RESEARCH ARTICLE

Generation of Genetically Modified Rats Using CRISPR/Cas9 Genome-Editing System to Reveal Novel Vitamin D Actions

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Abstract

We previously revealed that the anti-proliferative activity of 25-hydroxyvitamin D₃ (25(OH)D₃) in human prostate PZ-HPV-7 cells depends on the direct action of 25(OH)D₃ through the vitamin D receptor (VDR). We then attempted to confirm the direct action of 25(OH)D₃ *in vivo* using *Cyp27b1* knockout (KO) mice. Daily administration of 25(OH)D3 at 250 µg/kg bw/day rescued the rachitic conditions in *Cyp27b1* KO mice. The plasma levels of calcium, phosphorus, and parathyroid hormonea as well as the bone mineral density and female sexual cycle were normalized via the administration of 25(OH)D₃. These results strongly suggest the direct action of 25(OH)D₃. However, to our surprise, normal levels of 1 α ,25(OH)₂D₃ were detected in the plasma of *Cyp27b1* KO mice, probably due to Cyp27a1, which has a weak 1 α -hydroxylation activity toward 25(OH)D₃.

Next, we generated a novel *in vivo* system using genetically modified (GM) rats deficient in the *Cyp27b1* or *Vdr* gene to reveal the molecular mechanisms of vitamin D action. Human type II rickets model rats with mutant *Vdr* (R270L), which recognizes 1,25(OH)₂D₃ with an affinity equivalent to that of 25(OH)D₃, were also generated. *Cyp27b1*-knockout (KO), *Vdr*-KO, and *Vdr* (R270L) rats showed symptoms of rickets, including growth retardation and abnormal bone formation. Among these model animals, *Cyp27b1*-KO rats had notably low levels of calcium in the blood and the most severe growth retardation, while *Vdr*-KO rats showed abnormal skin formation and alopecia. Administration of 25(OH)D₃ restored rickets symptoms in *Cyp27b1*-KO and *Vdr* (R270L) rats. As shown in *Cyp27b1*-KO mice, 1,25(OH)₂D₃ was also synthesized in *Cyp27b1*-KO rats. In contrast, the effects of 25(OH)D₃ on *Vdr* (R270L) rats strongly suggest that 25(OH)D₃ exerts a direct action via VDR-genomic pathways. These results suggest that our novel *in vivo* system containing three different types of GM rats is useful for elucidating the molecular mechanism of vitamin D action.

Key words: vitamin D, vitamin D receptor, genome editing, CYP27B1, rickets, genome editing

Introduction

Vitamin D exerts various biological activities, including calcemic, osteogenic, and anti-cancer activities, along with the generation of immune responses¹. Vitamin D₃ is an animal vitamin D form obtained from diet via intestinal absorption or through biosynthesis from 7-dehydrocholesterol by UV irradiation of the skin. Vitamin D_3 is sequentially converted to biologically active 1,25-dihydroxyvitamin D_3 (1,25(OH)₂ D_3) in the body. Moreover, 25-hydroxyvitamin D₃ $(25(OH)D_3)$, which is converted from vitamin D3 via the hydroxylation of C-25 by CYP2R1

and CYP27A1 in the liver, circulates through the body by binding to vitamin D-binding protein (DBP). DBP-bound 25(OH)D₃ is taken into the proximal tubular cells in the kidney through megalin-mediated endocytosis of DBP, and then converted to $1,25(OH)_2D_3$ via 1α -hydroxylation by CYP27B1, which is a key enzyme in the production of $1,25(OH)_2D_3$ ² (Fig. 1). Furthermore, $1,25(OH)_2D_3$ is a ligand with a high affinity for the vitamin D receptor (VDR), whose binding affinity is approximately 1000-fold higher than that of 25(OH)D₃. The transcriptional activity of

 $1,25(OH)_2D_3$ by binding to VDR is a classic molecular mechanism of vitamin D, which is observed in tissues highly expressing VDR, such as the intestine, kidneys, and bones. Once $1,25(OH)_2D_3$ binds to VDR, it forms a heterodimer with 9-cis retinoid X receptor (RXR) to regulate the expression of target genes by binding to the vitamin D-responsive element (VDRE) in the promoter region of target genes involved in calcium homeostasis and osteogenesis ³. This transcriptional activity is also recognized as a VDRdependent genomic action of 1,25(OH)₂D₃. Dysfunction of the genomic action of 1,25(OH)₂D₃ causes typical bone disorders, such as osteomalacia and rickets associated with hypocalcemia and hyperparathyroidism. Recently, novel molecular mechanisms of vitamin D have been reported. Hence, there may be five types of molecular actions of vitamin D: (1)VDR-dependent genomic or non-genomic effects of 1,25(OH)₂D₃ (VDR- $1,25(OH)_2D_3$) ^{4, 5}, (2) VDR-independent effects of $1,25(OH)_2D_3$ (non VDR- $1,25(OH)_2D_3)^6$, (3) VDR-dependent effects of 25(OH)D₃ (VDR-25(OH)D₃)^{7,8}, (4) VDRindependent effects of 25(OH)D₃ (non VDR- $25(OH)D_3)$ ⁹, and (5) ligand-independent effects of VDR (VDR-no ligand) 10, 11 (Fig. 1 and Table 1). The elucidation of these vitamin D actions and biological activities is useful for the discovery of new therapeutic targets, because the classical vitamin D action (VDRdependent genomic action of 1,25(OH)₂D₃) has a high potency of exhibiting calcemic effect, which often causes side effects, such as hypercalcemia.

To reveal the molecular mechanisms underlying the action of vitamin D, it is important to identify the active form of vitamin D₃. A wide variety of vitamin D metabolites are present in the body to tightly regulate the concentration of $1,25(OH)_2D_3$ in the plasma. For example, $25(OH)D_3$ and $1,25(OH)_2D_3$ are also sequentially converted to biologically inactive metabolites by CYP24A1, whose conversion starts with C-23 and C-24 hydroxylation, respectively (Fig. 1). In the current review, we introduce new molecular mechanisms of action of vitamin D, which is revealed through a detailed analysis of vitamin D metabolism using cultured cells and genetically modified (GM) rickets model animals.

1. Direct biological action of 25(OH)D₃ via VDR in PZ-HPV-7 cells.

Recent reports have demonstrated that 25(OH)D₃ can regulate gene expression by directly binding to the VDR ¹²⁻¹⁵. We have demonstrated that 25(OH)D₃ is a potential VDR ligand in immortalized human prostate PZ-HPV-7 cells ⁷. Whereas the affinity of 25(OH)D₃ toward VDR is less than 100-fold lower than that of $1,25(OH)_2D_3$ ¹⁶, the plasma concentration of 25(OH)D₃ is several hundred-fold higher than that of $1,25(OH)_2D_3$ in the DBP-bound form. Based on its K_d value for the VDR and the plasma concentration of 25(OH)D₃, these biological and biochemical findings suggested that 25(OH)D₃ could be a physiologically important agonist of VDR. We revealed that transcellular localization of VDR was induced by 100 nM 25(OH)D₃ or 1 nM 1,25(OH)₂D₃ in PZ-HPV-7 cells, both of which are physiological conditions of vitamin D^{7} . We further examined the effect of vitamin D on cell growth because previous studies have shown that $1,25(OH)_2D_3$ has cell growth-inhibitory activity mediated by VDR

¹⁷. In agreement with VDR trans-localization, 1 and 10 nM $1,25(OH)_2D_3$ significantly inhibited cell growth. Similarly, 10 and 100 nM $25(OH)D_3$ exerted inhibitory effects on cell growth. We measured the amount of $1,25(OH)_2D_3$ synthesized from 100nM $25(OH)D_3$ in the PZ-HPV-7 cells, and confirmed that cell-derived $1,25(OH)_2D_3$ did not trigger the trans-localization of VDR. We further confirmed that the inhibition of cell growth was not inhibited by CYP27B1 knockdown by siRNA, strongly suggesting that $25(OH)D_3$ acts as a potential VDR ligand in PZ-HPV-7 cells⁷.

2. Cyp27b1-independent 1,25(OH)₂D₃ production in *Cyp27b1*-KO mice.

To confirm the direct action of 25(OH)D₃ in vivo. we examined its effect on osteogenesis in Cyp27b1 knockout (KO) mice ¹⁸. Cyp27b1-KO mice have no detectable $1,25(OH)_2D_3$ in the plasma and exhibit all the hallmarks of vitamin D-dependent rickets (VDDR) type I, such as lower bone mineral hypocalcemia. density and 25(OH)D₃ at 150 $\mu g \cdot kg^{-1} \cdot day^{-1}$ administration normalized bone and calcium phenotypes in Cyp27b1-KO mice. Unexpectedly, a normal level of 1,25(OH)₂D₃ was detected in Cyp27b1-KO mice administered 25(OH)D₃¹⁸. We previously reported that CYP27A1 has a weak 1α -hydroxylation activity toward 25(OH)D₃, which is a liver mitochondrial enzyme involved in 25-hydroxylation of vitamin D, and has 40% homology in amino acid sequence compared to CYP27B1¹⁹. Based on the activity of 1α -hydroxylation towards $25(OH)D_3$ in liver mitochondrial fractions prepared from Cyp27b1-KO mice, we assumed that Cyp27a1 compensatory

converted 25(OH)D₃ to $1,25(OH)_2D_3^{-18}$. Thus, high doses $25(OH)D_3$ of promote compensatory production of $1,25(OH)_2D_3$ by liver Cyp27a1 in Cyp27b1-KO mice, suggesting that 25(OH)D₃ treatment would be useful for patients with CYP27B1disfunction, including VDDR type I rickets and chronic kidney disease. However, we failed to demonstrate the direct action of 25(OH)D₃ in Cyp27b1-KO mice.

3. Elucidation of direct action of 25(OH)D₃ via Vdr in Vdr (R270L) mutant rats.

We hypothesized that the multiple deletion of CYP27B1 and CYP27A1 also fails to demonstrate the direct action of 25(OH)D₃ via VDR because CYPs form a gene superfamily and their substrate specificity often overlaps with others. Thus, we focused on mutated VDR derived from VDDR type II patients. Furthermore, we selected a rat model to generate GM animals because pharmacokinetic analysis of vitamin D metabolites using liquid chromatography/mass spectrometry (LC/MS) often requires a large volume of blood samples. To elucidate the new biological actions of vitamin D, including the direct action of 25(OH)D₃ via VDR, we used Cyp27b1-KO, Vdr(R270L)-knocked in, and Vdr-KO rats generated by the clustered interspaced regularly palindromic repeat/caspase 9 (CRISPR/Cas9) genome editing system.

Rat Vdr mutation, R270L, which is an ortholog of human VDR mutation, R274L, isolated from the VDDR type II rickets patient ²⁰⁻²², exerts significantly reduced affinity towards 1,25(OH)₂D₃ as Arg270 and Arg274

in rats and human VDR play pivotal roles in hydrogen bond formation with 1α hydroxy group of 1,25(OH)₂D₃. The affinity of these mutant VDRs toward 1,25(OH)₂D₃ is approximately 1000-fold lower than that of the wild-type ²², which has an equivalent affinity toward 25(OH)D₃. The patients with VDR (R274L) were hardly responsible for 1,25(OH)₂D₃. Hence, there is no ligand with high affinity for the mutated Vdr in *Vdr*(R270L) rats, which enables us to evaluate the direct action of exogenous 25(OH)D₃ via

Vdr(R270L) because the conversion of $25(OH)D_3$ to $1,25(OH)_2D_3$ is almost negligible for VDR-mediated action in *Vdr*(R270L) rats. In fact, bone disorders associated with hypocalcemia and subsequent hyperparathyroidism were observed in *Vdr*(R270L) rats, while the plasma levels of $1,25(OH)_2D_3$ were significantly elevated after weaning, indicating that $1,25(OH)_2D_3$ could not act as a high-affinity ligand of Vdr(R270L)²³.

Table1. vitamin D signals including canonical (VDR-1,25(OH)₂D₃) and non-canonical actions ²³.

- GM strain	Mode of actions				
	(1) VDR- 1,25(OH) ₂ D ₃	(2) non VDR- 1,25(OH) ₂ D ₃	(3) VDR- 25(OH)D3	(4) non VDR∙ 25(OH)D₃	(5) VDR- no ligand
WT	Yes	Yes	Yes	Yes	Yes
Vdr (R270L)	No	Yes	Yes	Yes	Yes
<i>Cyp27b1</i> ·KO	No	No	Yes	Yes	Yes
Vdr-KO	No	Yes	No	Yes	No

We compared the phenotypes of GM rats to determine the biological effects of different vitamin D actions ²³. As these GM strains lack different vitamin D actions, comparative analysis might reveal the biological effect of each vitamin D action (Fig. 1 and Table1). For example, as Vdr(R270L) rats are deficient in Vdr-dependent action of $1,25(OH)_2D_3$, phenotypes *Vdr*(R270L) abnormal of compared to wild type would be the effect of Vdr-dependent action of 1,25(OH)₂D₃. Similarly, the effect of Vdr-independent action of $1.25(OH)_2D_3$ was revealed by comparison with Cyp27b1-KO and Vdr-KO rats. All GM rats showed rickets symptoms, including bone disorders associated with hypocalcemia. Among these GM rats, Cyp27b1-KO rats showed severe rickets

symptoms, including notably low levels of calcium in the blood and the most severe growth retardation, suggesting that Vdrindependent 1,25(OH)₂D₃ action is also involved in these phenotypes in addition to Vdr-dependent action of 1,25(OH)₂D₃. In contrast, Vdr-KO rats showed abnormal skin formation and alopecia, whereas skin phenotypes Vdr(R270L) rats were almost normal, suggesting that the Vdr-dependent effect of 25(OH)D₃ or the ligand-independent effect of Vdr might be involved in these skin phenotypes ²³. Thus, these GM rats are useful for revealing the molecular mechanisms of vitamin D, including novel and conventional signaling, which lead to novel molecular targets for vitamin D-related diseases (Table 1).

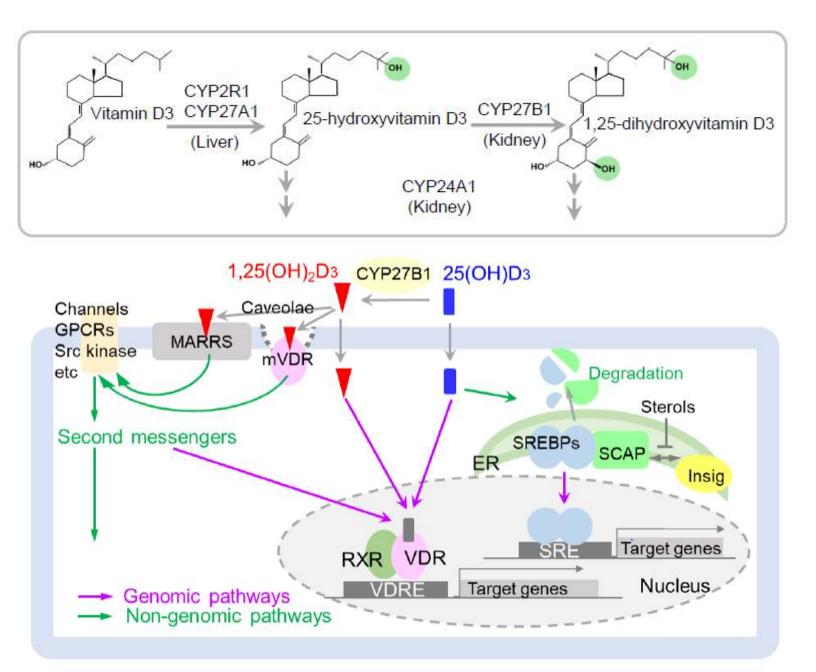


Fig.1. Metabolism and putative molecular mechanisms of vitamin D₃

Vitamin D_3 is converted into 25(OH) D_3 by CYP2R1 and CYP27A1 in the liver, and then further converted into 1,25(OH)₂ D_3 by CYP27B1 in the kidney. The resultant 1,25(OH)₂ D_3 is inactivated by CYP24A1 in the kidney

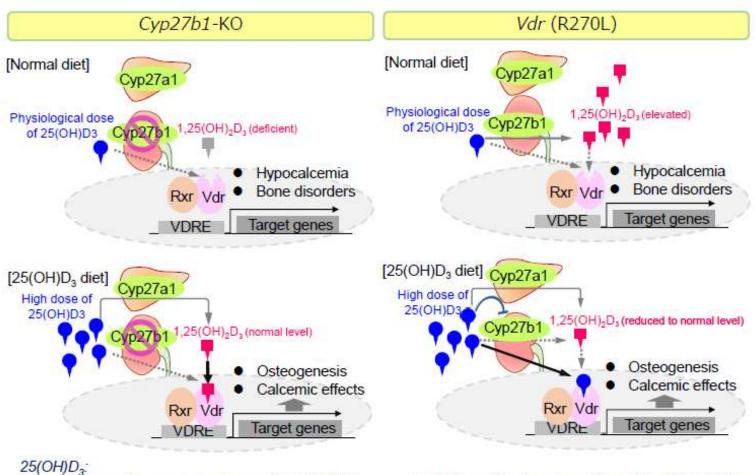
Purple and green arrows indicate genomic and non-genomic pathway, respectively.

GPCRs, G protein-coupled receptor; MARRS, membrane associated, rapid response steroid-binding receptor; VDR, vitamin D receptor; mVDR, membrane-bound vitamin D receptor; RXR, retinoid X receptor; VDRE, vitamin D response element; ER, endoplasmic reticulum; SREBPs, sterol regulatory element–binding proteins; SCAP, SREBP cleavage activating protein; SRE, sterol regulatory element.

Finally, we treated Cyp27b1-KO (VDDR type I model) and Vdr(R270L) (VDDR type II model) rats with $25(OH)D_3$ at $200 \ \mu g \cdot kg^{-1}$. day⁻¹ after weaning to evaluate the biological of 25(OH)D₃ ²³. 25(OH)D₃ activities treatment fully normalized their rickets phenotypes, including hypocalcemia and bone disorders. As expected, plasma 1,25(OH)₂D₃ deficiency in Сур27b1-КО rats was normalized by 25(OH)D₃ in accordance with a study using Cyp27b1-KO mice²³. In $25(OH)D_3$ decreased contrast. plasma $1.25(OH)_2D_3$ to the normal level in Vdr(R270L) rats, whose plasma 1,25(OH)₂D₃ levels were significantly elevated in the absence of 25(OH)D₃ treatment. These results suggest the following: (1) In Cyp27b1 deficient conditions, high-dose treatment of $25(OH)D_3$ is useful as a source for extrarenal

1,25(OH)₂D₃ synthesis, and hepatic Cyp27a1 might be involved in 1 α -hydroxylation of 25(OH)D₃ under these conditions. (2) 25(OH)D₃ itself acts as a ligand for Vdr(R270L) at a high dose of 25(OH)D₃²³ (Fig. 2). 1,25(OH)₂D₃ is clinically used for treatment of rickets, even in VDDR type II patients who are hardly responsible for 1,25(OH)₂D₃. Our findings demonstrated the usefulness of 25(OH)D₃ as a wide-safety-region compound for CYP27B1 deficient or VDR(R274L).

In conclusion, we demonstrated novel vitamin D actions involved in calcium homeostasis and osteogenesis by a comparative approach using GM rats deficient in different vitamin D signals. Our novel *in vivo* system is useful for elucidating the molecular mechanism of vitamin D action.



Intracellular pool to generate extrarenal 1,25(OH)₂D₃

25(OH)D₃: active form (weak ligand for Vdr(R270L)

Fig.2. Biological roles of 25(OH)D₃ in Cyp27b1-KO and Vdr(R270L) rats [23]

In the case of Cyp27b1-KO rats fed a normal diet, plasma 1,25(OH)₂D₃ level was dramatically reduced to result in hypocalcemia and bone disorders.

In the case of Cyp27b1-KO rats fed a 25(OH)D₃ containing diet, plasma 1,25(OH)₂D₃ level was increased to a normal level probably by Cyp27a1 to result in normal plasma

References

1. Plum LA, DeLuca HF. Vitamin D, disease and therapeutic opportunities. Nat Rev Drug Discov. 2010;9(12):941-955.

2. Sakaki T, Kagawa N, Yamamoto K, et al. Metabolism of vitamin D3 by cytochromes P450. Front Biosci. 2005;10:119-134.

3. Haussler MR, Whitfield GK, Kaneko I, et al. Molecular mechanisms of vitamin D action. Calcif Tissue Int. 2013;92(2):77-98.

4. Norman AW, Bishop JE, Collins ED, et al. Differing shapes of 1 alpha,25dihydroxyvitamin D3 function as ligands for the D-binding protein, nuclear receptor and membrane receptor: a status report. J Steroid Biochem Mol Biol. 1996;56 (1-6 Spec No): 13-22.

5. Mizwicki MT, Norman AW. The vitamin D sterol-vitamin D receptor ensemble model offers unique insights into both genomic and rapid-response signaling. Sci Signal. 2009;2(75): re4; 10.1126/scisignal.275re4.

6. Hii CS, Ferrante A. The Non-Genomic Actions of Vitamin D. Nutrients. 2016;8(3): 135; 10.3390/nu8030135.

7. Munetsuna E, Kawanami R, Nishikawa M, et al. Anti-proliferative activity of 25hydroxyvitamin D3 in human prostate cells. Mol Cell Endocrinol. 2014;382(2): 960-970.

8. Masuda S, Jones G. Promise of vitamin D analogues in the treatment of hyperproliferative conditions. Mol Cancer Ther. 2006;5(4),:797-808.

9. Asano L, Watanabe M, Ryoden Y, et al. Vitamin D Metabolite, 25-Hydroxyvitamin D, Regulates Lipid Metabolism by Inducing Degradation of SREBP/SCAP. Cell Chem Biol. 2017;24(2):207-217.

10. Malloy PJ, Feldman D. The role of vitamin D receptor mutations in the

development of alopecia. Mol Cel Endocrinol. 2011;347(1-2):90-96.

11. Skorija K, Cox M, Sisk JM, et al. Ligandindependent actions of the vitamin D receptor maintain hair follicle homeostasis. Mol Endocrinol. 2005;19(4):855-862.

12. Lou YR, Laaksi I, Syvälä H, et al. 25hydroxyvitamin D3 is an active hormone in human primary prostatic stromal cells. FASEB J. 2004;18(2):332-334.

13. Peng X, Hawthorne M, Vaishnav A, et al. 25-Hydroxyvitamin D3 is a natural chemopreventive agent against carcinogen induced precancerous lesions in mouse mammary gland organ culture. Breast Cancer Res Treat. 2009;113(1):31-41.

14. Lou YR, Molnár F, Peräkylä M, et al. 25-Hydroxyvitamin D(3) is an agonistic vitamin D receptor ligand. J Steroid Biochem Mol Biol. 2010;118(3):162-170.

15. Verone-Boyle AR, Shoemaker S, Attwood K, et al. Diet-derived 25hydroxyvitamin D3 activates vitamin D receptor target gene expression and suppresses EGFR mutant non-small cell lung cancer growth in vitro and in vivo. Oncotarget. 2016;7(1):995-1013.

16. Bouillon R, Okamura WH, Norman AW. Structure-function relationships in the vitamin D endocrine system. Endocr Rev. 1995;16(2):200-257.

17. Dusso AS, Brown AJ, Slatopolsky E.Vitamin D. Am J Physiol Renal Physiol.2005;289(1):F8-28.

18. Nishikawa M, Yasuda K, Takamatsu M, et al. Generation of 1,25-dihydroxyvitamin D3 in Cyp27b1 knockout mice by treatment with 25-hydroxyvitamin D3 rescued their rachitic phenotypes. J Steroid Biochem Mol Biol. 2019;185:71-79.

19. Sawada N, Sakaki T, Ohta M, et al. Metabolism of vitamin D(3) by human CYP27A1. Biochem Biophys Res Commun. 2000;273(3):977-984.

20. Kristjansson K, Rut AR, Hewison M, et al. Two mutations in the hormone binding domain of the vitamin D receptor cause tissue resistance to 1,25 dihydroxyvitamin D3. J Clin Invest. 1993;92(1):12-16.

21. Malloy PJ, Feldman D. Genetic disorders and defects in vitamin D action. Endocr Metab Clin. 2010;39(2):333–346. 22. Nakabayashi M, Tsukahara Y, Iwasaki-Miyamoto Y, et al. Crystal structures of hereditary vitamin D-resistant ricketsassociated vitamin D receptor mutants R270L and W282R bound to 1,25dihydroxyvitamin D3 and synthetic ligands. J Med Chem. 2013;56(17):6745-6760.

23. Nishikawa M, Yasuda K, Takamatsu M, et al. Generation of novel genetically modified rats to reveal the molecular mechanisms of vitamin D actions. Sci Rep. 2020;10(1):5677.