

Published: February 28, 2023

Citation: Arnsten AFT and Baudry M, 2023. Targeting Calpain-2 For Alzheimer's Disease Treatment, Medical Research Archives, [online] 11(2).
<https://doi.org/10.18103/mra.v11i2.3487>

Copyright: © 2023 European Society of Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

DOI
<https://doi.org/10.18103/mra.v11i2.3487>

ISSN: 2375-1924

RESEARCH ARTICLE

Targeting Calpain-2 for Alzheimer's Disease Treatment

Amy F.T. Arnsten¹ and Michel Baudry²

¹Department of Neuroscience, Yale University School of Medicine, New Haven, CT

²Department of Dental Medicine, Western University of Health Sciences, Pomona, CA

*mbaudry@westernu.edu

ABSTRACT

There is an urgent need for treatments for sporadic Alzheimer's Disease (sAD). Although antibodies removing β -amyloid have recently been shown to slow disease progression, the degenerative course continues. Thus, there is a need for strategies that intervene early in the degenerative process, before irreversible damage is done to neurons (e.g., by autophagic degeneration). This review will summarize the evidence indicating that targeting calpain-2 with a selective inhibitor might represent a novel strategy for the treatment of sAD. Calpains are neutral proteases that are activated by intracellular calcium. The two main isoforms are calpain-1, which is activated by low, micromolar levels of calcium and generally has beneficial effects for cellular health, and calpain-2, which is activated by high, almost millimolar levels of calcium and mediates many of calcium's toxic actions. Calcium signaling becomes dysregulated with advancing age due to loss of regulatory proteins such as calbindin, and is pronounced in sAD brain tissue, including signs of calcium leakage from the smooth endoplasmic reticulum (SER) through phosphorylated ryanodine receptors (pRyR2). Both calpain-1 and calpain-2 are elevated in AD brains and herald the rise in tau pathology, but only calpain-2 is localized with neurofibrillary tangles (NFTs) and pretangles. Calpain drives a spectrum of AD-related pathologies, and in particular, calpain drives tau hyperphosphorylation by cleaving, and thus disinhibiting, kinases central to tau hyperphosphorylation, i.e., GSK3 β and cdk5, as well as increasing A β formation and autophagic degeneration. Thus calpain-2 inhibitors may reduce a spectrum of sAD pathology, protecting neurons at very early stages of disease.

1. Introduction

Current treatment strategies for Alzheimer's disease (AD) have focused on targeting amyloid or tau pathology, the traditional hallmarks of AD neuropathology. Although this approach has begun to have some success in slowing progression, it has not cured or halted the disease. Ideally, we need to understand what causes the neurodegeneration and accompanying tau and amyloid pathologies, in order to intervene very early to prevent the initiation of the pathological processes, especially since even early stage tau and amyloid pathology can have toxic effects in neurons. However, very early intervention would require a very benign treatment that potentially could be taken for decades by healthy individuals, without interfering with normal cellular functioning. The current review describes the importance of calcium dysregulation as an early etiological factor in the development of AD, and the potential use of selective calpain-2 inhibitors to safely reduce pathological mechanisms prior to the onset of irreversible neuronal damage.

Calpains are soluble, neutral, calcium-dependent proteases, i.e., they cleave other proteins, and they are activated by intracellular calcium in the cytosol. While there are 15 members of the calpain family, two ubiquitous isoforms of calpains are calpain-1, which is activated by micromolar concentrations of calcium that occur under normal physiological conditions, and calpain-2, which is activated by almost millimolar concentrations of calcium. In addition, recent studies have shown that calpain-1 activation is neuroprotective while calpain-2 mediates many of calcium's toxic actions¹. As reviewed below, dysregulation of calcium with advancing age and/or inflammation leads to noxious levels of cytosolic calcium and activation of calpain-2, which in turn drives tau and amyloid pathology, synapse loss, and autophagic neurodegeneration. We propose that selective inhibition of brain calpain-2 may reduce early pathological events while leaving beneficial calcium actions intact, thus protecting neurons from future insults.

2. Alzheimer's disease pathology, and the amyloid vs. tau hypotheses

AD is a progressive neurodegenerative disorder characterized by memory loss and cognitive deficits, which ultimately lead to severe dementia². AD is the most prominent form of dementia in aged people, and the number of Americans suffering from the disease is projected

to reach about 15 million over the next few decades^{3,4}. AD brain pathology is characterized by the presence of senile plaques consisting of aggregated extracellular β -amyloid peptide, and intracellular neurofibrillary tangles, which are composed of hyperphosphorylated tau proteins². In addition, AD is associated with synapse loss and neuronal death in hippocampus, entorhinal cortex, basal forebrain, and neocortical association cortices⁵. Since AD affects the health and lives of so many people, there is a clear urgency to understand the molecular/cellular processes underlying the disease and to develop treatments that can slow down its progression and ultimately prevent or reverse it.

Although the brain pathology associated with AD is now widely agreed upon, the underlying mechanisms remain unclear and debated. For years, the similarities between the pathology in familial, early onset, autosomal dominant forms of AD, and the sporadic forms of AD, have suggested the existence of a common etiology. Familial forms of AD appear to be due to mutations in genes encoding the amyloid precursor protein⁶, and presenilin-1 and presenilin-2⁷, which all result in increased levels of β -amyloid peptides in brain⁸. However, familial forms of AD account for less than 5% of all AD cases⁹; thus, it is clear that the vast majority of AD patients develops AD sporadically (sAD).

The genetics of familial forms of AD have led to the amyloid hypothesis of AD, which postulates that A β accumulation initiates the pathological process and drives tau pathology, which results in synaptic and dendritic dysfunction, and ultimately neuronal death. This hypothesis has led to the use of numerous mouse models with mutations in various amyloid-related genes to reproduce the human pathology with some degree of success⁸. This has also inspired the development of numerous, potential therapies targeting amyloid deposition or clearing. However, most of these have failed in clinical trials¹⁰⁻¹². Only recently, lecanemab, an antibody against A β , which removes amyloid from AD brains, has shown a small but highly significant slowing in cognitive decline¹³, supporting the amyloid hypothesis. However, other compounds, successful in removing brain amyloid pathology, have had limited or no success¹⁴⁻¹⁶, and lecanemab does not stop progression of the disease and does not appear to help patients with aggressive forms of disease, e.g., those who are apoE4 homozygotes¹³.

Although much of the field has focused on the amyloid hypothesis, extensive postmortem evaluations of sporadic AD brains by Braak and colleagues have repeatedly shown that tau pathology is initiated 10 years *before* the appearance of amyloid plaques¹⁷. Moreover, the extent of tau pathology, and not amyloid pathology, correlates positively with the degree of loss of grey matter and of cognitive impairment¹⁸. The time-course and regional distribution of tau pathology are also different than those of the A β accumulation and are consistent with the progression of cognitive symptoms. Tau pathology is particularly pronounced in glutamatergic projection neurons with extensive cortical-cortical connections and not in GABAergic interneurons. In cortex, tau pathology is initiated in layer II of the entorhinal and perirhinal cortices, and then proceeds to interconnected limbic and association cortices, as well as hippocampus. This progression likely involves the trafficking of phosphorylated tau between connected neurons¹⁹, which are in a vulnerable state, e.g., with calcium dysregulation, as described below.

Recent data suggest that tau pathology may initiate or hasten amyloid pathology. Immunoelectron microscopy has shown that tau hyperphosphorylation and aggregation on microtubules could drive A β generation in neurons due to an "endosomal traffic jam", as it traps the amyloid protein precursor (APP) in endosomes containing β -secretase, increasing its cleavage to A β ²⁰. As A β in turn drives tau phosphorylation, this would establish a vicious cycle of neuropathology²¹.

3. Calcium dysregulation as an early pathological event in AD

AD specifically afflicts the limbic and association cortices, and aging is the largest risk factor for the disease, with inflammatory conditions (e.g., insulin insensitivity, traumatic brain injury) contributing additional risk. Why are glutamatergic neurons in the limbic and association especially vulnerable with advanced age and inflammation? Studies from a number of disciplines indicate that these neurons express the molecular machinery to magnify calcium signaling needed to sustaining neural representations in memory stores^{22,23}. However, dysregulation of calcium with advancing age and/or inflammation renders these neurons particularly vulnerable to AD pathology. The calcium hypothesis of AD has been discussed for decades²⁴⁻²⁷, with early studies

of AD brains by Nixon and Mattson showing that dysregulated calcium is likely a major initiating factor in AD pathology^{28,29}, and recent research from primate models has reinforced their groundbreaking work.

The pyramidal cells in layer III of the dlPFC have been an important focus of research, as these neurons are critical for higher cognitive functions^{30,31}, and are selectively vulnerable to tau pathology and neurodegeneration in AD^{32,33}, thus providing important clues about AD etiology. Studies of these neurons in rhesus monkeys have shown that they express the molecular machinery to magnify calcium signaling in dendritic spines at glutamate synapses (Fig. 1), where cAMP-PKA signaling increases calcium release from the smooth endoplasmic reticulum (SER), which in turn leads to more cAMP formation, thus producing a feedforward signaling³¹. Rhesus monkeys are all apoE4 homozygotes³⁴, a major risk factor for AD, which make them a particularly interesting animal model to study the etiology of AD. In addition to internal calcium release, cAMP-PKA signaling also magnifies calcium entry through NMDAR³⁵, and voltage-gated calcium channels³⁶, which can in turn further increase calcium-mediated calcium release from the SER³⁶. Interestingly, apoE4 also increases calcium entry at these sites³⁷⁻⁴⁰, suggesting that this may be one of the important ways apoE4 increases risk of AD. In the healthy, young dlPFC neurons, cAMP magnification of calcium signaling is tightly regulated by phosphodiesterases (PDE4s), which degrade cAMP, and by calbindin, which binds cytosolic calcium⁴¹. However, with advancing age and/or inflammation, PDE4s and calbindin are lost from layer III dlPFC dendrites, leading to excessive cAMP-PKA-calcium signaling. This process is worsened by PKA phosphorylation of ryanodine receptors (pRyR2), which flux calcium out of the SER, leading to calcium leak from the SER into the cytosol⁴². Studies of aging monkeys show that calcium leak through pRyR2 highly correlates with the initiation of early stage tau phosphorylation, e.g., by PKA at serine 214 (Fig. 1)⁴³. Tau phosphorylation at S214 causes tau to change shape and detach from microtubules, and primes it for hyperphosphorylation by other kinases, especially GSK3 β and cdk5. As described below, these events are particularly damaging when cytosolic calcium levels are sufficient to activate calpain-2.

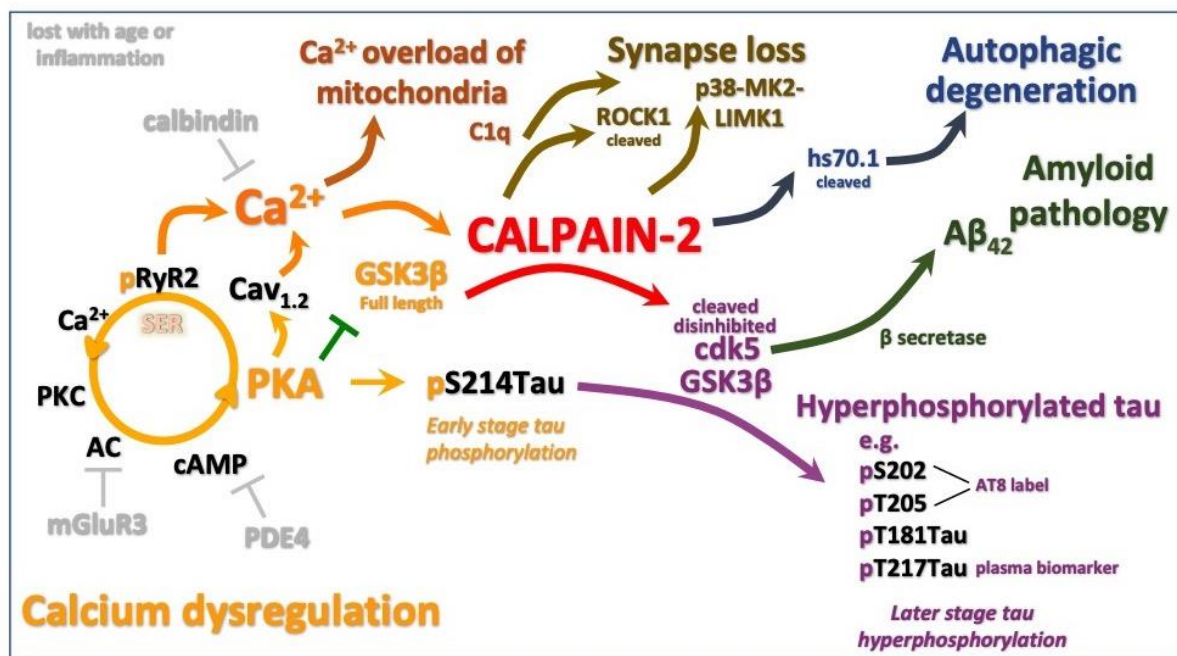


Figure 1. Role of calpain-2 in AD pathology. Dysregulation of feedforward, cAMP-calcium signaling with advancing age and/or inflammation generates high levels of cytosolic calcium, which activates calpain-2. Calpain-2 cleaves and disinhibits GSK3β and cdk5, which hyperphosphorylate tau. Cleaved cdk5 also increases BACE/Aβ production, driving amyloid pathology. Calpain-2 can also drive synapse loss and autophagic degeneration through cleavage of ROCK1 and hsp70.1, respectively, thus worsening multiple pathological events in AD. (Adapted from Arnsten et al, 2021¹⁷).

Importantly, evidence of calcium leak from the SER through pRyR2 has been documented in brain tissues from patients with sporadic AD, confirming its relevance to the human disease^{44,45}. Indeed, some of the earliest evidence of the importance of calcium leak to neural pathology came from Mattson's studies of tissues with PS1 mutations, the most common, and aggressive cause of autosomal dominant, early onset AD. He showed that either a PS1 or PS2 mutation causes dramatic calcium leak from the SER and hypothesized that this would be a significant driver of neuropathology^{28,46}.

Dysregulated calcium signaling has also been linked to heart failure, where calcium leak from the sarcoplasmic reticulum causes calcium overload of mitochondria in cardiac muscle, driving oxidative stress, which in turn causes more calcium toxicity⁴⁷. Evidence of these pathological events can also be seen in brain of the aged rhesus monkey dIPFC and of AD patients, where calcium overload causes an abnormal mitochondrial morphology called Mitochondria-On-A-String (MOAS)^{48,49}. In AD, the activity of the electron transport chain enzyme cytochrome c oxidase is significantly reduced^{50,51}, ATP production is diminished^{52,53}, and there is a decrease in both

mitochondrial mass and mitochondrial DNA content^{54,55}. As mitochondria normally regulate calcium signaling by taking up calcium, mitochondrial dysfunction can further drive calcium dysregulation. When cytosolic calcium levels become abnormally high, they can activate calpain-2, which can cause a large number of pathological events seen in AD brains, as described below, and summarized in Figure 1.

4. Links between calpain and AD

The calcium-dependent proteases, calpains, have long been associated with the pathology of AD^{56,57}. Calpains represent a 15-member family of calcium-dependent proteases, and it has been difficult to determine the roles of specific calpain isoforms in the brain in general and in AD in particular. The major calpain isoforms in the brain are calpain-1 and calpain-2, the so-called classical calpains⁵⁸. Recent findings indicate that calpain-1 and calpain-2 play opposite functions in the brain, with calpain-1 being neuroprotective and calpain-2 neurodegenerative¹. Calpain-2 was also identified as an early component of neurofibrillary tangles in AD using active-site directed antibodies⁵⁹. More recently, hyperactivation of calpain-2 was shown to occur presymptomatically

in a mouse model of AD and to correlate with memory deficits in AD patients⁶⁰.

Calpain has long been proposed to participate in the pathology of Alzheimer's disease. It was first speculated that impaired calpain activity could lead to the development of neurofibrillary degeneration and cytoskeletal alterations⁶¹. A few years later, the idea was advanced that excitotoxicity was involved in AD pathology and that activation of calpain-1 was responsible for this phenomenon⁶². Interestingly, Ralph Nixon changed his initial hypothesis and proposed that calpains could promote β -amyloidogenesis, neurofibrillary pathology and neuronal degeneration^{63,64}. In support of this idea, the Nixon group found that calpain-2 specifically is found in association with neurofibrillary tangles in the brains of patients with AD. Other groups have also found that the rise in calpain activity heralds the onset of tau pathology⁵⁷.

The Nixon lab tested whether reducing calpain activity would be protective in mouse models, and found that overexpression of the endogenous calpain inhibitor, calpastatin, reduced excitotoxic stress-induced neurodegeneration⁶⁵. The role of cdk5/p25 in the formation of neurofibrillary tangles also supported the involvement of calpain, as calpain is responsible for the cleavage of p35 into p25, the cdk5 activator⁶⁶. These findings led to the suggestion that calpain inhibitors could be used as neuroprotective drugs for the treatment of various neurodegenerative disorders, including AD⁶⁷⁻⁶⁹. A recent review summarized a whole body of evidence linking calpain to AD, including the roles of calpain in both A β and tau pathology, as well as in autophagy impairment⁵⁷. It also reviews many studies showing that a variety of calpain inhibitors provided beneficial effects on a range of pathological parameters, including learning and memory, A β accumulation and tau hyperphosphorylation in various animal models of AD. Depletion of the endogenous calpain inhibitor, calpastatin, resulting from the activity of caspases and calpain, has been shown to produce calpain-2 activation and tau and neurofilament hyperphosphorylation⁷⁰. All these findings led Abbvie to conduct a Phase I clinical trial with a non-selective calpain inhibitor, ABT-957, aka Alicapostat, for Alzheimer's disease. The study tested the effects of the calpain inhibitor in both healthy and MCI patients, with twice daily doses and for 12 weeks. However, the study was terminated due to lack of pharmacodynamic effects, assessed by changes in REM sleep, suggesting that the inhibitor did not reach high enough levels in the

brain⁷¹.

In addition to driving the hyperphosphorylation of tau, activated calpain increases BACE and A β levels via cleavage of cdk5⁷², and drives autophagic degeneration through the activation of heatshock protein 70.1 (Hsp70.1)⁷³, as summarized in Figure 1. As neurons die from autophagic degeneration in AD, and not by apoptosis, this latter finding suggests that calpain-2 may simultaneously drive tau pathology and autophagic degeneration within the same neurons. This would be consistent with Yamashima's "calpain-cathepsin hypothesis" for AD. In this model, calpain-mediated cleavage of Hsp70.1 results in the permeabilization of lysosomes and the leakage of cathepsin in the cytoplasm⁷⁴.

In most of these studies, there is little, if any, discussion regarding the respective roles of calpain-1 and calpain-2, the major calpain isoforms ubiquitously expressed in the brain. Both calpain-1 and calpain-2 appear to be elevated in AD brains and herald the rise in tau pathology⁷⁵, but only calpain-2 is localized with neurofibrillary tangles (NFTs) and pretangles⁵⁹. Importantly a recent study reported specific hyperactivation of calpain-2 in in both a mouse model of AD and in cortical tissues from post-mortem AD patient brains⁶⁰. Elevated calpain-2 was observed in presymptomatic 1 month-old APP^{swe}/PS1 Δ E9 mice and persisted up to 10 months of age. In humans, elevated calpain-2 activity was significantly correlated with cognitive impairment. Interestingly, while calpain-2 activity was positively correlated with A β load, there was no correlation between calpain-2 activity and hyperphosphorylated tau⁶⁰. The authors postulated that elevated calpain-1 activity might happen later in the disease progression and could be responsible for tau pathology. Clearly, more work remains to be done to clarify the respective roles of calpain-1 and calpain-2 on the various pathological manifestations of AD.

As mentioned above and by many reports⁷⁶, calpain drives a spectrum of AD-related pathologies, and in particular, tau hyperphosphorylation by cleaving, and thus disinhibiting, kinases central to tau hyperphosphorylation, i.e., GSK3 β and cdk5. GSK3 β and cdk5 hyperphosphorylate tau at key sites, including pS202 and pT205 (labeled by AT8), and pT181Tau and pT217Tau, which are important, emerging fluid biomarkers for AD pathology, including pT217Tau in plasma^{77,78}. Early stage tau phosphorylation at S214 by PKA primes tau for

subsequent hyperphosphorylation by GSK3 β ⁷⁹, and thus creates a reservoir of primed phosphorylated tau (pTau) species (Fig. 1). Calpain-2 cleavage of GSK3 β appears to be a critical step, as it removes the site where PKA normally inhibits GSK3 β ^{80,81}, and thus allows GSK3 β to phosphorylate tau without any regulation. Calpain-2 also selectively cleave and deactivate the tyrosine phosphatase, PTPN13, which can indirectly promote tau phosphorylation, and produces a stable breakdown product, P13BP, that can be measured in CSF and plasma^{82,83}.

Another important aspect of the role of calpain in AD pathology is the putative role of calpain in generating tau fragments that might seed the formation of neurofibrillary tangles^{76,84,85}. The issue of whether calpain-mediated tau cleavage results in neurotoxicity remains debated. Results in *Drosophila* convincingly showed that calpain-mediated generation of a 17 kD fragment of tau was responsible for neurotoxicity⁸⁶. On the other hand, Garg et al.⁸⁷ concluded that the 17 kD tau fragment was not a mediator of A β toxicity. More recently, aggregates of the 17 kD tau fragment (also referred to as tau₄₅₋₂₃₀) were found in AD brains and could be internalized and induce neurodegeneration in hippocampal neurons⁸⁸. However, there is a general agreement that calpain cleavage and activation of GSK3 β and cdk5 could drive AD pathology, and thus this would strongly support the idea that a selective calpain-2 inhibitor could have beneficial effects in AD and may preserve the beneficial effects of calpain-1 needed for housekeeping functions and neuroplasticity.

5. Calpain-2 and neurodegeneration

Calpain activation has been implicated in neurodegeneration since the early 80s, and most reviews written on calpains all mentioned the role of calpains in neurodegenerative processes⁸⁹⁻⁹². However, very few studies have explored the specific contributions of calpain-1 and calpain-2 in neurodegeneration. Since 2012, our laboratory has focused on evaluating the contributions of calpain-1 and calpain-2 in both synaptic plasticity and neurodegeneration. We showed that calpain-2, but not calpain-1, activation was responsible for NMDA-induced excitotoxicity through the activation of the striatal-enriched tyrosine phosphatase (STEP) in primary neuronal cultures⁹³. A similar study indicated that knock-down of calpain-2, but not calpain-1, increased neuronal survival following NMDA treatment of cultured hippocampal neurons⁹⁴. Because calpains

cleave a large number of proteins involved in neurodegeneration⁹⁵, it has been difficult to determine under various experimental conditions which of them are responsible cell death. Importantly, we found that calpain-2 is associated with a multi-protein complex including extrasynaptic NMDARs, which have been shown to be involved in neuronal death⁹⁶. In particular, NR2B subunits are enriched in extrasynaptic NMDARs⁹⁷, and their activation leads to calpain-2-mediated cleavage of STEP and to neuronal death⁹⁸. NR2B also interacts with RasGRF1, which leads to ERK activation⁹⁹, calpain-2 phosphorylation and prolonged calpain-2 activation¹⁰⁰. Numerous studies have shown that calpain cleaves (STEP), generating inactive fragments, resulting in the activation of p38 and downstream cell death signaling pathways^{98,101}. While this pathway is activated following acute brain injury, it might not be the one involved in chronic neurodegenerative diseases, as neurons appear to die by autophagy and not apoptosis.

As discussed above, over the last 20 years, Yamashima has developed the calpain-cathepsin hypothesis to account for several features of neuronal death in Alzheimer's disease^{74,102-104}. A main feature of this hypothesis is the truncation of carbonylated Hsp70.1 by calpain, leading to the destabilization of lysosomal membranes and the release of cathepsins in neuronal cytoplasm. Incorporated in this hypothesis is the concept that oxidative stress, which has often been associated with AD¹⁰⁵, could stimulate the formation of carbonylated Hsp70.1, and calpain activation through disruption of mitochondrial function. Reactive oxygen species (ROS), which accumulate as a result of mitochondrial dysfunction, have been shown to activate calpain and in particular, calpain-2 in several types of cells and under a variety of experimental conditions¹⁰⁶⁻¹⁰⁸. There is therefore a direct link between calpain-2 activation, lysosomal dysfunction and neuronal death.

6. Conclusions

Accumulating evidence indicates that overactivation of calpain-2 plays a critical role in the initiation and progression of AD pathology. Calcium dysregulation within glutamatergic neurons in the limbic and association cortices with advancing age may activate calpain-2 to spur AD pathology, including A β generation, tau hyperphosphorylation and autophagic degeneration. As calpain activation heralds tau pathology, and calpain-2 is specifically associated

with neurofibrillary tangles in AD brain, calpain-2 may be an important therapeutic target. Early treatment with a selective calpain-2 inhibitor may therefore provide a real preventive treatment for AD, leaving the beneficial effects of calpain-1 intact. This strategy may allow a treatment with a very low side effect profile, necessary for early, long-term treatment.

A challenge of preventive trials is how to measure early indices of therapeutic efficacy, e.g., at an age prior to the formation of fibrillated A β and pTau species measured by PET imaging. Plasma and CSF biomarkers may be a fruitful direction forward, given the recent successes in this arena. The effects of calpain-2 inhibition on tau and amyloid pathology could be assessed with plasma and CSF measures of pT217Tau, and CSF assays of A β ratios, which are altered at very early stages.

The efficacy of a calpain-2 inhibitor on calpain-2 activity could also be assessed using a new blood biomarker, consisting of the calpain-2-mediated fragment of the tyrosine phosphatase, PTPN13, and referred to as P13BP, which has been found to be elevated in postmortem samples from AD patients⁸², and thus may be used to track drug efficacy in addition to plasma and CSF markers of pT217Tau and A β ratios. These methods may allow the development of treatments that could actually prevent AD by reducing pathology at its earliest stages. Furthermore, new drug delivery devices are targeting brain more specifically by using intranasal delivery^{109,110}. Such an approach would mitigate the potential harmful peripheral effects of a chronic treatment with a selective calpain-2 inhibitor.

References

1. Baudry M, Bi X. Calpain-1 and Calpain-2: The Yin and Yang of Synaptic Plasticity and Neurodegeneration. *Trends Neurosci.* Feb 10 2016;doi:10.1016/j.tins.2016.01.007
2. Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev.* Apr 2001;81(2):741-66. doi:10.1152/physrev.2001.81.2.741
3. Katzman R, Saitoh T. Advances in Alzheimer's disease. *FASEB J.* Mar 1 1991;5(3):278-86.
4. Salmon DP, Thomas RG, Pay MM, et al. Alzheimer's disease can be accurately diagnosed in very mildly impaired individuals. *Neurology.* Oct 8 2002;59(7):1022-8. doi:10.1212/wnl.59.7.1022
5. DeKosky ST, Scheff SW, Styren SD. Structural correlates of cognition in dementia: quantification and assessment of synapse change. *Neurodegeneration.* Dec 1996;5(4):417-21. doi:10.1006/neur.1996.0056
6. Goate A, Chartier-Harlin MC, Mullan M, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature.* Feb 21 1991;349(6311):704-6. doi:10.1038/349704a0
7. Cruts M, Van Broeckhoven C. Presenilin mutations in Alzheimer's disease. *Hum Mutat.* 1998;11(3):183-90. doi:10.1002/(SICI)1098-1004(1998)11:3<183::AID-HUMU1>3.0.CO;2-J
8. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science.* Jul 19 2002;297(5580):353-6. doi:10.1126/science.1072994
9. Onyango IG, Khan SM. Oxidative stress, mitochondrial dysfunction, and stress signaling in Alzheimer's disease. *Curr Alzheimer Res.* Sep 2006;3(4):339-49. doi:10.2174/156720506778249489
10. Anderson R, Hadjichrysanthou C, Evans S, Wong M. Why do so many clinical trials of therapies for Alzheimer's disease fail? *Lancet.* 2017;390:2327-2329.
11. Cummings J. Lessons learned from Alzheimer Disease: Clinical trials with negative outcomes. *J Clin Transl Sci.* 2018;11:147-152.
12. Yiannopoulou KG, Anastasiou AI, Zachariou V, Pelidou SH. Reasons for Failed Trials of Disease-Modifying Treatments for Alzheimer Disease and Their Contribution in Recent Research. *Biomedicines.* Dec 9 2019;7(4)doi:10.3390/biomedicines7040097
13. van Dyck CH, Swanson CJ, Aisen P, et al. Lecanemab in Early Alzheimer's Disease. *N Engl J Med.* Nov 29 2022;doi:10.1056/NEJMoa2212948
14. Elmaleh DR, Farlow MR, Conti PS, Tompkins RG, Kundakovic L, Tanzi RE. Developing Effective Alzheimer's Disease Therapies: Clinical Experience and Future Directions. *J Alzheimers Dis.* 2019;71(3):715-732. doi:10.3233/JAD-190507
15. Jeremic D, Jimenez-Diaz L, Navarro-Lopez JD. Past, present and future of therapeutic strategies against amyloid-beta peptides in Alzheimer's disease: a systematic review. *Ageing Res Rev.* Dec 2021;72:101496. doi:10.1016/j.arr.2021.101496
16. Reiss AB, Montufar N, DeLeon J, et al. Alzheimer Disease Clinical Trials Targeting Amyloid: Lessons Learned From Success in Mice and Failure in Humans. *Neurologist.* Mar 4 2021;26(2):52-61. doi:10.1097/NRL.0000000000000320
17. Arnsten AFT, Datta D, Del Tredici K, Braak H. Hypothesis: Tau pathology is an initiating factor in sporadic Alzheimer's disease. *Alzheimers Dement.* Jan 2021;17(1):115-124. doi:10.1002/alz.12192
18. Giannakopoulos P, Hermann F, Bussiere T, et al. Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology.* 2003;60:1495-1500.
19. Peng C, Trojanowski JQ, Lee VM. Protein transmission in neurodegenerative disease. *Nat Rev Neurol.* Apr 2020;16(4):199-212. doi:10.1038/s41582-020-0333-7
20. Paspalas CD, Carlyle BC, Leslie S, et al. The aged rhesus macaque manifests Braak stage III/IV Alzheimer's-like pathology. *Alzheimers Dement.* May 2018;14(5):680-691. doi:10.1016/j.jalz.2017.11.005
21. Busche MA, Hyman BT. Synergy between amyloid-beta and tau in Alzheimer's disease. *Nat Neurosci.* Oct 2020;23(10):1183-1193. doi:10.1038/s41593-020-0687-6
22. Dash PK, Moore AN, Kobori N, Runyan JD. Molecular activity underlying working memory. *Learn Mem.* Aug 2007;14(8):554-63. doi:10.1101/lm.558707
23. Arnsten AF, Wang MJ, Paspalas CD. Neuromodulation of thought: flexibilities and vulnerabilities in prefrontal cortical network synapses. *Neuron.* Oct 4 2012;76(1):223-39. doi:10.1016/j.neuron.2012.08.038

24. Khatchaturian Z. Calcium, membranes, aging, and Alzheimer's disease. Introduction and overview. *Ann N Y Acad Sci.* 1989;568:1-4.
25. Khatchaturian Z. Calcium hypothesis of Alzheimer's disease and brain aging. *Ann N Y Acad Sci.* 1994;747:1-11.
26. Berridge MJ. Calcium signalling and Alzheimer's disease. *Neurochem Res.* Jul 2011;36(7):1149-56. doi:10.1007/s11064-010-0371-4
27. Workgroup AsACH. Calcium hypothesis of Alzheimer's disease and brain aging: A framework for integrating new evidence into a comprehensive theory of pathogenesis. *Alzheimers Dement.* 2017;13:178-182.
28. Mattson MP, Chan SL. Dysregulation of cellular calcium homeostasis in Alzheimer's disease: bad genes and bad habits. *J Mol Neurosci.* Oct 2001;17(2):205-24. doi:10.1385/JMN:17:2:205
29. McBrayer M, Nixon RA. Lysosome and calcium dysregulation in Alzheimer's disease: partners in crime. *Biochem Soc Trans.* Dec 2013;41(6):1495-502. doi:10.1042/BST20130201
30. Elston GN. Cortex, cognition and the cell: new insights into the pyramidal neuron and prefrontal function. *Cereb Cortex.* Nov 2003;13(11):1124-38. doi:10.1093/cercor/bhg093
31. Arnsten AF, Jin LE. Molecular influences on working memory circuits in dorsolateral prefrontal cortex. *Prog Mol Biol Transl Sci.* 2014;122:211-31. doi:10.1016/B978-0-12-420170-5.00008-8
32. Braak H, Del Tredici K. Spreading of Tau Pathology in Sporadic Alzheimer's Disease Along Cortico-cortical Top-Down Connections. *Cereb Cortex.* Sep 1 2018;28(9):3372-3384. doi:10.1093/cercor/bhy152
33. He B, Perez SE, Lee SH, Ginsberg SD, Malek-Ahmadi M, Mufson EJ. Expression profiling of precuneus layer III cathepsin D-immunopositive pyramidal neurons in mild cognitive impairment and Alzheimer's disease: Evidence for neuronal signaling vulnerability. *J Comp Neurol.* Nov 1 2020;528(16):2748-2766. doi:10.1002/cne.24929
34. Poduri A, Gearing M, Rebeck GW, Mirra SS, Tigges J, Hyman BT. Apolipoprotein E4 and beta amyloid in senile plaques and cerebral blood vessels of aged rhesus monkeys. *Am J Pathol.* Jun 1994;144(6):1183-7.
35. Skeberdis VA, Chevaleyre V, Lau CG, et al. Protein kinase A regulates calcium permeability of NMDA receptors. *Nat Neurosci.* Apr 2006;9(4):501-10. doi:10.1038/nn1664
36. Arnsten AFT, Datta D, Wang M. The genie in the bottle-magnified calcium signaling in dorsolateral prefrontal cortex. *Mol Psychiatry.* Aug 2021;26(8):3684-3700. doi:10.1038/s41380-020-00973-3
37. Veinbergs I, Everson A, Sagara Y, Masliah E. Neurotoxic effects of apolipoprotein E4 are mediated via dysregulation of calcium homeostasis. *J Neurosci Res.* Feb 1 2002;67(3):379-87. doi:10.1002/jnr.10138
38. Jiang L, Zhong J, Dou X, Cheng C, Huang Z, Sun X. Effects of ApoE on intracellular calcium levels and apoptosis of neurons after mechanical injury. *Neuroscience.* Aug 20 2015;301:375-83. doi:10.1016/j.neuroscience.2015.06.005
39. Tambini MD, Pera M, Kanter E, et al. ApoE4 upregulates the activity of mitochondria-associated ER membranes. *EMBO Rep.* Jan 2016;17(1):27-36. doi:10.15252/embr.201540614
40. Ramakrishna S, Jhaveri V, Konings SC, et al. APOE4 Affects Basal and NMDAR-Mediated Protein Synthesis in Neurons by Perturbing Calcium Homeostasis. *J Neurosci.* Oct 20 2021;41(42):8686-8709. doi:10.1523/JNEUROSCI.0435-21.2021
41. Datta D, Leslie SN, Wang M, et al. Age-related calcium dysregulation linked with tau pathology and impaired cognition in non-human primates. *Alzheimers Dement.* Jun 2021;17(6):920-932. doi:10.1002/alz.12325
42. Lacampagne A, Liu X, Reiken S, et al. Post-translational remodeling of ryanodine receptor induces calcium leak leading to Alzheimer's disease-like pathologies and cognitive deficits. *Acta Neuropathol.* Nov 2017;134(5):749-767. doi:10.1007/s00401-017-1733-7
43. Arnsten AFT, Datta D, Leslie S, Yang ST, Wang M, Nairn AC. Alzheimer's-like pathology in aging rhesus macaques: Unique opportunity to study the etiology and treatment of Alzheimer's disease. *Proc Natl Acad Sci U S A.* Dec 23 2019;doi:10.1073/pnas.1903671116
44. Popugaeva E, Pchitskaya E, Bezprozvanny I. Dysregulation of neuronal calcium homeostasis in Alzheimer's disease - A therapeutic opportunity? *Biochem Biophys Res Commun.* Feb 19 2017;483(4):998-1004. doi:10.1016/j.bbrc.2016.09.053
45. Chami M, Checler F. Alterations of the Endoplasmic Reticulum (ER) Calcium Signaling Molecular Components in Alzheimer's Disease. *Cells.* Dec 1 2020;9(12)doi:10.3390/cells9122577

46. Mattson MP. Calcium and neurodegeneration. *Aging Cell*. Jun 2007;6(3):337-50. doi:10.1111/j.1474-9726.2007.00275.x
47. Santulli G, Lewis D, des Georges A, Marks AR, Frank J. Ryanodine Receptor Structure and Function in Health and Disease. *Subcell Biochem*. 2018;87:329-352. doi:10.1007/978-981-10-7757-9_11
48. Zhang L, Trushin S, Christensen TA, et al. Altered brain energetics induces mitochondrial fission arrest in Alzheimer's Disease. *Sci Rep*. Jan 5 2016;6:18725. doi:10.1038/srep18725
49. Morozov YM, Datta D, Paspalas CD, Arnsten AFT. Ultrastructural evidence for impaired mitochondrial fission in the aged rhesus monkey dorsolateral prefrontal cortex. *Neurobiol Aging*. Mar 2017;51:9-18. doi:10.1016/j.neurobiolaging.2016.12.001
50. Kish SJ, Bergeron C, Rajput A, et al. Brain cytochrome oxidase in Alzheimer's disease. *J Neurochem*. Aug 1992;59(2):776-9. doi:10.1111/j.1471-4159.1992.tb09439.x
51. Mutisya EM, Bowling AC, Beal MF. Cortical cytochrome oxidase activity is reduced in Alzheimer's disease. *J Neurochem*. Dec 1994;63(6):2179-84. doi:10.1046/j.1471-4159.1994.63062179.x
52. Gibson GE, Sheu KF, Blass JP. Abnormalities of mitochondrial enzymes in Alzheimer disease. *J Neural Transm (Vienna)*. 1998;105(8-9):855-70. doi:10.1007/s007020050099
53. Castellani R, Hirai K, Aliev G, et al. Role of mitochondrial dysfunction in Alzheimer's disease. *J Neurosci Res*. Nov 1 2002;70(3):357-60. doi:10.1002/jnr.10389
54. Hirai K, Aliev G, Nunomura A, et al. Mitochondrial abnormalities in Alzheimer's disease. *J Neurosci*. May 1 2001;21(9):3017-23.
55. Anandatheerthavarada HK, Biswas G, Robin MA, Avadhani NG. Mitochondrial targeting and a novel transmembrane arrest of Alzheimer's amyloid precursor protein impairs mitochondrial function in neuronal cells. *J Cell Biol*. Apr 14 2003;161(1):41-54. doi:10.1083/jcb.200207030
56. Nixon RA, Saito KI, Grynspan F, et al. Calcium-activated neutral proteinase (calpain) system in aging and Alzheimer's disease. *Ann N Y Acad Sci*. Dec 15 1994;747:77-91.
57. Mahaman YAR, Huang F, Kessete Afewerky H, Maibouge TMS, Ghose B, Wang X. Involvement of calpain in the neuropathogenesis of Alzheimer's disease. *Med Res Rev*. Mar 2019;39(2):608-630. doi:10.1002/med.21534
58. Baudry M, Su W, Bi X. The calpain proteolytic system. *Encyclopedia of Cell Biology*. 2nd Edition ed2022.
59. Grynspan F, Griffin WR, Cataldo A, Katayama S, Nixon RA. Active site-directed antibodies identify calpain II as an early-appearing and pervasive component of neurofibrillary pathology in Alzheimer's disease. *Brain Res*. Jul 25 1997;763(2):145-58.
60. Ahmad F, Das D, Kommaddi RP, et al. Isoform-specific hyperactivation of calpain-2 occurs presymptotically at the synapse in Alzheimer's disease mice and correlates with memory deficits in human subjects. *Sci Rep*. Sep 3 2018;8(1):13119. doi:10.1038/s41598-018-31073-6
61. Nixon R. Calcium-activated neutral proteinases as regulators of cellular function. Implications for Alzheimer's disease pathogenesis. *Ann N Y Acad Sci*. 1989;568:198-208.
62. Siman R. Proteolytic mechanism for the neurodegeneration of Alzheimer's disease. *Ann N Y Acad Sci*. Dec 31 1992;674:193-202. doi:10.1111/j.1749-6632.1992.tb27488.x
63. Nixon R. A "protease activation cascade" in the pathogenesis of Alzheimer's disease. *Ann N Y Acad Sci*. 2000;924:117-131.
64. Nixon R. The calpains in aging and aging-related diseases. *Ageing Res Rev*. 2003;2:407-418.
65. Higuchi M, Tomioka M, Takano J, et al. Distinct mechanistic roles of calpain and caspase activation in neurodegeneration as revealed in mice overexpressing their specific inhibitors. *J Biol Chem*. Apr 15 2005;280(15):15229-37. doi:10.1074/jbc.M500939200
66. Tsai LH, Lee MS, Cruz J. Cdk5, a therapeutic target for Alzheimer's disease? *Biochim Biophys Acta*. Mar 11 2004;1697(1-2):137-42. doi:10.1016/j.bbapap.2003.11.019
67. Carragher NO. Calpain inhibition: a therapeutic strategy targeting multiple disease states. *Curr Pharm Des*. 2006;12(5):615-38. doi:10.2174/138161206775474314
68. Cagmat EB, Guingab-Cagmat JD, Vakulenko AV, Hayes RL, Anagli J. Potential Use of Calpain Inhibitors as Brain Injury Therapy. In: Kobeissy FH, ed. *Brain Neurotrauma: Molecular, Neuropsychological, and Rehabilitation Aspects*. 2015. *Frontiers in Neuroengineering*.
69. Donkor IO. An update on the therapeutic potential of calpain inhibitors: a patent review. *Expert Opin Ther Pat*. Sep 2020;30(9):659-675. doi:10.1080/13543776.2020.1797678

70. Rao MV, Mohan PS, Peterhoff CM, et al. Marked calpastatin (CAST) depletion in Alzheimer's disease accelerates cytoskeleton disruption and neurodegeneration: neuroprotection by CAST overexpression. *J Neurosci*. Nov 19 2008;28(47):12241-54. doi:10.1523/JNEUROSCI.4119-08.2008
71. Lon HK, Mendonca N, Goss S, et al. Pharmacokinetics, Safety, Tolerability, and Pharmacodynamics of Alicapostat, a Selective Inhibitor of Human Calpains 1 and 2 for the Treatment of Alzheimer Disease: An Overview of Phase 1 Studies. *Clin Pharmacol Drug Dev*. Apr 2019;8(3):290-303. doi:10.1002/cpdd.598
72. Liang B, Duan BY, Zhou XP, Gong JX, Luo ZG. Calpain activation promotes BACE1 expression, amyloid precursor protein processing, and amyloid plaque formation in a transgenic mouse model of Alzheimer disease. *J Biol Chem*. Sep 3 2010;285(36):27737-44. doi:10.1074/jbc.M110.117960
73. Sahara S, Yamashita T. Calpain-mediated Hsp70.1 cleavage in hippocampal CA1 neuronal death. *Biochem Biophys Res Commun*. Mar 19 2010;393(4):806-11. doi:10.1016/j.bbrc.2010.02.087
74. Yamashita T. Reconsider Alzheimer's disease by the 'calpain-cathepsin hypothesis'—a perspective review. *Progress in neurobiology*. 2013;105:1-23.
75. Kurbatskaya K, Phillips EC, Croft CL, et al. Upregulation of calpain activity precedes tau phosphorylation and loss of synaptic proteins in Alzheimer's disease brain. *Acta Neuropathol Commun*. Mar 31 2016;4:34. doi:10.1186/s40478-016-0299-2
76. Ferreira A, Bigio EH. Calpain-mediated tau cleavage: a mechanism leading to neurodegeneration shared by multiple tauopathies. *Mol Med*. 2011;17(7-8):676-85. doi:10.2119/molmed.2010.00220
77. Barthelemy NR, Horie K, Sato C, Bateman RJ. Blood plasma phosphorylated-tau isoforms track CNS change in Alzheimer's disease. *J Exp Med*. Nov 2 2020;217(11)doi:10.1084/jem.20200861
78. Janelidze S, Palmqvist S, Leuzy A, et al. Detecting amyloid positivity in early Alzheimer's disease using combinations of plasma Abeta42/Abeta40 and p-tau. *Alzheimers Dement*. Feb 2022;18(2):283-293. doi:10.1002/alz.12395
79. Liu F, Liang Z, Shi J, et al. PKA modulates GSK-3beta- and cdk5-catalyzed phosphorylation of tau in site- and kinase-specific manners. *FEBS Lett*. Nov 13 2006;580(26):6269-74. doi:10.1016/j.febslet.2006.10.033
80. Ma S, Liu S, Huang Q, et al. Site-specific phosphorylation protects glycogen synthase kinase-3beta from calpain-mediated truncation of its N and C termini. *J Biol Chem*. Jun 29 2012;287(27):22521-32. doi:10.1074/jbc.M111.321349
81. Jin N, Yin X, Yu D, et al. Truncation and activation of GSK-3beta by calpain I: a molecular mechanism links to tau hyperphosphorylation in Alzheimer's disease. *Sci Rep*. Feb 2 2015;5:8187. doi:10.1038/srep08187
82. Wang Y, Hall RA, Lee M, Kamgar-parsi A, Bi X, Baudry M. The tyrosine phosphatase PTPN13/FAP-1 links calpain-2, TBI and tau tyrosine phosphorylation. *Scientific Reports*. 2017;7
83. Wang Y, Brazdzionis J, Dong F, et al. P13BP, a Calpain-2-Mediated Breakdown Product of PTPN13, Is a Novel Blood Biomarker for Traumatic Brain Injury. *J Neurotrauma*. Nov 15 2021;38(22):3077-3085. doi:10.1089/neu.2021.0229
84. Chung DC, Carlomagno Y, Cook CN, et al. Tau exhibits unique seeding properties in globular glial tauopathy. *Acta Neuropathol Commun*. Mar 7 2019;7(1):36. doi:10.1186/s40478-019-0691-9
85. Cicognola C, Satir TM, Brinkmalm G, et al. Tauopathy-Associated Tau Fragment Ending at Amino Acid 224 Is Generated by Calpain-2 Cleavage. *J Alzheimers Dis*. 2020;74(4):1143-1156. doi:10.3233/JAD-191130
86. Reinecke JB, DeVos SL, McGrath JP, et al. Implicating calpain in tau-mediated toxicity in vivo. *PLoS One*. 2011;6(8):e23865. doi:10.1371/journal.pone.0023865
87. Garg S, Timm T, Mandelkow EM, Mandelkow E, Wang Y. Cleavage of Tau by calpain in Alzheimer's disease: the quest for the toxic 17 kD fragment. *Neurobiol Aging*. Jan 2011;32(1):1-14. doi:10.1016/j.neurobiolaging.2010.09.008
88. Afreen S, Ferreira A. The formation of small aggregates contributes to the neurotoxic effects of tau45-230. *Neurochem Int*. Jan 2022;152:105252. doi:10.1016/j.neuint.2021.105252
89. Croall DE, DeMartino GN. Calcium-activated neutral protease (calpain) system: structure, function, and regulation. *Physiol Rev*. Jul 1991;71(3):813-47. doi:10.1152/physrev.1991.71.3.813

90. Vanderklish PW, Bahr BA. The pathogenic activation of calpain: a marker and mediator of cellular toxicity and disease states. *International journal of experimental pathology*. 2000;81(5):323-339.
91. Huang Y, Wang KK. The calpain family and human disease. *Trends Mol Med*. Aug 2001;7(8):355-62. doi:10.1016/s1471-4914(01)02049-4
92. Geddes JW, Saatman KE. Targeting individual calpain isoforms for neuroprotection. *Exp Neurol*. Nov 2010;226(1):6-7. doi:10.1016/j.expneurol.2010.07.025
93. Wang Y, Briz V, Chishti A, Bi X, Baudry M. Distinct roles for mu-calpain and m-calpain in synaptic NMDAR-mediated neuroprotection and extrasynaptic NMDAR-mediated neurodegeneration. *J Neurosci*. Nov 27 2013;33(48):18880-92. doi:10.1523/JNEUROSCI.3293-13.2013
94. Bevers MB, Lawrence E, Maronski M, Starr N, Amesquita M, Neumar RW. Knockdown of m-calpain increases survival of primary hippocampal neurons following NMDA excitotoxicity. *Journal of neurochemistry*. 2009;108(5):1237-1250.
95. Bevers MB, Neumar RW. Mechanistic role of calpains in postischemic neurodegeneration. *Journal of Cerebral Blood Flow & Metabolism*. 2008;28(4):655-673.
96. Hardingham GE, Bading H. Synaptic versus extrasynaptic NMDA receptor signalling: implications for neurodegenerative disorders. *Nature Reviews Neuroscience*. 2010;11(10):682-696.
97. Papouin T, Oliet SH. Organization, control and function of extrasynaptic NMDA receptors. *Phil Trans R Soc B*. 2014;369(1654):20130601.
98. Xu J, Kurup P, Zhang Y, et al. Extrasynaptic NMDA receptors couple preferentially to excitotoxicity via calpain-mediated cleavage of STEP. *J Neurosci*. Jul 22 2009;29(29):9330-43. doi:10.1523/JNEUROSCI.2212-09.2009
99. Krapivinsky G, Krapivinsky L, Manasian Y, et al. The NMDA receptor is coupled to the ERK pathway by a direct interaction between NR2B and RasGRF1. *Neuron*. 2003;40(4):775-784.
100. Zadran S, Jourdi H, Rostamiani K, Qin Q, Bi X, Baudry M. Brain-derived neurotrophic factor and epidermal growth factor activate neuronal m-calpain via mitogen-activated protein kinase-dependent phosphorylation. *The Journal of Neuroscience*. 2010;30(3):1086-1095.
101. Gladding CM, Sepers MD, Xu J, et al. Calpain and STriatal-Enriched protein tyrosine phosphatase (STEP) activation contribute to extrasynaptic NMDA receptor localization in a Huntington's disease mouse model. *Human molecular genetics*. 2012;21(17):3739-3752.
102. Yamashima T. Implication of cysteine proteases calpain, cathepsin and caspase in ischemic neuronal death of primates. *Progress in neurobiology*. 2000;62(3):273-295.
103. Yamashima T. Ca²⁺-dependent proteases in ischemic neuronal death: a conserved 'calpain-cathepsin cascade' from nematodes to primates. *Cell calcium*. 2004;36(3):285-293.
104. Yamashima T. Can 'calpain-cathepsin hypothesis' explain Alzheimer neuronal death? *Ageing Research Reviews*. 2016;32:169-179.
105. Clausen A, Xu X, Bi X, Baudry M. Effects of the superoxide dismutase/catalase mimetic EUK-207 in a mouse model of Alzheimer's disease: protection against and interruption of progression of amyloid and tau pathology and cognitive decline. *Journal of Alzheimer's Disease*. 2012;30(1):183-208.
106. Páramo B, Montiel T, Hernández-Espinosa DR, Rivera-Martínez M, Morán J, Massieu L. Calpain activation induced by glucose deprivation is mediated by oxidative stress and contributes to neuronal damage. *The international journal of biochemistry & cell biology*. 2013;45(11):2596-2604.
107. Yokoyama Y, Maruyama K, Yamamoto K, et al. The role of calpain in an in vivo model of oxidative stress-induced retinal ganglion cell damage. *Biochemical and biophysical research communications*. 2014;451(4):510-515.
108. Chang H, Sheng JJ, Zhang L, et al. ROS-Induced Nuclear Translocation of Calpain-2 Facilitates Cardiomyocyte Apoptosis in Tail-Suspended Rats. *Journal of cellular biochemistry*. 2015;116(10):2258-2269.
109. Pandey M, Choudhury H, Verma RK, et al. Nanoparticles Based Intranasal Delivery of Drug to Treat Alzheimer's Disease: A Recent Update. *CNS Neurol Disord Drug Targets*. 2020;19(9):648-662. doi:10.2174/1871527319999200819095620
110. Fonseca LC, Lopes JA, Vieira J, et al. Intranasal drug delivery for treatment of Alzheimer's disease. *Drug Deliv Transl Res*. Apr 2021;11(2):411-425. doi:10.1007/s13346-021-00940-7