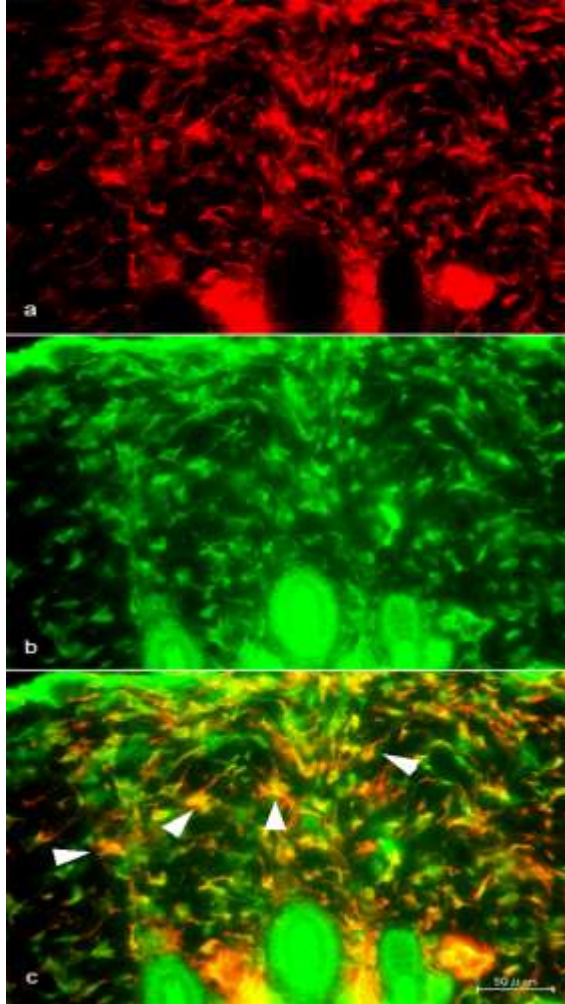
Figure 2: Double immunofluorescent staining in the wound section of the skin in a mouse-model (C57BL / 6N, 10-week-old male) on the 17th day after injury. Green: CD11b, red: LYVE-1, scale = 100µm. The blind ends of the lymphatic vessels were confirmed at the site of the wound on the 17th day after the injury (marked by arrows). In addition, CD11b-positive cells were seen to form the lymphatic endothelium (arrow markings in the figure inset; enlarged and highlighted by a white rectangle). These fluorescent images were observed under a Leica TCS-SL confocal laser scanning microscope (Leica, Wetzlar, Germany).



We also observed that some CD11b-positive macrophages were integrated into the lymphatic endothelium (Figure 2). On the 17th day, many blind ends of lymphatic vessels were observed, and the number of LYVE-1-positive macrophages was also seen to have decreased. Maruyama et al. had

previously reported that macrophages had transdifferentiated and been incorporated into the lymphatic endothelial cells in the cornea of mice⁵⁰. Similarly, in the skin, macrophages may have differentiated to form a part of the lymphatic endothelial cells.

Figure 3: Double immunofluorescent staining in the wound section of the skin in a mouse-model (C57BL / 6N, 8-week-old male) on day 1 after injury. Green: α -SMA, red: PDPN, scale = 50 μ m. Many PDPN positive cells appeared on the first day after injury (a). These cells had many cell processes. In addition, almost all PDPN positive cells co-expressed α -SMA (b, c; represented by arrows). These fluorescent images were observed under a KEYENCE BZ-9000 HS all-in-one microscope (KEYENCE, Osaka, Japan).



2.5 Podoplanin (PDPN) / PDPN⁺ cells

PDPN is among the most commonly used lymphatic markers along with LYVE-1 and VEGFR3. PDPN expression in lymphatic endothelial cells is regulated by PROX1. In the development of lymphatic vessels, PDPN has proven to be essential for isolating lymphatic budding from veins^{51,52}. In wound healing, PDPN is expressed in epithelial basal cells and is involved in epidermal cell migration⁵³. We also found that many PDPN⁺ cells appeared in the stroma using a mouse model to study wound

healing. These cells were α SMA⁺ myofibroblasts (Figure 3). Myofibroblasts differentiate from fibroblasts and epidermal cells. However, the origin of PDPN⁺ myofibroblasts is unknown. PDPN⁺ fibroblasts are involved in leukocyte migration as an “extravascular pathway” in lymphoid organs^{54,55}. In light of these, PDPN⁺ cells may not be directly involved in lymphangiogenesis, but may be indirectly involved in overall wound healing by causing the migration of various cells.

3. Summary

Angiogenesis and lymphangiogenesis are important processes in wound healing. Angiogenesis has been the focus of a lot of research; many manuscripts have studied embryology, tumors, and wound healing in various fields of study such as the molecular biology, physiology, pharmacology, and microanatomy. However, lymphangiogenesis is yet to be explored. Additionally, the contribution of cells which act as lymphatic markers involved in wound healing could be an interesting aspect for further research.

4. Acknowledgments

The authors would like to thank Ms. Ayaka Shibano, Dr. Sachiko Miyamoto-Kikuta, Dr. Masae Morishima, Dr. Shuji Kitahara and Professor Taichi Ezaki for their advice and contribution to this report. This work was supported by Grant-in-aid for Scientific Research (C) (JSPS KAKENHI Grant numbers JP15K10953 and JP18K09496).

5. Conflicts of interest

The authors declare no conflicts of interest associated with this manuscript.

6. *References*

1. Senger DR, Galli SJ, Dvorak AM, Perruzzi CA, Harvey VS, Dvorak HF. Tumor cells secrete a vascular permeability factor that promotes accumulation of ascites fluid. *Science*. 1983; 219(4587):983-985. doi: 10.1126/science.6823562 [doi].
2. Ferrara N, Houck KA, Jakeman LB, Winer J, Leung DW. The vascular endothelial growth factor family of polypeptides. *J Cell Biochem*. 1991; 47(3):211-218. doi: 10.1002/jcb.240470305 [doi].
3. Senger DR, Van de Water L, Brown LF, et al. Vascular permeability factor (VPF, VEGF) in tumor biology. *Cancer Metastasis Rev*. 1993; 12(3-4):303-324. doi: 10.1007/bf00665960 [doi].
4. Tischer E, Mitchell R, Hartman T, et al. The human gene for vascular endothelial growth factor. multiple protein forms are encoded through alternative exon splicing. *J Biol Chem*. 1991; 266(18):11947-11954.
5. Neufeld G, Cohen T, Gengrinovitch S, Poltorak Z. Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J*. 1999; 13(1):9-22.
6. Liu L, Marti GP, Wei X, et al. Age-dependent impairment of HIF-1alpha expression in diabetic mice: Correction with electroporation-facilitated gene therapy increases wound healing, angiogenesis, and circulating angiogenic cells. *J Cell Physiol*. 2008; 217(2):319-327. Accessed Nov 15, 2019. doi: 10.1002/jcp.21503.
7. Fagiani E, Christofori G. Angiopoietins in angiogenesis. *Cancer Lett*. 2013; 328(1):18-26. Accessed Nov 15, 2019. doi: 10.1016/j.canlet.2012.08.018.
8. Cao R, Bråkenhielm E, Pawliuk R, et al. Angiogenic synergism, vascular stability and improvement of hind-limb ischemia by a combination of PDGF-BB and FGF-2. *Nat Med*. 2003; 9(5):604-613. Accessed Nov 15, 2019. doi: 10.1038/nm848.
9. Ridiandries A, Bursill C, Tan J. Broad-spectrum inhibition of the CC-chemokine class improves wound healing and wound angiogenesis. *Int J Mol Sci*. 2017; 18(1). Accessed Nov 15, 2019. doi: 10.3390/ijms18010155.
10. Boniakowski AE, Kimball AS, Jacobs BN, Kunkel SL, Gallagher KA. Macrophage-mediated inflammation in normal and diabetic wound healing. *J Immunol*. 2017; 199(1):17-24. Accessed Nov 15, 2019. doi: 10.4049/jimmunol.1700223.
11. Rodero MP, Khosrotehrani K. Skin wound healing modulation by macrophages. *Int J Clin Exp Pathol*. 2010; 3(7):643-653. Accessed Nov 15, 2019.
12. Lucas T, Waisman A, Ranjan R, et al. Differential roles of macrophages in diverse phases of skin repair. *J Immunol*. 2010; 184(7):3964-3977. Accessed Nov 15, 2019. doi: 10.4049/jimmunol.0903356.
13. Brown LF, Yeo KT, Berse B, et al. Expression of vascular permeability factor (vascular endothelial growth factor) by epidermal keratinocytes during wound healing. *J Exp Med*. 1992; 176(5):1375-1379. Accessed Nov 15, 2019. doi: 10.1084/jem.176.5.1375.
14. Frank S, Hübner G, Breier G, Longaker MT, Greenhalgh DG, Werner S. Regulation of vascular endothelial growth factor expression in cultured keratinocytes. implications for normal and impaired wound healing. *J Biol Chem*. 1995; 270(21):12607-12613. Accessed Nov 15, 2019. doi: 10.1074/jbc.270.21.12607.
15. Grotendorst GR, Okochi H, Hayashi N. A novel transforming growth factor beta response element controls the expression of the connective tissue growth factor gene. *Cell Growth Differ*. 1996; 7(4):469-480. Accessed Nov 15, 2019.

16. Babic AM, Chen CC, Lau LF. Fisp12/mouse connective tissue growth factor mediates endothelial cell adhesion and migration through integrin alphavbeta3, promotes endothelial cell survival, and induces angiogenesis in vivo. *Mol Cell Biol.* 1999; 19(4):2958-2966. Accessed Nov 15, 2019. doi: 10.1128/mcb.19.4.2958.
17. Shimo T, Nakanishi T, Nishida T, et al. Connective tissue growth factor induces the proliferation, migration, and tube formation of vascular endothelial cells in vitro, and angiogenesis in vivo. *J Biochem.* 1999; 126(1):137-145. Accessed Nov 15, 2019. doi: 10.1093/oxfordjournals.jbchem.a022414.
18. Armulik A, Genové G, Betsholtz C. Pericytes: Developmental, physiological, and pathological perspectives, problems, and promises. *Dev Cell.* 2011; 21(2):193-215. Accessed Nov 15, 2019. doi: 10.1016/j.devcel.2011.07.001.
19. Orledge A, D'Amore PA. Inhibition of capillary endothelial cell growth by pericytes and smooth muscle cells. *J Cell Biol.* 1987; 105(3):1455-1462. Accessed Nov 15, 2019. doi: 10.1083/jcb.105.3.1455.
20. Sato Y, Rifkin DB. Inhibition of endothelial cell movement by pericytes and smooth muscle cells: Activation of a latent transforming growth factor-beta 1-like molecule by plasmin during co-culture. *J Cell Biol.* 1989; 109(1):309-315. Accessed Nov 15, 2019. doi: 10.1083/jcb.109.1.309.
21. von Tell D, Armulik A, Betsholtz C. Pericytes and vascular stability. *Exp Cell Res.* 2006; 312(5):623-629. Accessed Nov 15, 2019. doi: 10.1016/j.yexcr.2005.10.019.
22. Rhodin JA, Fujita H. Capillary growth in the mesentery of normal young rats. intravital video and electron microscope analyses. *J Submicrosc Cytol Pathol.* 1989; 21(1):1-34. Accessed Nov 15, 2019.
23. Schlingemann RO, Rietveld FJ, Kwaspen F, van de Kerkhof PC, de Waal RM, Ruiter DJ. Differential expression of markers for endothelial cells, pericytes, and basal lamina in the microvasculature of tumors and granulation tissue. *Am J Pathol.* 1991; 138(6):1335-1347. Accessed Nov 15, 2019.
24. Nehls V, Denzer K, Drenckhahn D. Pericyte involvement in capillary sprouting during angiogenesis in situ. *Cell Tissue Res.* 1992; 270(3):469-474. Accessed Nov 15, 2019. doi: 10.1007/bf00645048.
25. Wesseling P, Schlingemann RO, Rietveld FJ, Link M, Burger PC, Ruiter DJ. Early and extensive contribution of pericytes/vascular smooth muscle cells to microvascular proliferation in glioblastoma multiforme: An immuno-light and immuno-electron microscopic study. *J Neuropathol Exp Neurol.* 1995; 54(3):304-310. Accessed Nov 15, 2019. doi: 10.1097/00005072-199505000-00003.
26. Reynolds LP, Redmer DA. Expression of the angiogenic factors, basic fibroblast growth factor and vascular endothelial growth factor, in the ovary. *J Anim Sci.* 1998; 76(6):1671-1681. Accessed Nov 15, 2019. doi: 10.2527/1998.7661671x.
27. Amselgruber WM, Schäfer M, Sinowatz F. Angiogenesis in the bovine corpus luteum: An immunocytochemical and ultrastructural study. *Anat Histol Embryol.* 1999; 28(3):157-166. Accessed Nov 15, 2019. doi: 10.1046/j.1439-0264.1999.00195.x.
28. Reynolds LP, Grazul-Bilska AT, Redmer DA. Angiogenesis in the corpus luteum. *Endocrine.* 2000; 12(1):1-9. Accessed Nov 15, 2019. doi: 10.1385/ENDO:12:1:1.
29. Morikawa S, Baluk P, Kaidoh T, Haskell A, Jain RK, McDonald DM. Abnormalities in pericytes on blood vessels and endothelial sprouts in tumors. *Am J Pathol.* 2002; 160(3):985-1000. Accessed

- Nov 15, 2019. doi: 10.1016/S0002-9440(10)64920-6.
30. Ozerdem U, Stallcup WB. Early contribution of pericytes to angiogenic sprouting and tube formation. *Angiogenesis*. 2003; 6(3):241-249. Accessed Nov 15, 2019. doi: 10.1023/B:AGEN.0000021401.58039.a9.
31. Morikawa S, Ezaki T. Phenotypic changes and possible angiogenic roles of pericytes during wound healing in the mouse skin. *Histol Histopathol*. 2011; 26(8):979-995. Accessed Nov 15, 2019. doi: 10.14670/HH-26.979.
32. Noishiki C, Yuge S, Ando K, et al. Live imaging of angiogenesis during cutaneous wound healing in adult zebrafish. *Angiogenesis*. 2019; 22(2):341-354. Accessed Nov 15, 2019. doi: 10.1007/s10456-018-09660-y.
33. François M, Short K, Secker GA, et al. Segmental territories along the cardinal veins generate lymph sacs via a ballooning mechanism during embryonic lymphangiogenesis in mice. *Dev Biol*. 2012; 364(2):89-98. Accessed Nov 15, 2019. doi: 10.1016/j.ydbio.2011.12.032.
34. Joukov V, Sorsa T, Kumar V, et al. Proteolytic processing regulates receptor specificity and activity of VEGF-C. *EMBO J*. 1997; 16(13):3898-3911. Accessed Nov 15, 2019. doi: 10.1093/emboj/16.13.3898.
35. Joukov V, Pajusola K, Kaipainen A, et al. A novel vascular endothelial growth factor, VEGF-C, is a ligand for the Flt4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases. *EMBO J*. 1996; 15(2):290-298. Accessed Nov 15, 2019.
36. Achen MG, Jeltsch M, Kukk E, et al. Vascular endothelial growth factor D (VEGF-D) is a ligand for the tyrosine kinases VEGF receptor 2 (Flk1) and VEGF receptor 3 (Flt4). *Proc Natl Acad Sci U S A*. 1998; 95(2):548-553. Accessed Nov 15, 2019. doi: 10.1073/pnas.95.2.548.
37. Alam A, Hault J, Barron P, et al. Heterodimerization with vascular endothelial growth factor receptor-2 (VEGFR-2) is necessary for VEGFR-3 activity. *Biochem Biophys Res Commun*. 2004; 324(2):909-915. Accessed Nov 15, 2019. doi: 10.1016/j.bbrc.2004.08.237.
38. Dixelius J, Makinen T, Wirzenius M, et al. Ligand-induced vascular endothelial growth factor receptor-3 (VEGFR-3) heterodimerization with VEGFR-2 in primary lymphatic endothelial cells regulates tyrosine phosphorylation sites. *J Biol Chem*. 2003; 278(42):40973-40979. Accessed Nov 15, 2019. doi: 10.1074/jbc.M304499200.
39. Goldman J, Rutkowski JM, Shields JD, et al. Cooperative and redundant roles of VEGFR-2 and VEGFR-3 signaling in adult lymphangiogenesis. *FASEB J*. 2007; 21(4):1003-1012. Accessed Nov 15, 2019. doi: 10.1096/fj.06-6656com.
40. Wigle JT, Harvey N, Detmar M, et al. An essential role for Prox1 in the induction of the lymphatic endothelial cell phenotype. *EMBO J*. 2002; 21(7):1505-1513. Accessed Nov 15, 2019. doi: 10.1093/emboj/21.7.1505.
41. Hong Y, Harvey N, Noh Y, et al. Prox1 is a master control gene in the program specifying lymphatic endothelial cell fate. *Dev Dyn*. 2002; 225(3):351-357. Accessed Nov 15, 2019. doi: 10.1002/dvdy.10163.
42. Hong Y, Detmar M. Prox1, master regulator of the lymphatic vasculature phenotype. *Cell Tissue Res*. 2003; 314(1):85-92. Accessed Nov 15, 2019. doi: 10.1007/s00441-003-0747-8.
43. Seo M, Choi J, Rho CR, Joo C, Lee SK. MicroRNA miR-466 inhibits lymphangiogenesis by targeting prospero-related homeobox 1 in the alkali burn corneal injury model. *J Biomed Sci*. 2015; 22:3. Accessed Nov 15, 2019. doi: 10.1186/s12929-014-0104-0.

44. Dagenais SL, Hartsough RL, Erickson RP, Witte MH, Butler MG, Glover TW. Foxc2 is expressed in developing lymphatic vessels and other tissues associated with lymphedema-distichiasis syndrome. *Gene Expr Patterns*. 2004; 4(6):611-619. Accessed Nov 15, 2019. doi: 10.1016/j.modgep.2004.07.004.
45. Petrova TV, Karpanen T, Norrmén C, et al. Defective valves and abnormal mural cell recruitment underlie lymphatic vascular failure in lymphedema distichiasis. *Nat Med*. 2004; 10(9):974-981. Accessed Nov 15, 2019. doi: 10.1038/nm1094.
46. Fatima A, Wang Y, Uchida Y, et al. Foxc1 and Foxc2 deletion causes abnormal lymphangiogenesis and correlates with ERK hyperactivation. *J Clin Invest*. 2016; 126(7):2437-2451. Accessed Nov 15, 2019. doi: 10.1172/JCI80465.
47. Norrmén C, Ivanov KI, Cheng J, et al. FOXC2 controls formation and maturation of lymphatic collecting vessels through cooperation with NFATc1. *J Cell Biol*. 2009; 185(3):439-457. Accessed Nov 15, 2019. doi: 10.1083/jcb.200901104.
48. Banerji S, Ni J, Wang SX, et al. LYVE-1, a new homologue of the CD44 glycoprotein, is a lymph-specific receptor for hyaluronan. *J Cell Biol*. 1999; 144(4):789-801. Accessed Nov 15, 2019. doi: 10.1083/jcb.144.4.789.
49. Mouta Carreira C, Nasser SM, di Tomaso E, et al. LYVE-1 is not restricted to the lymph vessels: Expression in normal liver blood sinusoids and down-regulation in human liver cancer and cirrhosis. *Cancer Res*. 2001; 61(22):8079-8084. Accessed Nov 15, 2019.
50. Maruyama K, Ii M, Cursiefen C, et al. Inflammation-induced lymphangiogenesis in the cornea arises from CD11b-positive macrophages. *J Clin Invest*. 2005; 115(9):2363-2372. Accessed Nov 15, 2019. doi: 10.1172/JCI23874.
51. Schacht V, Ramirez MI, Hong Y, et al. T1alpha/podoplanin deficiency disrupts normal lymphatic vasculature formation and causes lymphedema. *EMBO J*. 2003; 22(14):3546-3556. Accessed Nov 15, 2019. doi: 10.1093/emboj/cdg342.
52. Suzuki-Inoue K, Fuller GLJ, García A, et al. A novel syk-dependent mechanism of platelet activation by the C-type lectin receptor CLEC-2. *Blood*. 2006; 107(2):542-549. Accessed Nov 15, 2019. doi: 10.1182/blood-2005-05-1994.
53. Asai J, Hirakawa S, Sakabe J, et al. Platelets regulate the migration of keratinocytes via podoplanin/CLEC-2 signaling during cutaneous wound healing in mice. *Am J Pathol*. 2016; 186(1):101-108. Accessed Nov 15, 2019. doi: 10.1016/j.ajpath.2015.09.007.
54. Shimizu K, Morikawa S, Kitahara S, Ezaki T. Local lymphogenic migration pathway in normal mouse spleen. *Cell Tissue Res*. 2009; 338(3):423-432. Accessed Nov 15, 2019. doi: 10.1007/s00441-009-0888-5.
55. Herzog BH, Fu J, Wilson SJ, et al. Podoplanin maintains high endothelial venule integrity by interacting with platelet CLEC-2. *Nature*. 2013; 502(7469):105-109. Accessed Nov 15, 2019. doi: 10.1038/nature12501.