Medical Research Archives



3 OPEN ACCESS

Published: December 31, 2023

Citation: Davis, J., P., et.al. 2023. Documented and projected actions *in vitro* of thyroid hormone as L-thyroxine (T4) on basal cell carcinoma of the skin. Medical Research Archives, [online] 11(12).

https://doi.org/10.18103/mra.v11i12.4370

Copyright: © 2023 European Society of Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

DOI:

https://doi.org/10.18103/mra. v11i12.4370

ISSN: 2375-1924

RESEARCH ARTICLE

Documented and projected actions *in vitro* of thyroid hormone as L -thyroxine (T4) on basal cell carcinoma of the skin

Paul J. Davis^{1,2*}, Aleck Hercbergs³, Hung-Yun Lin⁴, Matthew Leinung¹, Shaker A. Mousa^{2,5}

¹Department of Medicine, Albany Medical College, Albany, NY USA; ²NanoPharmaceuticals LLC, Troy, NY USA;

³Department of Radiation Oncology, The Cleveland Clinic, Cleveland, OH USA;

⁴PhD Program for Cancer Molecular Biology and Drug Discovery, College of Medical Science and Technology, Taipei Medical University, Taipei, Taiwan;

⁵Vascular Vision Company, Troy, NY.

*pdavis.ordwayst@gmail.com

ABSTRACT

Thyroid hormone as L-thyroxine (T4) at physiological concentrations acts at its cell surface receptor on integrin $\alpha v\beta 3$ to stimulate cancer cell proliferation¹. These proliferation studies have been conducted in vitro, but pharmacological reduction of T4 and substitution of nuclear receptor ligand 3,3',5-triiodo-L-thyronine (T3) is a state of euthyroid hypothyroxinemia that has been shown clinically to arrest tumor growth in patients with cancer. T3 is inactive at physiological levels at the plasma membrane integrin receptor. A preclinical study of human basal cell carcinoma (BCC) cells has shown that the integrin thyroid hormone receptor regulates BCC radiosensitivity. While the large majority of BCCs are very manageable clinically, a small number of such tumors are aggressive. In this review of documented and proposed effects of T4 on BCC cells, we raise the possibility that BCC aggressiveness reflects T4 actions on its thyrointegrin target. The functions affected by T4 at the integrin in other human cancers include enhanced cell proliferation, anti-apoptosis, immune checkpoint regulation and metastasis, as well as state of radiosensitivity. The importance of investigating this possible pathophysiology is that euthyroid hypothyroxinemia may be tested as a treatment option.

Keywords: integrin, thyrointegrin, radioresistance, apoptosis, metastasis, L-thyroxine (T4), 3,5,3;-triiodo-L-thyronine (T3).

Introduction

Classical genomic actions of thyroid hormone (TH) are initiated at nuclear TH receptors (TRs), for which the principal ligand is 3,5,3'triiodo-L-thyronine (T3)². The major product of the thyroid gland is L-thyroxine (T4), the prohormone for T3^{1,2}. Nongenomic actions of TH are initiated at a cell surface receptor for thyroid hormone analogues extracellular domain of plasma membrane integrin $\alpha v \beta 3^{1,3}$. Functionally, the principal TH ligand of this receptor is T4. Downstream of the integrin receptor for thyroid hormone, signal transduction systems may convert T4generated signals into expression of specific genes¹. In normal adult cells, integrin $\alpha v\beta 3$ is modestly expressed and is largely in an inactive (folded) state⁴. In cancer cells and in rapidly-dividing endothelial cells, however, integrin $\alpha v\beta 3$ is usually overexpressed and is in an activated (extended) state¹. In such cells, T4 in physiological concentrations may importantly stimulate cell division modulation of transcription of specific genes. Expression of a large number of other genes in cancer cells is differentially regulated by T4 at the integrin to foster anti-apoptosis⁵, radioresistance⁶ and angiogenesis^{7,8}, as discussed in this review. Many types of cancer cells have in preclinical studies been shown to respond to T4^{1, 9-13}. Clinical evidence has shown that the growth of a variety of cancers is slowed by hypothyroidism¹⁴⁻¹⁶ and that elimination of host T4 with maintenance of euthyroidism with T3 ('euthyroid hypothyroxinemia')¹⁷ prolongs survival of cancer patients.

Cutaneous cancers shown to be responsive to thyroid hormone in clinical or preclinical studies are melanoma¹⁸⁻²¹ and squamous cell carcinoma^{22,23}. That both genomic and nongenomic molecular mechanisms of TH action are involved in these two types of cancer cells has been suggested. In an in vitro study of radioresistant human basal cell carcinoma (BCC) cells, Leith et al. described induction of radiosensitivity by tetraiodothyroacetic acid (tetrac)²⁴, a deaminated analogue of T4. This set of observations indicates that integrin $\alpha \nu \beta 3$ is expressed by these cells and is in an activated state. The report also shows that BCC cells that are radioresistant can have the latter state reversed. Finally, Leith provides a basis for speculating—because of the expression and activation of the thyrointegrin—that when BCC is sometimes difficult to manage, the T4 receptor and its ligand T4 may be contributory²⁵. In this brief review, we discuss possible contributions to BCC functional states that the thyrointegrin may support. These states described in BCC cells include radioresistance^{24,25}, pro-angiogenesis,²⁶ inhibition²⁷, checkpoint immune apoptosis^{28, 29} and metastasis³⁰. Much of this evidence has been developed via in vitro studies.

ανβ3 and Radioresistance in BCC cells

Lovett et al.³¹ found that tumor control was achieved in 91% of BCC patients treated with one or more radiotherapeutic modalities (superficial ex-irradiation, electrons or megavoltage irradiation). Achievement of control was primarily related to lesion size, so that 97% of tumors <1 cm in diameter were successfully treated and control was obtained in 87% of lesions that were 1-to-5 cm in diameter. As noted above in the Introduction,



BCC patients with radioresistance are of particular interest to us because we have shown that radioresistance in BCC cells²⁴ and in other tumor cells³² is subject to regulation by the cell surface integrin receptor for thyroid hormone and whether $\alpha v\beta 3$ is activated/extended. The activated integrin is a basis for increased intercellular affinity (integrin-integrin and integrin-extracellular matrix protein interactions) and decreased intercellular space, resulting in inhibition of cell division and, thus, radioresistance³².

Tetrac binds to the thyroid hormone receptor on integrin ntegrin ανβ3, reduces integrinintegrin affinity and restores radiosensitivity^{25,} ³². The liganding of T4 by the integrin has activating functions that deal largely downstream (signal transduction) differential expression of a large number of genes related to cancer cell self-protection¹, discussed below. systematic as investigation of possible effects of T4 on radioresistance has not yet been carried out.

αvβ3, T4 and Angiogenesis

Examined in the fertilized chicken egg chorioallantoic membrane (CAM) model, T4 at physiological concentratiomns has been enhance shown to significantly transcription of a variety of genes relevant to angiogenesis^{7, 8}. These include genes coding for vascular endothelial growth factor (VEGF), basis fibroblast growth factor (bFGF), plateletderived growth factor (PDGF) and epidermal growth factor (EGF)1,7,8. T3 is active only at supraphysiological levels in the CAM system. These effects of T4 on angiogenesis are initiated at the thyroid hormone analogue receptor on $\alpha \nu \beta 3$. We propose that this

activity of T4 on vasculature is likely to be reproduced in BCC that is metastatic.

ανβ3, T4 and Immune Checkpoint Proteins

A cohort of BCC patients was recently reported to have substantial circulating levels of soluble immune checkpoint proteins³³. These proteins included PD-1/PD-L1, CTLA-4 and TIM3. More than 50% of the 40 patients studied had infiltrating tumors, suggesting that the patients were in a state of systemic immunosuppression imposed by the checkpoint proteins and fostering aggressive skin disease. This raises the possibility that administration of checkpoint inhibitors might improve BCC management when the disease is extensive.

Acting at tumor cell surface $\alpha\nu\beta3$, T4 has been shown to regulate signal transduction pathway activities that determine cellular accumulation of PD-L1 and PD-1 that is antiapoptotic^{34, 35}. This may also serve to reduce the effectiveness of host immune checkpoint-based anti-cancer defense. Thyroid function testing and measurements of immune checkpoint proteins is needed in additional patient populations with aggressive BCC, e.g., metastatic BCC.

ανβ3, T4 and Anti-Apoptosis

Thyroid hormone as T4 has anti-apoptotic properties in cancer cells³⁶. The mechanisms include blockade of Ser-15 phosphorylation (activation) of p53 and upregulation of expression of anti-apoptotic genes such as X-linked inhibitor of apoptosis (XIAP) and hypoxia-inducible factor-1 α (HIF-1 χ). The excessive ultraviolet (UV) light that induces



BCC is also known to inhibit p53-dependent apoptosis and, via the hedgehog signaling pathway, to upregulate transcription of antiapoptotic genes³⁷. Other anti-apoptotic mechanisms may be operative in BCC³⁸ and, as mentioned above, T4 action on certain immune checkpoint proteins may inhibit apoptosis. We raise the possibility that borderline elevated levels of T4, e.g., in the upper tertile of the normal range or frankly elevated free T4 levels that may occur in non-thyroidal illness syndrome (NTIS)³⁹, may synergize with continuing host UV exposure to support aggressive behavior of BCC cells.

αvβ3, T4 and Metastasis

Metastasis of BCC occurs infrequently, but can be fatal $^{40-42}$. Mechanisms supporting BCC metastasis are incompletely defined. Because thyrointegrin $\alpha v \beta 3$ is functional in BCC cells 24 , we raise the possibility here that endogenous T4 that is elevated in NTIS 39 or simply in the upper tertile of the normal range may contribute to metastasis of tumors of many types 1,43 . We propose that this applies to BCC and that host free T4 be measured clinically in BCC patients with and without metastases to test this hypothesis.

Mousa et al.⁴³ have reviewed the mechanisms by which T4 at the TH receptor on $\alpha\nu\beta3$ may support metastasis. Integrin $\alpha\nu\beta3$ molecules on tumor cells and on the platelet surface are both subject to activation—conversion of the folded state to the extended state of the extracellular component of the integrin—by physiological concentrations of T4^{1,44}. In the T4-containing circulation in patients, platelets and free tumor cells bearing activated integrin will form complexes. The complexes are resistant to shear forces in the circulation and

resistant to natural killer (NK) cells⁴⁵. Platelet ATP freed up by T4 in the complexes may be additional support for tumor cell invasiveness⁴⁴. T4 may also potentiate the local actions of any platelet-derived growth factors (PDGFs)¹.

Tumor cell matrix metalloproteinase (MMP) gene expression (MMP-2, MMP-9) supports solubilizing the extracellular matrix at the primary tumor site and at potential sites of metastases⁴³. Thyroid hormone analogues acting at $\alpha v \beta 3$ regulate transcription of these genes; a synthetic T4 antagonist, nanodiamo-tetrac (NDAT), downregulates MMP gene expression in cancer cells and we have proposed that the primary endogenous ligand of the integrin, T4, facilitates metastasis by upregulating MMP genes⁴³.

As noted above, we have shown that T4 in the setting of cancer is highly pro-angiogenic via various mechanisms that are regulated from the thyroid hormone receptor on $\alpha v \beta 3^1$. The pro-angiogenic activity is an important support factor for tumor metastasis. Cancer driver genes are genes whose mutation may support cancer growth in the primary tumor and in metastases. Actions of thyroid hormone analogues at integrin $\alpha v \beta 3$ modulate expression of at least a dozen driver genes⁴³. We have suggested that this is another mechanism by which T4 stimulates metastasis and by which T4 antagonists retard the metastatic process⁴³.

The processes listed here are possible factors contributing to metastasis when the latter arises in s BCC.

Discussion and Conclusions

Skin cancers such as melanoma and squamous cell carcinoma (SCC) are aggressive clinical

entities that are difficult to manage. There is evidence that the TH receptor on integrin αvβ3 may mediate adverse clinical behaviors in melanoma²⁰. A role for the integtrin TH receptor in SCC has not been fully established, but T4—the principal ligand for the integrin thyroid hormone receptor—may be a growth factor for this type of cancer^{22, 23}. Our review here of the mechanisms that underlie aggressive behavior that occasionally complicates BCC was enabled by the demonstration by Leith^{24, 25, 32} that thyroid hormone analogue actions in BCC cells can alter the state of radiosensitivity. This means that $\alpha v\beta 3$ is in a functional (activated) state in BCC cells and can contribute additional procancer actions we considered here, namely, anti-apoptosis, angiogenesis and metastasis.

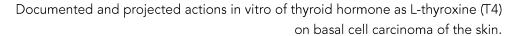
We propose that *in vitro* comparison studies of aggressive BCC cells vs. non-aggressive BCC cells be carried out to look for T4-sensitive changes in cell proliferation, radiosensitivity and metastasis. We raised the issue of immune checkpoint function in BCC cells that have activated integrin TH receptors for two reasons:

1) Lin et al. have demonstrated T4 control of PD-1/PD-L1 immune checkpoints in several types of cancer cells^{34, 35} and 2) BCC patients have been shown to have systemic evidence of increased immune checkpoint activity³³.

There are several clinical settings in which T4 actions on BCC cells at integrin $\alpha v\beta 3$ may be manifested. First, systemically ill patients with the nonthyroidal illness syndrome (NTIS) may have elevated circulating T4 levels. ³⁹. Second, we feel that the sensitivity of the integrin to T4 is such that levels of endogenous free T4 or pharmacological T4 replacement may result in free T4 concentrations in the upper tertile of the normal range; these levels may be sufficient

to induce $\alpha v \beta 3$ -mediated enhancement of cell proliferation. Third, patients with thyroid cancer whose management includes suppression of endogenous thyrotropin (TSH) with exogenous T4 may have elevated circulating free T4. While there may be an epidemiologic association of basal cell carcinoma of the skin with thyroid cancer^{46,47}, this has not yet been critically examined from the standpoints of relative dates of onset of the two conditions and aggressiveness of the skin disease.

There are potentially important therapeutic implications of the presence of activated, T4responsive $\alpha v\beta 3$ in aggressive BCC cells. We have pointed out above that induction of euthyroid hypothyroxinemia—ridding the patient of T4, but maintaining the normal metabolic acid (tetrac)—is an option. A hallmark of tumor support actions of T4 initiated at the cancer cell surface integrin is the susceptibility of such actions to blockade with tetrac1. In preclinical studies, tetrac and chemicallymodified tetrac in the absence of T4 have anticancer activity against a variety of tumor types^{11,12,50-54} and a clinical trial of chemicallymodified tetrac as an anticancer against nervous system tumor is currently in progress (SA Mousa, J Lynch, PJ Davis). The immune checkpoint actions of tetrac mentioned above are attractive for application to locally aggressive BCC55. Tetrac will also disrupt T4 action on epithelial-to-mesenchymal transition (EMT)⁵⁶ because this action is $\alpha v \beta 3$ -mediated; BCC is EMT-dependent⁵⁷. Actions of tetrac on ERBB2 and AKT driver gene expression¹ may be relevant to Hedgehog pathway signaling⁵⁸ which is prominent in BCC^{55,59}. Thus, from the standpoints of pathogenesis and possible treatment, roles for T4 in aggressive BCC require further examination.



Medical Research Archives

Funding Statement:

None

Acknowledgement Statement:

None

Conflict of Interest Statement:

Co-authors Davis and Mousa own stock in NanoPharmaceuticals, LLC.



References:

- 1. Davis PJ, Mousa SA, Lin H-Y. Nongenomic actions of thyroid hormone: the integrin component. *Physiol Rev.* 2021 Jan 1; 101(1): 319-352.
- 2. Cheng SY, Leonard JL, Davis PJ. Molecular aspects of thyroid hormone actions. *Endocr Rev.* 2010 (Apr); 31(2):139-170.
- 3. Davis PJ, Davis FB, Mousa SA, Luidens MK, Lin H-Y. Membrane receptor for thyroid hormone: physiologic and pharmacologic implications. *Annu Rev Pharmacol Toxicol*. 2011; 51:99-115.
- 4. Desgrosellier JS, Cheresh DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer*. 2010 (Jan); 10(1):9-22.
- 5. Lin H-Y, Glinsky GV, Mousa SA, Davis PJ. Thyroid hormone and anti-apoptosis in tumor cells. *Oncotarget*. 2014 Jun 20; 6(17):14735-14743.
- 6. Leith JT, Mousa SA, Hercbergs A, Lin H-Y, Davis P. Radioresistance of cancer cells, integrin $\alpha \nu \beta 3$ and thyroid hormone. *Oncotarget*. 2018 Dec11; 9(97):37069-37075.
- 7. Luidens MK, Mousa SA, Davis FB, Lin H-Y, Davis PJ. Thyroid hormone and angiogenesis. *Vascul Pharmacol.* 2010 Mar-Apr; 52(3-4):142-145.
- 8. Rajabi M, Mousa SA. The roles of angiogenesis in cancer treatment. *Biomedicines*. 2017 Jun; 5(2):34.
- 9. Davis PJ, Lin H-Y, Hercbergs A, Mousa SA. Actions of L-thyroxine (T4) and tetraiodothyroacetic acid (tetrac) on gene expression in thyroid cancer cells. *Genes* (*Basel*). 2020 Jul 7; 11(7):755.
- 10. Davis PJ, Glinsky GV, Lin HY, Leith JT, Hercbergts A, Tang HY, Ashur-Fabian O,

- Incerpi S, Mousa SA. Cancer cell gene expression modulated from plasma membrane integrin $\alpha v \beta 3$ by thyroid hormone and nanoparticulate tetrac. *Front Endocrinol (Lausanne)*. 2015 Jan 12; 5:240
- 11. Shinderman-Maman E, Cohen K, Weingarten C, Nabriski D, Twito O, Baraf L, Hercbergs A, Davis PJ, Werner H, Ellis M, Ashur-Fabian O. The thyroid hormone-ανβ3 integrin axis in ovarian cancer: regulation of gene transcription and MAPK-dependent proliferation. *Oncogene*. 2016 Apr 14; 35 (15):1977.
- 12. Cohen K, Flint N, Shalev S, Erez D, Baharal T, Davis PJ, Hercbergs A, Ellis M, Ashur-Fabioan O. Thyroid hormone regulates adhesion, migration, and matrix metalloproteinase 9 activity via ανβ3 integrin in myeloma cells. *Oncotarget*. 2014 Aug15;5 (15):6312-6322.
- 13. Hercbergs A, Lin H-Y, Mousa SA, Davis PJ. (Thyroid) Hormonal regulation of breast cancer cells. *Front Endocrinol (Lausanne)*. 2023 Jan 11; 13:1109555.
- 14. Schmidinger M, Vogl UM, Bojic M, Lamm W, Heinzl H, Haitel A, Clodi M, Kramer G, Zielinski CC. Hypothyroidism in patients with renal cell carcinoma: blessing or curse? *Cancer.* 2011 Feb 1; 117(3):534-544.
- 15. Cristofanilli M, Yamamura Y, Kau S-W, Beveers T, Strom S, Patangan M Hsu L, Krishnamurthy S, Theriault RL, Horotbagyi GN. Thyroid hormone and breast cancer. Primary hypothyroidism is associatred with a reduced incidence of primary breast carcinoma. *Cancer.* 2005 Mar 15; 103(6):1122-1128
- 16. Mathew A, Fuhrer D, Lahner H. Sunitinibinduced hypothyroidism and survival in

pancreatic neuroendocrine tumors. *Horm Metab Res.* 2021(Dec); 53(12):794-800.

- 17. Hercbergs A, Johnson RE, Ashur-Fabian O, Garfield DH, Davis PJ. Medically induced euthyroid hypothyroxinemia may extend survival in compassionate need cancer patients: an observational study. *Oncologist.* 2015 Jan; 20(1):72-76.
- 18. Bilen MA, Patel A, Hess KR, Munoz J, Busidy NL, Whelan JJ, Janku F, Falchook GS, Hong DS, Meric-Bernstam FM, Habar MA, Naing A. Association between new-onset hypothyroidism and clinical response in patients treated with tyrosine kinase inhibitor therapy in phase I clinical trials. *Cancer Chemother Pharmacol*. 2016 Jul; 78(1):167-171.
- 19. Atkins MB, Mier JW, Parkinson DR, Gould JA, Berkman EM, Kaplan MM. Hypothyroidism after treatment with interleukin-2 and lymphokine-activatd killer cells. *N Engl J Med.* 1988 Jun 16; 318(24):1557-1563.
- 20. Fabian ID, Rosner M, Fabian I, Vishnevskia-Dan V, Zioto O, Shinderman Maman E, Cohen K, Ellis M, Lin H-Y, Hercbergs A, Davis PJ, Ashur-Fabian O. Low thyroid hormone levels improve survival in murine model for ocular melanoma. *Oncotarget*. 2015 May 10; 6(13): 11038-11046.
- 21. Wu Y, Wang Z, Bai H, Gao Y. Thyroid dysfunction during PD-1 inhibitor treatment in patients with cancer: incidence and association with progression-free survival. *Oncol Lett.* 2022 Jul 13; 24(3):309.
- 22. Miro C, Di Cicco E, Ambrosio R, Mancir G, Di Girolamo D, Cicatiello AG, Sagliocchi S, Nappi A, De Stefano MA, Luongo C, Antonini D, Visconte F, Varricchio S, Ilardi G, Del Vecchio L, Staibano S, Boelen A, Bianpain C, Missero C, Salvatore D, Dentice M. Thyroid

- hormone induces progression and invasiveness of squamous cell carcinomas by promoting a ZEB-1/E-cadherin switch. *Nat Commun.* 2019 Nov 27; 10 (1):5410.
- 23. Lupulesco A. Hormonal regulation of epidermal tumor development. *J Invest Dermatol.* 1981 Aug; 77(2):186-195.
- 24. Leith JT, Davis PJ, Mousa SA, Hercbergs AA. In vitro effects of tetraiodothyroacetic acid combined with X-irradiation on basal cell carcinoma cells. *Cell Cycle*. 2017 Feb 16; 16(4):367-373.
- 25. Leith JT, Mousa SA, Hercbergs A, Lin H-Y, Davis PJ. Radioresistance of cancer cells, integrin $\alpha \nu \beta 3$ and thyroid hormone. *Oncotarget*. 2018 Dec; 9(97):37069-37075.
- 26. Lupu M, Caruntu C, Popa MI, Voiculescu VM, Zurac S, Boda D. Vascular patterns in basal cell carcinoma: dermoscopic, confocal and histopathological perspectives. *Oncol Lett.* 2019 May; 17(5):4112-4125.
- 27. Moujaess E, Merhy R, Kattan J, Sarkis A-S, Tomb R. Immune checkpoint inhibition for advanced or metastatic basal cell carcinoma: how much evidence do we need? *Immunotherapy.* 2021 Oct; 13(15):1293-1304.
- 28. Koyun E, Karadag R, Ozkanli S, Oguztuzun S, Kocdogan AK. Caspase-3, p53 and Bcl-2 expression in basal cell carcinoma of the eyelid. *Postepy Dermatol Alergol.* 2020 Aug; 37(4):535-539.
- 29. Mendez-Flores RG, Martinez-Fernandez DE, Vega-De La Torre DE, Zambrano-Roman M, Munoz-Valle JF, Toledo-Lelewer MG, Guevara-Gutierrez E, Ramirsez-Padilla M, Vales-Alvarado E. Role of Bcl-2, p53, and Ki-67 expression in basal cell carcinoma and their association with aggressive and non-

- aggressive histological phenotypes. *Postepy Dermatol Allergol.* 2022 Jun; 39(3):517-523.
- 30. Laga AC, Schaefer IM, Sholl LM, Fremnch CA, Hanna J. Metastatic basal cell carcinoma. Am J Clin Pathol. 2019(Nov 4); 152(6):706-717.
- 31. Lovett RD, Perez CA, Shapiro ST, Garcia DM. External irradiation of epithelial skin cancer. *Int J Radiol Oncol Biol Phys.* 1990 (Aug); 19(2):235-242.
- 32. Leith JT, Hercbergs A, Kenney S, Mousa SA, Davis PJ. Activation of tumor cell integrin $\alpha v\beta 3$ by radiation and reversal of activation by chemically modified tetraiodothyroacetic acid (tetrac). *Endocr Res.* 2018 Nov; 43(4):215-219.
- 33. Malinga NZ, Siwele SC, Steel HC, Kwofie LLI, Meyer PWA, Smit T, Anderson R, Rapoport BL, Kgokolo MCM. Systemic levels of the soluble co-inhibitory immune checkpoints, CTLA-4, LAG-3, POD-1/PD-L1 and TIM-3, are markedly increased in basal cell carcinoma. *Translat Oncol.* 2022; 19:101384.
- 34. Lin HY, Chin YT, Shin YG, Chen YR, Leinung M, Keating KA, Mousa SA, Davis PJ. In tumor cells, thyroid hormone analogues non-immunologically regulate PD-L1 and PD-1 accumulation that is anti-apoptotic. *Oncotarget*. 2018 Sept 24; 9(75):34033-34037.
- 35. Lin HY, Chin YT, Nana AW, Shih YJ, Lai HY, Tang HY, Leinung M, Mousa SA, Davis PJ. Actions of L-thyroxine and nano-diaminotetrac (Nanotetrac) on PD-L1 in cancer cells. *Steroids*. 2016 Oct; 114:59-67.
- 36. Lin H-Y, Glinsky GV, Mousa SA, Davis PJ. Thyroid hormone and anti-apoptosis in tumor cells. *Oncotarget*. 2015 Jun 20; 6(17):14735-14743.
- 37. Erb P, Ji J, Wernli M, Kump E, Glaser A, Buchner SA. Role of apoptosis in basal cell

- and squamous cell carcinoma formation. *Immunol Lett.* 2005 Aug 15; 100(1):68-72.
- 38. Jee SH, Shen SC, Chiu HC, Tsai WL, Kuo ML. Overexpression of interleukin-6 in human basal carcinoma cell lines increases antiapoptotic activity and tumorigenic potency. *Oncogene*. 2001 Jan 11; 20(2):198-208.
- 39. Hercbergs A, Mousa SA, Davis PJ. Nonthyroidal illness syndrome and thyroid hormone actions at integrin $\alpha v\beta 3$. *J Clin Endocrinol Metab.* 2018 Aptr; 103(4):1291-1295.
- 40. McCuskey M, Basset-Seguin N, Dummer R, Lewis K, Schadendorf D, Sekulic A, Hou J, Wang L, Yue H, Hauschid A. Metastatic basal cell carcinoma: prognosis dependent on anatomic site and spread of disease. *Eur J Cancer*. 2014 Mar; 50(4):774-783.
- 41. Laga AC, Schaefer IM, Sholl LM, French CA, Hanna J. Metastatic basal cell carcinoma. *Am J Clin Pathol.* 2019 Nov 4; 152(6):707-717.
- 42. Mochel MC, Liaquat S, Moore JB, Hoang MP. Metastatic basal cell carcinoma: a clinicopathologic and immunohistochemical study of 22 cases. *J Cutan Pathol.* 2021 Mar; 48(3):374-383.
- 43. Mousa SA, Glinsky GV, Lin H-Y, Ashur-Fabian O, Hercbergs A, Keating KA, Davis PJ. Contributions of thyroid hormone to cancer metastasis. *Biomedicines*. 2018 Aug 22; 6(3):89.
- 44. Davis PJ, Mousa SA, Schechter GP, Lin H-Y. Platelet ATP, thyroid hormone receptor on integrin $\alpha v\beta 3$ and cancer metastasis. *Horm Cancer.* 2020 Feb; 11(1):13-16.
- 45. Schlesinger M. Role of platelets and platelet receptors in cancer metastasis. *J Hematol Oncol.* 2018; 11:125.
- 46. Krilaviciute A, Vincerzevskiene I, Smailyte G. Basal cell skin cancer and the risk of second



- primary cancers: a cancer registry-based study in Lithuania. *Ann Epidemiol*. 2016 Jul; 26(7):511-514.
- 47. Friedman GD, Tekawa IS. Association of basal cell skin cancers with other cancers (United States). *Cancer Causes Control.* 2000 Dec; 11(10)891-897.
- 48. Davis PJ, Tang HY, Hercbergs A, Lin HY, Keating KA, Mousa SA. Bioactivity of thyroid hormone analogs at cancer cells. *Front Endocrinol (Lausanne)*. 2018 Dec 4; 9:739.
- 49. Davis PJ, Glinsky GV, Lin HY, Leith JT, Hercbergs A, Tang HY, Ashur-Fabian O, Incerpi S, Mousa SA. Cancer cell gene expression modulated from plasma membrane integrin $\alpha \nu \beta 3$ by thyroid hormone and nanoparticulate tetrac. *Front Endocrinol (Lausanne)*. 2015 Jan 12; 5:240.
- 50. Sudha T, Godugxu K, Darwish NHE, Nazeer T, Mousa SA. Novel polyethylene glycol-conjugated triazole derivative with high thyrointegrin $\alpha \nu \beta 3$ affinity in acute myeloid leukemia management. *Cancers (Basel)*. 2921 Aug13;1 3(16):4070.
- 51. Gionfra F, De Vito P, Pallottini V, Lin H-Y, Davis PJ, Pederson JZ, Incerpi S. The roles of thyroid hormones in hepatocyte proliferation and liver cancer. *Front Endocrinol (Lausanne)*. 2019 Aug 30; 10:532.
- 52. Mousa SA, Yalcin M, Bharali DJ, Meng R, Tang H-Y, Lin H-Y, Davis FB, Davis PJ. Tetraiodotnhyroacetic acid and its nanoformulation inhibit thyroid hormone stimulation of non-small cell lung cancer cells in vitro and its growth in xenografts. *Lung Cancer*. 2012 Apr; 76(1):39-45.
- 53. Chin Y-T, He Z-R, Chen C-L, Chu H-C, Ho Y, Su P-Y, Yang Y-C SH, Wang K, Shih Y-J, Chen Y-R, Pedersen JZ, Incerpi S, Nana AW,

- Tang H-Y, Lin H-Y, Mousa SA, Davis PJ, Whang-Peng J. Tetrac and NDAT induce antiproliferation via integrin $\alpha v \beta 3$ in colorectal cancers with different K-RAS status. Front Endocrinol (Lausanne). 2019 Mar 12; 10:130.
- 54. Coskun MD, Sudha T, Bharali DJ, Celikler S, Davis PJ, Mousa SA. $\alpha v \beta 3$ Integrin antagonists enhance chemotherapy response in an orthotopic pancreatic cancer model. *Front Pharmacol.* 2020 Feb 27; 11:95.
- 55. Gupta N, Ruiz ES. Current perspectives in the treatment of locally advanced basal cell carcinoma. *Drug Design Development Ther*.2022; 16:183-190.
- 56. Weingarten C, Jenudi Y, Tshuva RY, Moskovich D, Alfanda A, Hercbergs A, Davis PJ, Ellis M, Ashur-Fabian O. The interplay between epithelial-mesenchymal treansition (EMT) and the thyroid hormone- $\alpha\nu\beta$ 3 axis in ovarian carcinoma. *Horm Cancer*. 2018 Feb; 9(1):22-32.
- 57. Lupu M, Caruntu C, Ghia MA, Voiculescu V, Voiculescu S, Rosca AE, Caruntu A, Moraru L, Popa IM, Calenic B, Greabu M, Costea DE. Gene expression and proteome analysis as sources of biomarkers in basal cell carcinoma. *Disease Markers*. 2016; article ID 9831237.
- 58. Gajjar A, Robinson GW, Smith KS, Lin T, Merchant TE, Chintagumpala M, Mahajan A, Su J, Bouffet E, Bartels U, Schechter T, Hassall T, Robertson T, Nicholls W, Gururangan S, Schroeder K, Sullivan M, Wheeler G, Hansford JR, Kellie S, McCowage G, Cohn R, Fisher MJ, Krasin MJ, Stewart CF, Broniscer Buchhalter I, Tatevossian RG, Orr BA, Neale G, Klimo P Jr, Boop F, Srinivasan A, Pfister Sm, Gilbertson RJ, Onar-Thomas A, Ellison DW, Northcott PAS. Outcomes byt clinical and molecular with features in children



medulloblastsoma treated with risk-adapted therapy: results of an international Phase III trial (SJMB03). *J Clin Oncol.* 2021 Mar 1; 39(7)-822-835.

59. Grund-Groschka S, Ortner D, Szenes-Nagy AB, Zaborsky N, Weiss R, Neureite D, Wipplinger M, Risch A, Hammerl P, Greil R, Sibilia M, Gratz IK, Stoitzner P, Aberger F. Epidermal activation of Hedgehog signaling establishes an immunosuppressive microenvironment in basal cell carcinoma by modulating skin immunity. *Molec Oncol.* 2020; 14:1930-1946.