

## REVIEW ARTICLE

# Pathology and Molecular Mechanisms of Perineural Spread of Tumors

### Authors

Yasra Sayyed<sup>1†</sup>, Rengpeng Ji<sup>1</sup>, Tean Zaheer<sup>2</sup> Luyuan Li<sup>1</sup>

### Affiliations

<sup>1</sup> Nankai University College of Pharmacy, Tianjin, China

<sup>2</sup> Faculty of Veterinary Science, University of Agriculture Faisalabad, Pakistan

† Corresponding Author

### Corresponding author:

Yasra Sayyed

Address: No. 38 Tongyan Road, College of Pharmacy, Nankai University, Tianjin

E-mail: [yasra.syed14@gmail.com](mailto:yasra.syed14@gmail.com)

Phone/Fax: +8618822033746

### Abstract

Perineural invasion (PNI) is an important but under-reported route of metastasis of many cancers, in which neoplasm invades and spreads along the nerves. In recent years, PNI has been identified to contribute to the pathology of malignant tumors in the breast, stomach, head and neck, pancreas, prostate and large intestine. PNI of neoplasm may be attributable to poor prognosis of the patients, and sometimes appears to be the only cause of long-distance metastasis. Recent studies have furnished latest insights into the pathology and clinical features of PNI, characterized by continuous and interlinked multiple steps, starting from the formation of a perineural niche, comprising of neural cells, inflammatory cells, stromal cells, extracellular matrix, and blood vessels, in addition to the cancer cells. The critical step of PNI involves the establishment of connections between tumor and nerve through a number of signaling pathways consisting of soluble factors such as nerve growth factor, interleukins, and matrix metalloproteinases. Upon invasion into the nervous system, the cancer cells bring changes to neural cells and their microenvironment, leading to neoplastic spread along the nerves and alteration of normal nerve functions. In this review, we attempt to comprehensively cover the cellular and molecular mechanisms of perineural spread of tumors.

**Keywords:** Perineural invasion, Tumor, Perineural spread, Neoplasm, Perineural niche

## Introduction

Perineural spread of tumor may be well-defined as an expansion of the prime tumor through the neural sheath tissues (perineurium and epineurium) of a nerve<sup>1</sup>. It is well-accepted that typical routes of malignant tumor metastasis and invasion are blood and lymphatic vessels<sup>2</sup>. Perineural spread of tumors, in which neoplasm invades and spreads along the nerves, is a relatively under-studied process of cancer progression even though nerves were established to be involved in epithelial cell adenoma and later on in breast, bladder, pancreatic and colorectal tumors about three decades ago<sup>3-7</sup>. Initially described as the invasion of cancer cells near or passing through tumor nerves<sup>2, 8</sup>, perineural spread of cancer cells was termed perineural invasion (PNI) by Batsakis in 1985 when the characteristics of this unique route of tumour invasion began to draw the attention of oncologists. PNI is now recognized as a distinctive pathological entity attributable to distant tumour spread beyond local invasion<sup>8</sup>. Mechanistically, PNI may involve reciprocal signalling interactions between nerves and tumor, and the invading cancer cells may have acquired the ability to respond to pro-invasion signals within the peripheral nerve milieu<sup>9, 10</sup>. The axon part of neuron is covered by a covering sheath known as neurolemma which is made up of epineurium, perineurium and endoneurium from outside to inside respectively. Nerves can grow in the direction of a tumor in various patterns, such as concentric lamellar, complete surrounding, partial surrounding, and tangent contact. PNI could be the sole route of tumor spread in the absence of all other routes<sup>10</sup>. Chronic initiation of the sympathetic nervous system by enhanced norepinephrine levels in the tumor microenvironment has been responsible for

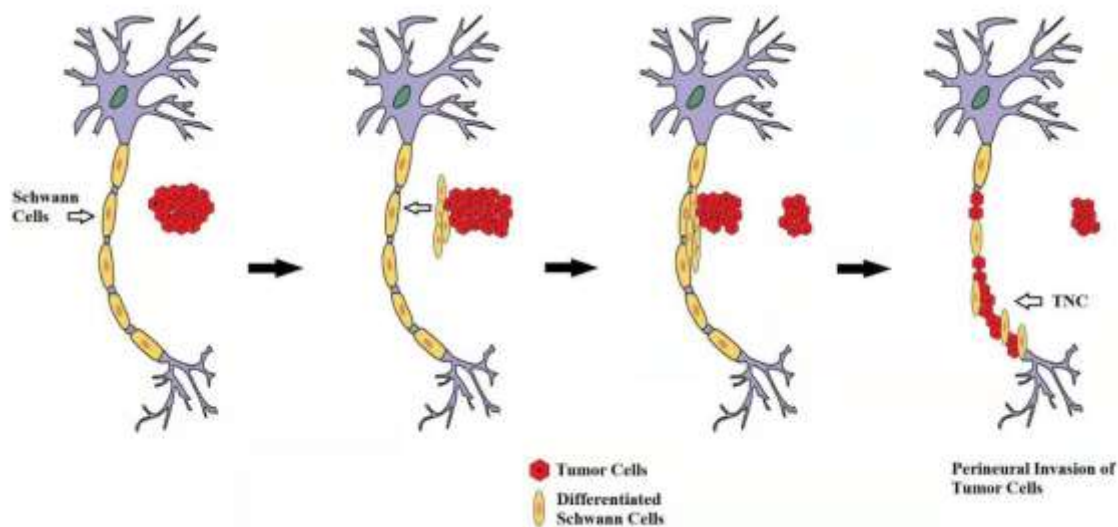
tumor spread. Neoneurogenesis is another key driver for progression of tumor<sup>11, 12</sup>. PNI has been shown as a pathological feature in cancers of multiple organs including stomach, large intestine, pancreas, head and neck, accompanied by low survival rates of the patients<sup>10</sup>. It is therefore important to take PNI into consideration while studying cancer pathology and devising better personalized point of care therapies in PNI patients.

## Multistep and continuous process of PNI

The importance of PNI in cancer is evident from its widespread occurrence in various organs of the body (**Table 1**). Previously, it was thought that PNI was a relatively less common route of cancer invasion and metastasis, requiring the availability of a “low resistance channel”<sup>13, 14</sup>. Based on the aforementioned hypothesis, PNI occurs only when the tumor is present in close proximity to the nerves, and neoplastic cells gain entry into the low resistance and cells-free perineural space with the help of nerve sheath and surrounding lymphatics. As research into the pathology of PNI deepens, evidence begin to support that occurrence of PNI involves multiple steps and a variety of factors<sup>2</sup>. A new hypothesis of tumor neural connects (TNC)<sup>14</sup> was developed to explain that the presence of a complex signaling network between tumor and nerve is chiefly responsible for the initiation of PNI (**Figure 1**). This complex signal network is produced by a number of factors including, supporting cells, inflammatory cells, immune components, blood vessels and the extracellular matrix. Additionally, number of soluble signal molecules and their receptors are discovered to take part in the process<sup>15</sup>, including interleukin 6 (IL-6)<sup>16</sup>, matrix metalloproteinases (MMPs)<sup>17</sup>, nerve growth factor (NGF)<sup>18</sup>, and glial cell line-derived neurotrophic factor (GDNF)<sup>19</sup>.

**Table 1:** Importance of PNI in cancer of different organs of the body

PNI in cancer of different organs	Recurrence rate	Prognosis	References
Adenocarcinoma of the pancreatic duct	70%-100%	Poor	94-98
Stomach carcinoma	6.8%-75.6%	Poor	99-102
Prostate cancer	83.6%	Poor	103-105
Cervical cancer	8.6%-31.3%	Shortened disease-free survival time	106-108
Biliary tract cancer	56%-88%	28% patients survived up to 5 years	108 107
Colorectal cancer	15.7%-38.9%	Poor	109-112
Head and neck cancer	30-100%	Poor	113, 114

**Figure 1:** Schematic representation of perineural spread of cancer cells. Depicted are formation of tumor-neural connect (TNC) and spread of tumor cells into neuronal axon

PNI is a continuous and multistep process<sup>20, 21</sup>. The initial step is the direct interaction between cancer cells and nerve cells. The interaction is apparently bilateral: both the growth of a tumor to engulf a branch of nerves as well as the in-growth of nerves into a nearby tumor are observed<sup>22</sup>. A number of interlinked events include survival of

neoplastic cells, neural formation, initiation of inflammation, recruitment of neoplastic cells to the nerves, regeneration of nerves, binding of neurolemma with tumor cells and, eventually, cancer cell invasion into the nerve. Apart from these fundamental steps, variation in supporting cells, nerve cells and perineural matrix, increased tumor cells

invasion, neogenesis of nerve cells, interaction of tumor cells with nerve cells and escape of tumor cells from apoptosis, are also associated with PNI<sup>23</sup>.

### **Nerve-Tumor Interaction**

Growth and extension of axon and increased thickening of nerve fibers are the main steps in the interaction of nerve and tumor. The production of growth factors and neurotrophic factors in response to the nerve-tumor interaction leads to nerve growth in tumor, axon lengthening and nerve thickening<sup>24</sup>. Using nerve innervated mouse prostatic epithelial cells to study histological and genetic aspects of tumor, it was shown that the innervated integral nerves of tumor play a primary role in cancer development<sup>25</sup>. Usually characterized by abnormal energy metabolism pathways and high ratio of cancer cell cytoplasm; when the tumor was denervated, reduced cancer cell cytoplasm, condensed chromatin, and increased cancer cell apoptosis occurred<sup>26</sup>. Similar observations were made with mouse prostate cancer<sup>27</sup> and stomach cancer cells<sup>28</sup>. These findings indicate that the innervation of tumors and tumor invasion into the nerves may take place simultaneously during the course of PNI.

The importance of neurotransmitters in PNI has been underscored by numerous studies. The products of sympathetic nervous system (adrenergic fibers) and parasympathetic system (cholinergic fibers) participate in the initiation of tumor succession and tumor invasion, respectively<sup>28</sup>. Two important nerve cell protein families, namely neuroligins and neurexins, affect the communication between nerve cells and blood vessels<sup>29, 30</sup>. Two subtypes of neuroligins, NLGN1 and NLGN2, affect the glutamic acid reaction and secretion of catecholamine, respectively<sup>31</sup>.

More recent studies focus on the relation of micro RNA (miRNA) and long non-coding

RNA (lncRNA) with PNI. Abnormal expression of lncRNA-associated lung adenocarcinoma transcript 1 (lncRNA-MALAT-1) has been reported in several types of cancers, such as in hilar cholangiocarcinoma, linking the occurrence of PNI to tumor progression and metastasis<sup>32</sup>. Increased expression of lncRNA-MALAT-1 is also implicated in PNI in colon cancer<sup>33</sup> and renal cancer<sup>34</sup>. Similarly, upregulated expression of miR-21 in oral tumor is associated with depressed expression of phosphatase and tensin homolog, promoting PNI by allowing invasion and dispersion of tumor cells in nerves, leading to a poor survival rate of patients<sup>35</sup>. High levels of miR-21 but low levels of miR-100 and miR-125b are shown to occur in head and neck squamous cell carcinoma, again facilitating PNI<sup>36</sup>. In breast cancer, PNI is promoted by high levels of miR-370<sup>37</sup>. Developing the miRNA based prophylaxis for PNI tumors can be a major breakthrough, preventing poor prognostic rates.

### **The Role of Schwann Cells in PNI**

Regeneration of axons and survival time of transformed nerve cells are promoted by Schwann cells (also known as neuroglia). Movement of Schwann cells towards the tumor was described prior to the occurrence of PNI<sup>38</sup>. Acted upon by glial fibrillary acidic protein of Schwann cells, cancer cells can break up from a tumor mass and migrate towards Schwann cells, promoted by neural cell adhesion molecule 1 (NCAM-1)<sup>39</sup>. Additionally, Schwann cells secrete soluble signal molecules (chemokines, growth factors, biologically active small molecules and cytokines), that enhance the alteration of extracellular matrix, invasion of newly formed nerves, axon growth, tumor cell survival and neogenesis of damaged nerves<sup>40</sup>. Schwann cells have also been reported to induce salivary adenoid cystic carcinoma (SACC) cells to change into

mesenchymal form through the action of brain derived nerve growth factor (BDNF) or tropomyosin receptor kinase B (TrkB), indicating that the Schwann cells are highly capable of enhancing the epithelial mesenchymal transition (EMT), aiding cancer invasion and metastasis<sup>41, 42</sup>. Moreover, the occurrence of mesenchymal epithelial transition (MET) has been reported when Schwann cells were cultivated with pancreatic cancer cells; upregulation of membrane expression of E-cadherin protein and down-regulation of vimentin and small mothers-against-ecapentaplegic (SMAD-3) proteins expression were detected during this interaction<sup>43</sup>. Decreased expression of SMAD3 is associated with activation of transforming growth factor  $\beta$  (TGF- $\beta$ ), apparently enhancing the migration of neoplastic cells<sup>44</sup>.

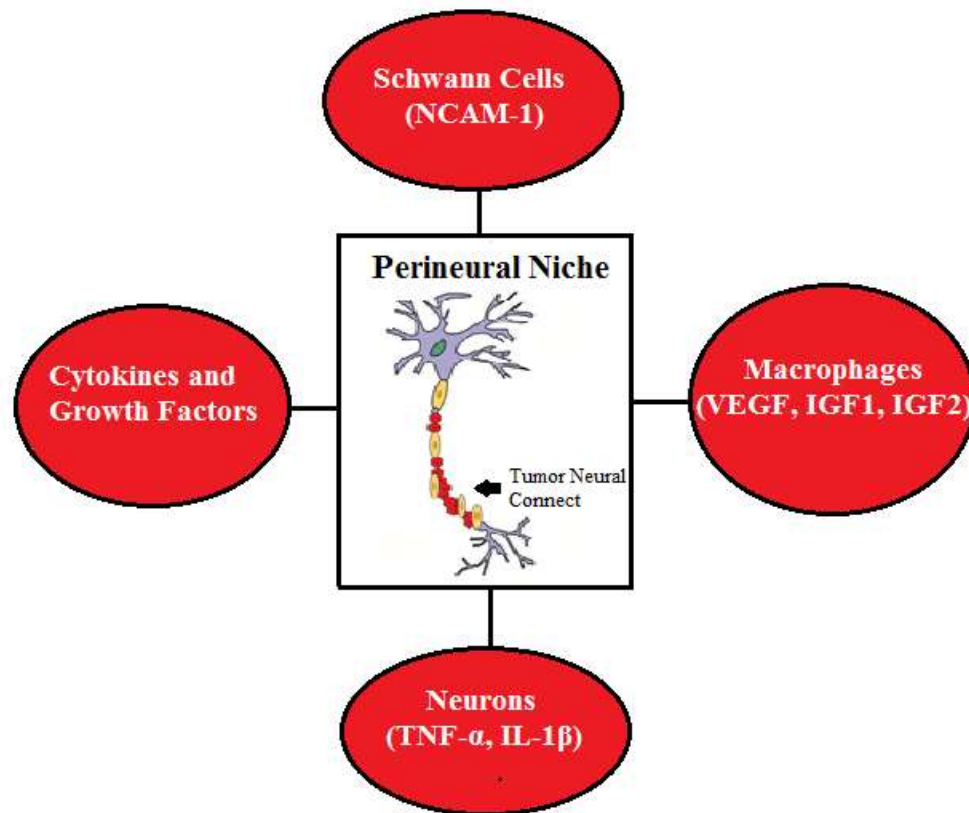
### Microenvironmental Factors Involved in PNI

An altered microenvironment is characteristic of PNI. This includes the occurrence of immature collagen and accumulation of fibroblasts in the extracellular matrix<sup>45</sup>, low oxygen levels, enrichment of inflammatory cells, high glucose levels, and activation of sympathetic nervous system<sup>23</sup>. Hypoxia in the tumor leads to activation of hypoxia inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ) that accelerates the neogenesis of blood vessels<sup>46</sup> and the induction of the gene expression of chemokines and their receptors such as CXR4, CXCL12 and CX3CR1, which leads to upregulation of stem cell factor (SCF) and vascular endothelial growth factor (VEGF)<sup>47</sup>. Additionally, glucose transfer and glycolytic enzyme expression are enhanced by glucoamylase 1 (GLU-1) upregulation<sup>48, 49</sup>. Moreover, neoplastic cell resistance to apoptosis and increase in cell proliferation

rates are strengthened by increased activity of telomerase reverse transcriptase(TERT) and surviving enzymes. These changes apparently attribute to PNI<sup>49</sup>. Furthermore, the activation of sympathetic system also leads to an increase of norepinephrine levels in the tumor microenvironment, which in turn gives rise to upregulation of chemotactic promoter by  $\beta$ -adrenergic receptor signal transduction pathway, facilitating the tumor growth<sup>50</sup>. Axonal sprouting, nerve homeostasis and dendritic growth are the roles played by the key factors that are secreted from Schwann cells, neurons, cytokines and growth factors in a perineural microenvironment, which ultimately endure and initiate cancer metastasis and invasion<sup>20, 51</sup> (**Figure 2**).

Other types of cells in the tumor that facilitate tumor growth may also promote PNI. For instance, in pancreatic tumor, pancreatic stellate cells may play a key role. Activation of pancreatic stellate cells along with Matrix Metalloproteinase 2 (MMP-2) and MMP-9 enzymes, is induced by Sonic Hedgehog (SHH) transcription pathway. As the stellate cells and fibroblasts cover the pancreatic tumor cells in microenvironment, their activation by SHH leads to increased tumor invasion and tumor cell survival<sup>52</sup>. Fibroblasts also play key role in tumor invasion by synthesizing chemotactic factors, growth factors and perineurium sheath<sup>2</sup>. Macrophages play role in PNI; more accurately, a subgroup of macrophages which is known as endoneurial macrophages, is responsible for the neogenesis of damaged nerves and continuity of nerve action in steady state. Pancreatic tumor cells secretes Macrophage Colony Stimulating Factor 1 (MCSF-1) which leads to the invasion of macrophages towards the chemical signals, hence promoting the PNI<sup>51</sup>.





**Figure 2:** Key cells involved in Perineural Niche that can potentially facilitate cancer invasion

### Medkine, Pleiotrophin, and Metalloproteinases in PNI

Medkine (MDK) and pleiotrophin (PTN) of heparin binding growth factor family are implicated in PNI. It was shown that binding of PTN to its high affinity receptor N-syndecan causes neovascularization, which facilitates growth of tumor cells and axons, and thus promote PNI<sup>53, 54</sup>. Upon the invasion of tumor cells into pancreatic nerves, Schwann cells and main nerve cells release N-syndecan. The PTN-N-syndecan complex causes the influx of more PTN-activated cells to the nerve<sup>55</sup>. Similarly, the binding of MDK to the syndecan-3 receptor stimulates lymphatic metastasis of cancer cells, promoting PNI as a result<sup>56</sup>. Upregulated expression of the Zinc-dependent endopeptidase family of matrix metalloproteinases (MMPs), which are involved in tissue remodeling and

degradation of extracellular matrix, has been linked to the occurrence of PNI in stomach cancer, lung cancer, liver cancer, breast cancer, pancreatic cancer and cholangiocarcinoma<sup>57-59</sup>. Specifically, extracellular signal regulated kinases (ERK)/MAPK pathway is shown to be responsible for the phosphorylation of ETS1 gene by  $\beta 6$  integrin which induces MMP-3 and MMP-9 expression, leading to PNI<sup>60</sup>. In another study, PNI of stomach cancer was found to be promoted by higher expression of phosphorylated epidermal growth factor receptor (EGFR), MMP7 and ERK1/2<sup>61</sup>. The mechanisms of MMP-promoted PNI include invasion of tumor cells facilitated by the degradation of extracellular matrix and reduced adhesion of tumor cells, inhibition of apoptosis and upregulation of tumor cell proliferation, activation of EMT, promotion of metastasis of cancer cells by activating

growth factors, and upregulation of tumor angiogenesis<sup>62-70</sup>.

Few other molecules involved in the process of PNI are survivin, slug, galanin, neuropeptide Y and substance P (**Table 2**). Survivin has been recently reported to block the mitochondrial function and receptor's apoptosis, thus block the tumor cell apoptosis<sup>71</sup>. Slug is the member of Zinc Finger Transcription Factor (ZFTF) family and found to promote PNI through the enhancement of EMT process by blocking E-cadherin and positively regulating the MMP promoting factors<sup>72, 73</sup>. After binding to the galanin receptor 2, Galanin promotes the

transcription of cyclooxygenase-2 (COX-2) to produce prostaglandin E2 (PGE2). Consequentially, perineural invasion is promoted by increased production of PGE2<sup>24, 74</sup>. The neuropeptide Y is produced and secreted by affected nerve cells and tumor cells itself<sup>75</sup>. Substance P is not only responsible for distant invasion of neoplastic cells by activating MAPK pathway but is also related to the pain associated with tumor<sup>76</sup>. A host of growth factors and cytokines, produced in the tumor microenvironment by tumor cells and non-tumor cell, have been described to support the mechanism of neural tracking in which cancer cells show capacity to actively migrate alongside nerves.

**Table 2: Molecules involved in the process of PNI**

Molecules/Cytokines Involved	Major Role	Mechanism	Expression	Ref.
SURVIVIN	Anti-apoptotic	Blocks the mitochondrial function & receptor's apoptosis	High, especially in Pancreatic cancer	71
SLUG	Enhance EMT, regulation of the MMP	Blocks E-cadherin	High in basal layers of Epidermis	72, 73, 115
Galanin	Increased production of PGE2	Transcription of COX-2)to produce PGE2, after binding to the galanin receptor 2	Down regulated in gastric cancer	24, 74, 116
Neuropeptide Y	Angiogenesis	Act via malleability of NPY receptors	High in adrenal cortical tumors	75, 117, 118
Substance P	Angiogenesis & distant invasion of neoplastic cells	Activates the MAPK pathway	High in colorectal cancer	119, 76, 120
Nerve growth factor (NGF)	Neurogenesis	Uptake of neurotrophic factor by nerve terminals	High in Breast Cancer	18, 121, 122

Glial cell line-derived neurotrophic factor (GDNF)	Proliferation, migration of enteric progenitors, differentiation of neurons	Neurotrophic action via GDNF/GFR $\alpha$ /Ret complex formation	Human pancreatic cancer cells	123 19, 124-126
Neural cell adhesion molecule (NCAM)	Neural development	Homophilic, Ca <sup>2+</sup> -independent, binding mechanism	Small-cell lung carcinoma and neuroblastoma	127-130
Matrix metalloproteinases (MMPs)	Morphogenesis	Degrade ECM proteins during the process of cancer metastasis and invasion	Human breast cancer	17, 131-133
Tumor Necrosis Factor (TNF- $\alpha$ )	Carcinogenesis	Induces IL-6 release through the inhibitory kappa B (I $\kappa$ B)-nuclear factor kappa B (NF $\kappa$ B) pathway	High in Ovarian cancer, Breast cancer, prostate cancer	134-138
Interleukin 6 (IL-6)	Protumorigenic effect	Agonist for cells expressing gp130. Induce the transcription of factors in many paths of inflammation	Prostate cancer	139-141
Transforming Growth Factor $\beta$ (TGF- $\beta$ )	Invasion and metastasis by Increased TGF- $\beta$ 1 mRNA in cancers	Induce epithelial-mesenchymal transition	High in Breast cancer	142-144
Interleukin 10 (IL-10)	Pro- and antitumoral effects	Inhibits NF- $\kappa$ B signalling	High in cervical cancer	145, 146

### Neurotrophin factors

Neurotrophin factors produced by tumor cells and nerve tissues are important protein molecules for the regulation of tumor nerve generated signal pathway, growth and survival of nerves associated with tumor. Nerve Growth Factor (NGF) integrates with Tyrosine Kinase (Trk) high affinity receptor and p75 Neurotrophin (p75NTR) low affinity

receptors are expressed in tumor cells. This activated NGF in tumor cells and nerve sheath contributes to its important role in PNI by forming the axons, differentiating the nerves and enhancing the damaged nerve survival time<sup>77-79</sup>. Expression level of NGF can vary in different conditions; in case it is overexpressed it can reduce the apoptosis of tumor cell of pancreas, enhance the proliferation of nerves, boost up the TrkA



signal, enhances the MMP-2 and also activates the Mitogen Activated Protein Kinase (MAPK) p44/42 signal pathway. In short, overexpression of NGF leads to PNI of pancreatic cancer<sup>49</sup>. In a study, negative feedback is also shown with respect to the NGF-TrkA pathway; reporting that the pathway helps the tumor cells to survive and migrate to the dorsal root ganglia but upon reaching there, the occurrence of PNI gets reduced due to inhibition of NGF-TrkA pathway by neutralizing antibodies and interfering RNA<sup>80</sup>. Neurotrophin 3 (NT-3) and TrkB are high affinity receptors of Brain Derived Neurotrophic Factor (BDNF); this neurotrophin-receptor complex promotes PNI and poor prognosis by production of MMP-2 from tumor cells. *In vitro* studies showed that growth and advancement of pancreatic and prostatic cancer in mouse was stopped by blocking the NT-3 receptor<sup>81-84</sup>. The survival of many types of neurons in humans is potentially promoted by a small protein “Glial Cell Derived Neurotrophic Factor” (GDNF). Other three members of same family are neurturin, artemin and persephin. Survival of tumor cells, differentiation of nerves, axon growth and signal pathways of growth are related to GDNF. For its expression in tumor, GDNF uses its receptor GFR $\alpha$ 1/RET and it has been reported that the occurrence of PNI was reduced by blocking the GFR receptor<sup>85, 86</sup>. Nerves and macrophages secrete GDNF, that forms GDNF-GFR $\alpha$ 1 complex after binding with GFR receptor. A gene of tumor cells known as “Rearranged During Transfection gene” (RET), binds with GDNF-GFR $\alpha$ 1 complex and controls the metastasis of cancer cells, enhances the invasion, initiates the MMPs excitation, growth of axons and ultimately increases growth of cancer cells by activating MAPK pathway through the induction signal transduction pathway<sup>20, 51, 87, 88</sup>. Similar to GDNF, the second member of same family, artemin also uses a GFR receptor (GFR $\alpha$ 3) to form a

complex and expressed in pancreatic cancer leading to PNI<sup>89</sup>.

### Perspectives

The function of systemic therapies in the controlling perineural spread and perineural invasion remains uncertain. Treatment procedures for patients with PNI have not been recognized. A study revealed that after a median time of 21 months, treatment on PNI with surgery followed by radiation therapy and chemotherapy, 1/3<sup>rd</sup> of patients died from the disease<sup>90, 91</sup>. There is a versatile technology called as CRISPR associated protein 9 (CRISPR/ Cas9), in a sequence-specific manner delivers the capacity to enhance or eliminate DNA in the genome<sup>92,93</sup>. Vigorous research is required, proceeding to increase the efficacy of CRISPR/Cas9 targeting. Schwann cells show affinity to cancer cells, and secrete soluble signal molecules (chemokines, growth factors, biologically active small molecules and cytokines)<sup>22</sup>. Therefore, targeting these signal molecules can inhibit nerve invasion. Considering the range of benign procedures that may look like perineural connection, might assist to prevent excessively aggressive treatments, diagnostic misperception, and misdiagnosis. Efforts should be made to explain the utmost common patterns of spread to increase treatment results but reducing the effects of radiation.

Perineural invasion (PNI) has attracted the attention of cancer researchers as an important aspect in tumor invasion and metastasis. The process involves interlinked multiple steps, beginning from the development of a perineural niche, which includes inflammatory cells, neural cells, supporting cells, and blood vessels. A number of signalling pathways have been identified, as well as many soluble protein factors and extracellular matrix components that act to connect tumors and the nerves. PNI is now recognized as an important feature in the

pathology of malignant tumors of several organs, in which neoplasm invades and spreads along the nerves, yet it is under-stated route of cancer metastasis. As this perineural invasion raises disease sternness, an improved consideration of how the process is controlled may benefit in the improvement of therapeutics aiming at tumor-neuronal connections. Identification of new molecular mechanisms of PNI will undoubtedly emerge from in-depth research in this field, opening

gates to new approaches in more personalized cancer treatment.

**Acknowledgement:** The work is supported in part by grants from the National Science Foundation of China to Luyuan Li (82073064, 81874167). We thank Mr. Ammar Ahmed for his skilful assistance on drawing the illustrations. We thank Mr. Zubair Khalid and Ms. Samantha Bradley for proofreading the manuscript.

## References

1. Stambuk HE. Perineural tumor spread involving the central skull base region. *Seminars in ultrasound, CT, and MR*. Oct 2013;34(5):445-58. doi:10.1053/j.sult.2013.09.002
2. Chen S-H, Zhang B-Y, Zhou B, Zhu C-Z, Sun L-Q, Feng Y-J. Perineural invasion of cancer: a complex crosstalk between cells and molecules in the perineural niche. *Am J Cancer Res*. 2019;9(1):1-21.
3. Ronaghy A, Yaar R, Goldberg LJ, Mahalingam M, Bhawan J. Perineural Involvement: What Does it Mean? *The American Journal of Dermatopathology*. 2010;32(5)
4. Seifert P, Spitznas M. Tumours may be innervated. *Virchows Archiv : an international journal of pathology*. 03/01 2001;438:228-31. doi:10.1007/s004280000306
5. Seifert P, Benedic M, Effert P. Nerve fibers in tumors of the human urinary bladder. *Virchows Archiv*. 2002/03/01 2002;440(3):291-297. doi:10.1007/s004280100496
6. Lü S-H, Zhou Y, Que H-P, Liu S-J. Peptidergic innervation of human esophageal and cardiac carcinoma. *World J Gastroenterol*. 2003;9(3):399-403. doi:10.3748/wjg.v9.i3.399
7. Liang Y-J, Zhou P, Wongba W, Guardiola J, Walker J, Yu J. Pulmonary innervation, inflammation and carcinogenesis. *Sheng li xue bao : [Acta physiologica Sinica]*. 06/25 2010;62:191-5.
8. Batsakis JG. Nerves and neurotropic carcinomas. *The Annals of otology, rhinology, and laryngology*. Jul-Aug 1985;94(4 Pt 1):426-7.
9. Ayala GE, Wheeler TM, Shine HD, et al. In vitro dorsal root ganglia and human prostate cell line interaction: Redefining perineural invasion in prostate cancer. <https://doi.org/10.1002/pros.1137>. *The Prostate*. 2001/11/01 2001;49(3):213-223. doi:<https://doi.org/10.1002/pros.1137>
10. Liebig C, Ayala G, Wilks JA, Berger DH, Albo D. Perineural invasion in cancer: a review of the literature. *Cancer*. Aug 1 2009;115(15):3379-91. doi:10.1002/cncr.24396
11. Lutgendorf SK, DeGeest K, Dahmouch L, et al. Social isolation is associated with elevated tumor norepinephrine in ovarian carcinoma patients. *Brain Behav Immun*. 2011;25(2):250-255. doi:10.1016/j.bbi.2010.10.012
12. Hassan S, Karpova Y, Baiz D, et al. Behavioral stress accelerates prostate cancer development in mice. *J Clin Invest*. Feb 2013;123(2):874-86. doi:10.1172/jci63324
13. Karak S, Quatrano N, Buckley J, Ricci A. Prevalence and significance of perineural invasion in invasive breast carcinoma. *Connecticut medicine*. 01/01 2010;74:17-21.
14. Liebig C, Ayala G, Wilks JA, Berger DH, Albo D. Perineural invasion in cancer. *Cancer*. 2009/08/01 2009;115(15):3379-3391. doi:10.1002/cncr.24396
15. Scuteri A, Miloso M, Foudah D, Orciani M, Cavaletti G, Tredici G. Mesenchymal stem cells neuronal differentiation ability: a real perspective for nervous system repair? *Current stem cell research & therapy*. Jun 2011;6(2):82-92. doi:10.2174/157488811795495486
16. Heikkilä K, Ebrahim S, Lawlor DA. Systematic review of the association between circulating interleukin-6 (IL-6) and cancer. *European journal of cancer (Oxford, England : 1990)*. May 2008;44(7):937-45. doi:10.1016/j.ejca.2008.02.047
17. Jabłońska-Trypuć A, Matejczyk M, Rosochacki S. Matrix metalloproteinases (MMPs), the main extracellular matrix (ECM) enzymes in collagen degradation, as a

- target for anticancer drugs. *Journal of enzyme inhibition and medicinal chemistry*. 2016;31(sup1):177-183. doi:10.3109/14756366.2016.1161620
18. Aloe L, Rocco ML, Balzamino BO, Micera A. Nerve Growth Factor: A Focus on Neuroscience and Therapy. *Curr Neuropharmacol*. 2015;13(3):294-303. doi:10.2174/1570159x13666150403231920
19. Gash DM, Gerhardt GA, Slevin JT. GDNF (Including Neurturin)☆. *Reference Module in Neuroscience and Biobehavioral Psychology*. Elsevier; 2017.
20. Marchesi F, Piemonti L, Mantovani A, Allavena P. Molecular mechanisms of perineural invasion, a forgotten pathway of dissemination and metastasis. *Cytokine & growth factor reviews*. Feb 2010;21(1):77-82. doi:10.1016/j.cytogfr.2009.11.001
21. Bakst R, Wong R. Mechanisms of Perineural Invasion. *Journal of Neurological Surgery Part B: Skull Base*. 03/10 2016;77doi:10.1055/s-0036-1571835
22. Azam SH, Pecot CV. Cancer's got nerve: Schwann cells drive perineural invasion. *J Clin Invest*. 2016;126(4):1242-1244. doi:10.1172/JCI86801
23. Amit M, Na'ara S, Gil Z. Mechanisms of cancer dissemination along nerves. *Nature Reviews Cancer*. 2016/06/01 2016;16(6):399-408. doi:10.1038/nrc.2016.38
24. Olar A, He D, Florentin D, Ding Y, Wheeler T, Ayala G. Biological correlates of prostate cancer perineural invasion diameter. *Human pathology*. Jul 2014;45(7):1365-9. doi:10.1016/j.humpath.2014.02.011
25. Reavis HD, Chen HI, Drapkin R. Tumor Innervation: Cancer Has Some Nerve. *Trends in Cancer*. 2020/08/14/ 2020;doi:<https://doi.org/10.1016/j.trecan.2020.07.005>
26. Yates C. Influence of Tumor Microenvironment on the Molecular Regulation of Prostate Cancer Progression. 2009.
27. Li X, Ma Q, Xu Q, et al. SDF-1/CXCR4 signaling induces pancreatic cancer cell invasion and epithelial-mesenchymal transition in vitro through non-canonical activation of Hedgehog pathway. *Cancer letters*. 2012;322(2):169-176. doi:10.1016/j.canlet.2012.02.035
28. Zhao C-M, Hayakawa Y, Kodama Y, et al. Denervation suppresses gastric tumorigenesis. *Science Translational Medicine*. 2014;6(250):250ra115. doi:10.1126/scitranslmed.3009569
29. Samarelli AV, Riccitelli E, Bizzozero L, et al. Neuroligin 1 induces blood vessel maturation by cooperating with the  $\alpha 6$  integrin. *The Journal of biological chemistry*. Jul 11 2014;289(28):19466-76. doi:10.1074/jbc.M113.530972
30. Graziano S, Marchiò S, Bussolino F, Arese M. A peptide from the extracellular region of the synaptic protein  $\alpha$  Neurexin stimulates angiogenesis and the vascular specific tyrosine kinase Tie2. *Biochemical and Biophysical Research Communications*. 2013/03/22/ 2013;432(4):574-579. doi:<https://doi.org/10.1016/j.bbrc.2013.02.045>
31. Marco Arese FB, Margherita Pergolizzi, Laura Bizzozero, Davide Pascal. Tumor progression: the neuronal input Review. *Annals of Translational Medicine*. 2018;6(5):89. doi:10.21037/atm.2018.01.01
32. Tan X, Huang Z, Li X. Long Non-Coding RNA MALAT1 Interacts With miR-204 to Modulate Human Hilar Cholangiocarcinoma Proliferation, Migration, and Invasion by Targeting CXCR4. *Journal of cellular biochemistry*. Nov 2017;118(11):3643-3653. doi:10.1002/jcb.25862
33. Zheng HT, Shi DB, Wang YW, et al. High expression of lncRNA MALAT1 suggests a biomarker of poor prognosis in colorectal cancer. *International journal of clinical and experimental pathology*. 2014;7(6):3174-81.

34. Hirata H, Hinoda Y, Shahryari V, et al. Long Noncoding RNA MALAT1 Promotes Aggressive Renal Cell Carcinoma through Ezh2 and Interacts with miR-205. *Cancer Res.* Apr 1 2015;75(7):1322-31. doi:10.1158/0008-5472.Can-14-2931
35. Yu EH, Tu HF, Wu CH, Yang CC, Chang KW. MicroRNA-21 promotes perineural invasion and impacts survival in patients with oral carcinoma. *Journal of the Chinese Medical Association : JCMA.* Jun 2017;80(6):383-388. doi:10.1016/j.jcma.2017.01.003
36. Sousa LO, Sobral LM, Matsumoto CS, et al. Lymph node or perineural invasion is associated with low miR-15a, miR-34c and miR-199b levels in head and neck squamous cell carcinoma. *BBA Clinical.* 2016/12/01/2016;6:159-164. doi:<https://doi.org/10.1016/j.bbacli.2016.11.001>
37. Sim J, Ahn H, Abdul R, et al. High MicroRNA-370 Expression Correlates with Tumor Progression and Poor Prognosis in Breast Cancer. *Journal of breast cancer.* Dec 2015;18(4):323-8. doi:10.4048/jbc.2015.18.4.323
38. Demir IE, Boldis A, Pfitzinger PL, et al. Investigation of Schwann cells at neoplastic cell sites before the onset of cancer invasion. *Journal of the National Cancer Institute.* Aug 2014;106(8)doi:10.1093/jnci/dju184
39. Deborde S, Omelchenko T, Lyubchik A, et al. Schwann cells induce cancer cell dispersion and invasion. *J Clin Invest.* Apr 1 2016;126(4):1538-54. doi:10.1172/jci82658
40. Azam SH, Pecot CV. Cancer's got nerve: Schwann cells drive perineural invasion. *J Clin Invest.* Apr 1 2016;126(4):1242-4. doi:10.1172/jci86801
41. Tanaka K, Okugawa Y, Toiyama Y, et al. Brain-Derived Neurotrophic Factor (BDNF)-Induced Tropomyosin-Related Kinase B (Trk B) Signaling Is a Potential Therapeutic Target for Peritoneal Carcinomatosis Arising from Colorectal Cancer. *PloS one.* 2014;9(5):e96410. doi:10.1371/journal.pone.0096410
42. Shan C, Wei J, Hou R, et al. Schwann cells promote EMT and the Schwann-like differentiation of salivary adenoid cystic carcinoma cells via the BDNF/TrkB axis. *Oncology reports.* Jan 2016;35(1):427-35. doi:10.3892/or.2015.4366
43. Ferdoushi A, Li X, Griffin N, et al. Schwann Cell Stimulation of Pancreatic Cancer Cells: A Proteomic Analysis. *Frontiers in oncology.* 2020;10:1601. doi:10.3389/fonc.2020.01601
44. Yoko Fujii-Nishimura KY, Yohei Masugi, Junya Douguchi, Yutaka Kurebayashi, Naoto Kubota, Hidenori Ojima, Minoru Kitago, Masahiro Shinoda, Akinori Hashiguchi, Michiie Sakamoto. Mesenchymal–epithelial transition of pancreatic cancer cells at perineural invasion sites is induced by Schwann cells. Research Article. *Pathology International.* 19 February 2018 2018;68(4):214-223. doi:doi.org/10.1111/pin.12641
45. Jeffus SK, Gehlot A, Holthoff E, et al. A fibromyxoid stromal response is associated with an infiltrative tumor morphology, perineural invasion, and lymph node metastasis in squamous cell carcinoma of the vulva. *The American journal of surgical pathology.* Sep 2015;39(9):1226-33. doi:10.1097/pas.0000000000000486
46. Mitani T, Harada N, Nakano Y, Inui H, Yamaji R. Coordinated action of hypoxia-inducible factor-1 $\alpha$  and  $\beta$ -catenin in androgen receptor signaling. *The Journal of biological chemistry.* 2012;287(40):33594-33606. doi:10.1074/jbc.M112.388298
47. Ramakrishnan R, Pena-Martinez P, Agarwal P, et al. CXCR4 Signaling Has a CXCL12-Independent Essential Role in Murine MLL-AF9-Driven Acute Myeloid Leukemia. *Cell Rep.* May 26 2020;31(8):107684. doi:10.1016/j.celrep.2020.107684



48. Gummy LF, Bampton ETW, Tolkovsky AM. Hyperglycaemia inhibits Schwann cell proliferation and migration and restricts regeneration of axons and Schwann cells from adult murine DRG. *Molecular and Cellular Neuroscience*. 2008/02/01/2008;37(2):298-311. doi:<https://doi.org/10.1016/j.mcn.2007.10.004>
49. Li J, Ma J, Han L, et al. Hyperglycemic tumor microenvironment induces perineural invasion in pancreatic cancer. *Cancer Biol Ther*. 2015;16(6):912-921. doi:10.1080/15384047.2015.1040952
50. Armaiz-Pena GN, Cole SW, Lutgendorf SK, Sood AK. Neuroendocrine influences on cancer progression. *Brain Behav Immun*. 2013;30 Suppl(Suppl):S19-S25. doi:10.1016/j.bbi.2012.06.005
51. Cavel O, Shomron O, Shabtay A, et al. Endoneurial macrophages induce perineural invasion of pancreatic cancer cells by secretion of GDNF and activation of RET tyrosine kinase receptor. *Cancer Res*. Nov 15 2012;72(22):5733-43. doi:10.1158/0008-5472.Can-12-0764
52. Tang D, Wang D, Yuan Z, et al. Persistent activation of pancreatic stellate cells creates a microenvironment favorable for the malignant behavior of pancreatic ductal adenocarcinoma. *International journal of cancer*. Mar 1 2013;132(5):993-1003. doi:10.1002/ijc.27715
53. Yao J, Li W-Y, Li S-G, Feng X-S, Gao S-G. Midkine promotes perineural invasion in human pancreatic cancer. *World Journal of Gastroenterology: WJG*. 2014;20(11):3018.
54. Yao J, Li W-Y, Gao S-G. The advances of Midkine with peripheral invasion in pancreatic cancer. *American journal of cancer research*. 2015;5(9):2912.
55. Yao J, Zhang L-L, Huang X-M, Li W-Y, Gao S-G. Pleiotrophin and N-syndecan promote perineural invasion and tumor progression in an orthotopic mouse model of pancreatic cancer. *World journal of gastroenterology*. 2017;23(21):3907.
56. Yao J, Li W-Y, Li S-G, Feng X-S, Gao S-G. Midkine promotes perineural invasion in human pancreatic cancer. *World J Gastroenterol*. 2014;20(11):3018-3024. doi:10.3748/wjg.v20.i11.3018
57. Chen P, Cescon M, Bonaldo P. The Role of Collagens in Peripheral Nerve Myelination and Function. *Molecular neurobiology*. Aug 2015;52(1):216-25. doi:10.1007/s12035-014-8862-y
58. Guo D, Sun W, Zhu LEI, et al. Knockdown of BDNF suppressed invasion of HepG2 and HCCLM3 cells, a mechanism associated with inactivation of RhoA or Rac1 and actin skeleton disorganization. <https://doi.org/10.1111/j.1600-0463.2011.02855.x>. *APMIS*. 2012/06/01 2012;120(6):469-476. doi:<https://doi.org/10.1111/j.1600-0463.2011.02855.x>
59. Cossa G, Gatti L, Cassinelli G, Lanzi C, Zaffaroni N, Perego P. Modulation of sensitivity to antitumor agents by targeting the MAPK survival pathway. *Current pharmaceutical design*. 2013;19(5):883-94.
60. Gao H, Peng C, Liang B, et al.  $\beta 6$  integrin induces the expression of metalloproteinase-3 and metalloproteinase-9 in colon cancer cells via ERK-ETS1 pathway. *Cancer letters*. Nov 28 2014;354(2):427-37. doi:10.1016/j.canlet.2014.08.017
61. Xu L, Hou Y, Tu G, et al. Nuclear Drosha enhances cell invasion via an EGFR-ERK1/2-MMP7 signaling pathway induced by dysregulated miRNA-622/197 and their targets LAMC2 and CD82 in gastric cancer. *Cell Death & Disease*. 2017/03/01 2017;8(3):e2642-e2642. doi:10.1038/cddis.2017.5
62. Xiang T, Xia X, Yan W. Expression of Matrix Metalloproteinases-2/-9 is Associated With Microvessel Density in Pancreatic Cancer. *American journal of*



- therapeutics*. Jul/Aug 2017;24(4):e431-e434. doi:10.1097/mjt.0000000000000424
63. Klimczak-Bitner AA, Kordek R, Bitner J, Musiał J, Szemraj J. Expression of MMP9, SERPINE1 and miR-134 as prognostic factors in esophageal cancer. *Oncol Lett*. Nov 2016;12(5):4133-4138. doi:10.3892/ol.2016.5211
64. Juchniewicz A, Kowalczyk O, Milewski R, et al. MMP-10, MMP-7, TIMP-1 and TIMP-2 mRNA expression in esophageal cancer. *Acta biochimica Polonica*. 2017;64(2):295-299. doi:10.18388/abp.2016\_1408
65. Li Q, Wang Y, Lai Y, Xu P, Yang Z. HspB5 correlates with poor prognosis in colorectal cancer and prompts epithelial-mesenchymal transition through ERK signaling. *PloS one*. 2017;12(8):e0182588. doi:10.1371/journal.pone.0182588
66. Klupp F, Neumann L, Kahlert C, et al. Serum MMP7, MMP10 and MMP12 level as negative prognostic markers in colon cancer patients. *BMC cancer*. Jul 18 2016;16:494. doi:10.1186/s12885-016-2515-7
67. Fu Y-Z, Su S, Gao Y-Q, et al. Human Cytomegalovirus Tegument Protein UL82 Inhibits STING-Mediated Signaling to Evade Antiviral Immunity. *Cell Host & Microbe*. 2017/02/08/ 2017;21(2):231-243. doi:<https://doi.org/10.1016/j.chom.2017.01.01>
68. Han Y, Wu Z, Wu T, et al. Tumor-suppressive function of long non-coding RNA MALAT1 in glioma cells by downregulation of MMP2 and inactivation of ERK/MAPK signaling. *Cell Death Dis*. Mar 3 2016;7(3):e2123. doi:10.1038/cddis.2015.407
69. Liu D, Duan W, Guo H, Xu X, Bai Y. Meta-analysis of associations between polymorphisms in the promoter regions of matrix metalloproteinases and the risk of colorectal cancer. *International Journal of Colorectal Disease*. 2011/05/03 2011;26(9):1099. doi:10.1007/s00384-011-1198-4
70. Du J, Zhang L. Analysis of salivary microRNA expression profiles and identification of novel biomarkers in esophageal cancer. *Oncol Lett*. Aug 2017;14(2):1387-1394. doi:10.3892/ol.2017.6328
71. Nigri J, Gironella M, Bressy C, et al. PAP/REG3A favors perineural invasion in pancreatic adenocarcinoma and serves as a prognostic marker. *Cellular and molecular life sciences : CMLS*. Nov 2017;74(22):4231-4243. doi:10.1007/s00018-017-2579-9
72. Mallini P, Lennard T, Kirby J, Meeson A. Epithelial-to-mesenchymal transition: What is the impact on breast cancer stem cells and drug resistance. *Cancer Treatment Reviews*. 2014/04/01/ 2014;40(3):341-348. doi:<https://doi.org/10.1016/j.ctrv.2013.09.008>
73. He Q, Zhou X, Li S, et al. MicroRNA-181a suppresses salivary adenoid cystic carcinoma metastasis by targeting MAPK–Snai2 pathway. *Biochimica et Biophysica Acta (BBA) - General Subjects*. 2013/11/01/ 2013;1830(11):5258-5266. doi:<https://doi.org/10.1016/j.bbagen.2013.07.028>
74. Scanlon CS, Banerjee R, Inglehart RC, et al. Galanin modulates the neural niche to favour perineural invasion in head and neck cancer. *Nature communications*. Apr 28 2015;6:6885. doi:10.1038/ncomms7885
75. Tilan J, Kitlinska J. Neuropeptide Y (NPY) in tumor growth and progression: Lessons learned from pediatric oncology. *Neuropeptides*. Feb 2016;55:55-66. doi:10.1016/j.npep.2015.10.005
76. Huang C, Li Y, Guo Y, et al. MMP1/PAR1/SP/NK1R paracrine loop modulates early perineural invasion of pancreatic cancer cells. *Theranostics*. 2018;8(11):3074-3086. doi:10.7150/thno.24281

77. Shen WR, Wang YP, Chang JY, Yu SY, Chen HM, Chiang CP. Perineural invasion and expression of nerve growth factor can predict the progression and prognosis of oral tongue squamous cell carcinoma. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. Apr 2014;43(4):258-64. doi:10.1111/jop.12133
78. Mancino M, Ametller E, Gascón P, Almendro V. The neuronal influence on tumor progression. *Biochimica et biophysica acta*. Dec 2011;1816(2):105-18. doi:10.1016/j.bbcan.2011.04.005
79. Doebele RC, Davis LE, Vaishnavi A, et al. An Oncogenic NTRK Fusion in a Patient with Soft-Tissue Sarcoma with Response to the Tropomyosin-Related Kinase Inhibitor LOXO-101. *Cancer discovery*. Oct 2015;5(10):1049-57. doi:10.1158/2159-8290.Cd-15-0443
80. Bapat AA, Munoz RM, Von Hoff DD, Han H. Blocking Nerve Growth Factor Signaling Reduces the Neural Invasion Potential of Pancreatic Cancer Cells. *PloS one*. 2016;11(10):e0165586. doi:10.1371/journal.pone.0165586
81. Jia S, Wang W, Hu Z, et al. BDNF mediated TrkB activation contributes to the EMT progression and the poor prognosis in human salivary adenoid cystic carcinoma. *Oral Oncology*. 2015/01/01/ 2015;51(1):64-70. doi:<https://doi.org/10.1016/j.oraloncology.2014.10.008>
82. Okugawa Y, Tanaka K, Inoue Y, et al. Brain-derived neurotrophic factor/tropomyosin-related kinase B pathway in gastric cancer. *British journal of cancer*. Jan 15 2013;108(1):121-30. doi:10.1038/bjc.2012.499
83. Amit M, Na'ara S, Sharma K, et al. Elective Neck Dissection in Patients With Head and Neck Adenoid Cystic Carcinoma: An International Collaborative Study. *Annals of Surgical Oncology*. 2015/04/01 2015;22(4):1353-1359. doi:10.1245/s10434-014-4106-7
84. Gao L, Bo H, Wang Y, Zhang J, Zhu M. Neurotrophic Factor Artemin Promotes Invasiveness and Neurotrophic Function of Pancreatic Adenocarcinoma In Vivo and In Vitro. *Pancreas*. 2015;44(1)
85. Bakst RL, Lee N, He S, et al. Radiation impairs perineural invasion by modulating the nerve microenvironment. *PloS one*. 2012;7(6):e39925. doi:10.1371/journal.pone.0039925
86. He S, Chen C-H, Chernichenko N, et al. GFR $\alpha$ 1 released by nerves enhances cancer cell perineural invasion through GDNF-RET signaling. *Proc Natl Acad Sci U S A*. 2014;111(19):E2008-E2017. doi:10.1073/pnas.1402944111
87. DeLancey JO, Wood DP, Jr., He C, et al. Evidence of perineural invasion on prostate biopsy specimen and survival after radical prostatectomy. *Urology*. Feb 2013;81(2):354-7. doi:10.1016/j.urology.2012.09.034
88. Chuang J-Y, Tsai C-F, Chang S-W, et al. Glial cell line-derived neurotrophic factor induces cell migration in human oral squamous cell carcinoma. *Oral Oncology*. 2013/12/01/ 2013;49(12):1103-1112. doi:<https://doi.org/10.1016/j.oraloncology.2013.08.009>
89. Gao L, Bo H, Wang Y, Zhang J, Zhu M. Neurotrophic factor artemin promotes invasiveness and neurotrophic function of pancreatic adenocarcinoma in vivo and in vitro. *Pancreas*. 2015;44(1):134.
90. Gregory E, Dugan R, David G, Song YH. The biology and engineered modeling strategies of cancer-nerve crosstalk. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer*. 2020/12/01/ 2020;1874(2):188406. doi:<https://doi.org/10.1016/j.bbcan.2020.188406>

91. Warren TA, Nagle CM, Bowman J, Panizza BJ. The Natural History and Treatment Outcomes of Perineural Spread of Malignancy within the Head and Neck. *Journal of neurological surgery Part B, Skull base*. Apr 2016;77(2):107-12. doi:10.1055/s-0036-1579777
92. Tian X, Gu T, Patel S, Bode AM, Lee MH, Dong Z. CRISPR/Cas9 - An evolving biological tool kit for cancer biology and oncology. *NPJ precision oncology*. 2019;3:8. doi:10.1038/s41698-019-0080-7
93. Koonin EV, Makarova KS. CRISPR-Cas: evolution of an RNA-based adaptive immunity system in prokaryotes. *RNA biology*. May 2013;10(5):679-86. doi:10.4161/rna.24022
94. Bapat AA, Hostetter G, Von Hoff DD, Han H. Perineural invasion and associated pain in pancreatic cancer. *Nature reviews Cancer*. Sep 23 2011;11(10):695-707. doi:10.1038/nrc3131
95. Deshmukh SD, Willmann JK, Jeffrey RB. Pathways of extrapancreatic perineural invasion by pancreatic adenocarcinoma: evaluation with 3D volume-rendered MDCT imaging. *AJR American journal of roentgenology*. Mar 2010;194(3):668-74. doi:10.2214/ajr.09.3285
96. Demir IE, Friess H, Ceyhan GO. Nerve-cancer interactions in the stromal biology of pancreatic cancer. *Front Physiol*. 2012;3:97-97. doi:10.3389/fphys.2012.00097
97. Yang YH, Liu JB, Gui Y, Lei LL, Zhang SJ. Relationship between autophagy and perineural invasion, clinicopathological features, and prognosis in pancreatic cancer. *World J Gastroenterol*. Oct 28 2017;23(40):7232-7241. doi:10.3748/wjg.v23.i40.7232
98. Schorn S, Demir IE, Haller B, et al. The influence of neural invasion on survival and tumor recurrence in pancreatic ductal adenocarcinoma - A systematic review and meta-analysis. *Surgical oncology*. Mar 2017;26(1):105-115. doi:10.1016/j.suronc.2017.01.007
99. Jiang N, Deng JY, Liu Y, Ke B, Liu HG, Liang H. Incorporation of perineural invasion of gastric carcinoma into the 7th edition tumor-node-metastasis staging system. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. Sep 2014;35(9):9429-36. doi:10.1007/s13277-014-2258-5
100. España-Ferrufino A, Lino-Silva LS, Salcedo-Hernández RA. Extramural Perineural Invasion in pT3 and pT4 Gastric Carcinomas. *Journal of pathology and translational medicine*. Mar 2018;52(2):79-84. doi:10.4132/jptm.2017.11.01
101. Aurello P, Berardi G, Tierno SM, et al. Influence of perineural invasion in predicting overall survival and disease-free survival in patients With locally advanced gastric cancer. *The American Journal of Surgery*. 2017/04/01/ 2017;213(4):748-753. doi:<https://doi.org/10.1016/j.amjsurg.2016.05.022>
102. Deng J, You Q, Gao Y, et al. Prognostic value of perineural invasion in gastric cancer: a systematic review and meta-analysis. *PloS one*. 2014;9(2):e88907. doi:10.1371/journal.pone.0088907
103. Tollefson MK, Karnes RJ, Kwon ED, et al. Prostate cancer Ki-67 (MIB-1) expression, perineural invasion, and gleason score as biopsy-based predictors of prostate cancer mortality: the Mayo model. *Mayo Clinic proceedings*. Mar 2014;89(3):308-18. doi:10.1016/j.mayocp.2013.12.001
104. Zhang LJ, Wu B, Zha ZL, et al. Perineural invasion as an independent predictor of biochemical recurrence in prostate cancer following radical prostatectomy or radiotherapy: a systematic review and meta-analysis. *BMC urology*. Feb 1 2018;18(1):5. doi:10.1186/s12894-018-0319-6
105. Lubig S, Thiesler T, Müller S, Vorreuther R, Leipner N, Kristiansen G.

Quantitative perineural invasion is a prognostic marker in prostate cancer. *Pathology*. 2018/04/01/ 2018;50(3):298-304. doi:<https://doi.org/10.1016/j.pathol.2017.09.013>

106. Feo C, Cossu M, Ginesu G, et al. Perineural infiltration as a prognostic factor in surgically treated gallbladder cancer: A single center experience and literature review. *Annali italiani di chirurgia*. 10/04 2017;6

107. Murakami Y, Uemura K, Sudo T, et al. Perineural Invasion in Extrahepatic Cholangiocarcinoma: Prognostic Impact and Treatment Strategies. *Journal of Gastrointestinal Surgery*. 2013/08/01 2013;17(8):1429-1439. doi:10.1007/s11605-013-2251-0

108. Wellner UF, Shen Y, Keck T, Jin W, Xu Z. The survival outcome and prognostic factors for distal cholangiocarcinoma following surgical resection: a meta-analysis for the 5-year survival. *Surgery today*. Mar 2017;47(3):271-279. doi:10.1007/s00595-016-1362-0

109. Quintana JM, González N, Lázaro S, et al. Predictors of 1- and 2-year mortality in patients with rectal cancer. *Colorectal disease : the official journal of the Association of Coloproctology of Great Britain and Ireland*. Aug 2018;20(8):676-687. doi:10.1111/codi.14250

110. Zare Bandamiri M, Fararouei M, Zohourinia S, Daneshi N, Dianatinasab M. Risk Factors Predicting Colorectal Cancer Recurrence Following Initial Treatment: A 5-year Cohort Study. *Asian Pacific journal of cancer prevention: APJCP*. 09/18 2017;18:2465-2470. doi:10.22034/APJCP.2017.18.9.2465

111. Huang Y, He L, Dong D, et al. Individualized prediction of perineural invasion in colorectal cancer: development and validation of a radiomics prediction model. *Chinese journal of cancer research = Chung-kuo yen cheng yen chiu*. Feb

2018;30(1):40-50. doi:10.21147/j.issn.1000-9604.2018.01.05

112. Milica Stojkovic Lalosevic TM, Marjan Micev, Mirjana Stojkovic, Sanja Dragasevic, Milos Stulic, Ivan Rankovic, Vladimir Dugalic, Zoran Krivokapic, Aleksandra Pavlovic Markovic Perineural invasion as a prognostic factor in patients with stage I-III rectal cancer – 5-year follow up. *World J Gastrointest Oncol*. May 15, 2020; 12(5): 592-600doi:10.4251/wjgo.v12.i5.592

113. Amit M, Eran A, Billan S, et al. Perineural Spread in Noncutaneous Head and Neck Cancer: New Insights into an Old Problem. *Journal of neurological surgery Part B, Skull base*. Apr 2016;77(2):86-95. doi:10.1055/s-0036-1571834

114. Bakst RL, Glastonbury CM, Parvathaneni U, Katabi N, Hu KS, Yom SS. Perineural Invasion and Perineural Tumor Spread in Head and Neck Cancer. *International Journal of Radiation Oncology\*Biophysics*. 2019/04/01/ 2019;103(5):1109-1124. doi:<https://doi.org/10.1016/j.ijrobp.2018.12.009>

115. Turner FE, Broad S, Khanim FL, et al. Slug regulates integrin expression and cell proliferation in human epidermal keratinocytes. *The Journal of biological chemistry*. Jul 28 2006;281(30):21321-31. doi:10.1074/jbc.M509731200

116. Zhang L, Fang P, Chai C, et al. Galanin expression is down-regulated in patients with gastric cancer. *J Int Med Res*. 2019;47(3):1241-1249. doi:10.1177/0300060518819382

117. Körner M, Waser B, Reubi JC. High Expression of Neuropeptide Y Receptors in Tumors of the Human Adrenal Gland and Extra-Adrenal Paraganglia. *Clinical Cancer Research*. 2004;10(24):8426. doi:10.1158/1078-0432.CCR-04-0821

118. Wu JQ, Jiang N, Yu B. Mechanisms of action of neuropeptide Y on stem cells and



its potential applications in orthopaedic disorders. *World journal of stem cells*. Sep 26 2020;12(9):986-1000.

doi:10.4252/wjsc.v12.i9.986

119. Li S, Sun Y, Gao D. Role of the nervous system in cancer metastasis. *Oncol Lett*. 2013;5(4):1101-1111.

doi:10.3892/ol.2013.1168

120. Chen X-Y, Ru G-Q, Ma Y-Y, et al. High expression of substance P and its receptor neurokinin-1 receptor in colorectal cancer is associated with tumor progression and prognosis. *Onco Targets Ther*. 2016;9:3595-3602.

doi:10.2147/OTT.S102356

121. Campenot RB. NGF and the local control of nerve terminal growth. *Journal of neurobiology*. Jun 1994;25(6):599-611.

doi:10.1002/neu.480250603

122. Adriaenssens E, Vanhecke E, Saule P, et al. Nerve Growth Factor Is a Potential Therapeutic Target in Breast Cancer. *Cancer Research*. 2008;68(2):346.

doi:10.1158/0008-5472.CAN-07-1183

123. Shishkina TV, Mishchenko TA, Mitroshina EV, et al. Glial cell line-derived neurotrophic factor (GDNF) counteracts hypoxic damage to hippocampal neural network function in vitro. *Brain Research*. 2018/01/01/ 2018;1678:310-321.

doi:<https://doi.org/10.1016/j.brainres.2017.10.023>

124. Saffrey MJ. Enteric Nervous System: Neurotrophic Factors☆. *Reference Module in Neuroscience and Biobehavioral Psychology*. Elsevier; 2017.

125. Dufour S, Broders-Bondon F, Bondurand N. Chapter 13 -  $\beta$ 1-Integrin Function and Interplay during Enteric Nervous System Development. In: Pruszk J, ed. *Neural Surface Antigens*. Academic Press; 2015:153-166.

126. Sawai H, Okada Y, Kazanjian K, et al. The G691S RET Polymorphism Increases Glial Cell Line-Derived Neurotrophic Factor-Induced Pancreatic Cancer Cell

Invasion by Amplifying Mitogen-Activated Protein Kinase Signaling. *Cancer Research*. 2005;65(24):11536. doi:10.1158/0008-5472.CAN-05-2843

127. Saarma M. GFL Neurotrophic Factors: Physiology and Pharmacology. *Encyclopedia of Neuroscience*. 01/01 2010:711-720. doi:10.1016/B978-008045046-9.00501-5

128. Schachner M, Leshchyn'ska I, Sytnyk V. Functions of the Neural Cell Adhesion Molecule (NCAM) at the Synapse. 2017.

129. Phimister E, Kiely F, Kemshead JT, Patel K. Expression of neural cell adhesion molecule (NCAM) isoforms in neuroblastoma. *J Clin Pathol*. Jul 1991;44(7):580-5. doi:10.1136/jcp.44.7.580

130. Schachner M, Leshchyn'ska I, Sytnyk V. Neural Cell Adhesion Molecules and Synapse Regulation. In: Squire LR, ed. *Encyclopedia of Neuroscience*. Academic Press; 2009:91-96.

131. Merdad A, Karim S, Schulten HJ, et al. Expression of matrix metalloproteinases (MMPs) in primary human breast cancer: MMP-9 as a potential biomarker for cancer invasion and metastasis. *Anticancer research*. Mar 2014;34(3):1355-66.

132. Nagase H, Visse R, Murphy G. Structure and function of matrix metalloproteinases and TIMPs. *Cardiovascular Research*. 2006;69(3):562-573. doi:10.1016/j.cardiores.2005.12.002

133. Murphy G. Matrix Metalloproteinases. In: Bradshaw RA, Stahl PD, eds. *Encyclopedia of Cell Biology*. Academic Press; 2016:621-629.

134. Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA. Chronic inflammation and cytokines in the tumor microenvironment. *J Immunol Res*. 2014;2014:149185-149185. doi:10.1155/2014/149185

135. Carswell-Richards EA, Williamson BD. A man of vision and the discovery of

tumor necrosis factor. *Cancer Immun.* 2012;12:4-4.

136. Muthukumaran N, Miletti-González KE, Ravindranath AK, Rodríguez-Rodríguez L. Tumor Necrosis Factor- $\alpha$  Differentially Modulates CD44 Expression in Ovarian Cancer Cells. *Molecular Cancer Research.* 2006;4(8):511. doi:10.1158/1541-7786.MCR-05-0232

137. Cruceriu D, Baldasici O, Balacescu O, Berindan-Neagoe I. The dual role of tumor necrosis factor-alpha (TNF- $\alpha$ ) in breast cancer: molecular insights and therapeutic approaches. *Cellular oncology (Dordrecht).* Feb 2020;43(1):1-18. doi:10.1007/s13402-019-00489-1

138. Tanabe K, Matsushima-Nishiwaki R, Yamaguchi S, Iida H, Dohi S, Kozawa O. Mechanisms of tumor necrosis factor-alpha-induced interleukin-6 synthesis in glioma cells. *J Neuroinflammation.* 2010;7:16-16. doi:10.1186/1742-2094-7-16

139. Culig Z. Proinflammatory cytokine interleukin-6 in prostate carcinogenesis. *Am J Clin Exp Urol.* 2014;2(3):231-238.

140. Jones SA, Takeuchi T, Aletaha D, Smolen J, Choy EH, McInnes I. Interleukin 6: The biology behind the therapy. *Considerations in Medicine.* 2018;2(1):2. doi:10.1136/conmed-2018-000005

141. Heikkilä K, Ebrahim S, Lawlor DA. Systematic review of the association between circulating interleukin-6 (IL-6) and cancer.

*European Journal of Cancer.* 2008/05/01/2008;44(7):937-945.

doi:<https://doi.org/10.1016/j.ejca.2008.02.047>

142. Morrison CD, Parvani JG, Schiemann WP. The relevance of the TGF- $\beta$  Paradox to EMT-MET programs. *Cancer letters.* Nov 28 2013;341(1):30-40. doi:10.1016/j.canlet.2013.02.048

143. Bierie B, Moses HL. TGF-beta and cancer. *Cytokine & growth factor reviews.* Feb-Apr 2006;17(1-2):29-40. doi:10.1016/j.cytogfr.2005.09.006

144. Zhao Y, Ma J, Fan Y, et al. TGF- $\beta$  transactivates EGFR and facilitates breast cancer migration and invasion through canonical Smad3 and ERK/Sp1 signaling pathways. *Molecular oncology.* Mar 2018;12(3):305-321. doi:10.1002/1878-0261.12162

145. Schottelius AJ, Mayo MW, Sartor RB, Baldwin AS, Jr. Interleukin-10 signaling blocks inhibitor of kappaB kinase activity and nuclear factor kappaB DNA binding. *The Journal of biological chemistry.* Nov 5 1999;274(45):31868-74. doi:10.1074/jbc.274.45.31868

146. Wang Y, Liu X-H, Li Y-H, Li O. The paradox of IL-10-mediated modulation in cervical cancer. *Biomed Rep.* 2013;1(3):347-351. doi:10.3892/br.2013.69