

Published: February 29, 2024

Citation: AY Ahmed, HL Abdullahi, et al., 2024 Effect of Estrogen Beta Receptor Polymorphism in Male Infertility: A Review, Medical Research Archives, [online] 12(2). https://doi.org/10.18103/mra.v 12i2.5009

Copyright: © 2024 European Society of Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. **DOI**

<u>https://doi.org/10.18103/mra.v</u> 12i2.5009

ISSN: 2375-1924

REVIEW ARTICLE

Effect of Estrogen Beta Receptor Polymorphism in Male Infertility: A Review

Ahmed, A.Y^{*1}, Abdullahi, H.L²., Gwarzo M.Y²., Maryam, I. R¹., Yusuf, A¹., Abdulkadir, S.S¹., Muhammad, Y⁴., Gamde, S.M³.

¹Department of Medical Laboratory Science, Kaduna State University, Nigeria.

²Department of Medical Laboratory Science, Bayero University Kano, Nigeria.

³Department of Medical Laboratory Science, Bingham University Karu, Nigeria.

⁴Department of Medical Laboratory Science, Rasheed Shekoni Teaching Hospital, Jigawa, Nigeria.

*Corresponding author: <u>ahmedarmiyau@gmail.com</u> or <u>ahmed.armiyau@kasu.edu.ng</u>

ABSTRACT

Infertility is one of the major health problems of reproductive age couples. The suggestion that oestrogen beta receptor polymorphism has unfavorable effects on the male fertility is contentious. This review study aimed to identify the impact of estrogen beta receptor polymorphism on male infertility. The comprehensive literature review focused on the role of estrogen beta receptor (ER β) in the testis, its effects on sperm motility, longevity, and fertilizing capacity, as well as the relationship between $ER\beta$ mutation and male infertility. The review highlighted the critical role of ERB in sperm function, emphasizing its strong expression in the testis and its significance for sperm motility and fertilizing capacity. The absence of $ER\beta$ was found to diminish epididymal sperm content, sperm motility, and fertility, thereby contributing to male infertility. Additionally, the review discussed the influence of environmental, physiological, and genetic factors on male factor infertility and emphasized the need for further molecular studies to determine the implications of ERB mutation in a multifactorial manner.

The literature cited in the review provided insights into the prevalence of male infertility worldwide, with a focus on the African region, where male factor infertility was found to account for a significant percentage of infertility cases. Genetic evaluations and lifestyle changes, including the inclusion of antioxidant vitamins and minerals in treatment regimens to prevent oxidative DNA damage linked to estrogen-induced effects on male reproduction, were recommended. Furthermore, healthy lifestyle choices and the avoidance of tobacco smoking, marijuana, and alcohol use may play central role in preserving male fertility.

Introduction

Infertility is one of the major health problems of reproductive age couples. People begin to show concern if conception cannot be achieved after 12 months of regular unprotected intercourse with increasing incidence rates in males ¹. Without a doubt, semen analysis remains the first step in the diagnosis of male infertility. Research findings associated approximately 50% of childlessness to male factor infertility, while women have been wrongly stigmatized for inability to conceive ². This is in most cases is due to cultural believe and insufficient knowledge and misunderstanding. Male infertility is well-thought-out when identifiable female causes of infertility are excluded and semen quantity and quality fail to fulfill WHO criteria ³. Essentially, factors identified in sperm dysfunction and male factor infertility are mainly environmental, physiological, and genetic ¹.

Researchers had revealed that male factors account for 40-50% of infertility in human. Male infertility is commonly due to deficiencies in the semen and semen quality is used as a surrogate measure of male fertility. It is estimated that 60% of married couples having regular unprotected sexual intercourse, achieve pregnancy after 6 months of co-habitation, 90% achieve pregnancy by 12 months and 95% between 18-24 months ⁴. In more than 50% of male infertility cases, the causes remain unknown, and the infertility is thus classified as idiopathic. Idiopathic infertility affects a high percentage of infertile men who cannot be effectively treated by the treatment modalities available ⁴.

Male-factor infertility is a notable health issue all over the world including Africa and other developing nations. The prevalent rate differs between and within countries. For instance, in the United Kingdom and the United States of America, it is estimated to be 6% and 10% respectively⁵.

The prevalence of infertility in Sub-Sahara Africa ranges from 20% to 40%. Although the Africa socio-cultural setting has before now focused on the female, fertility problems are shared by both male and female sexes ⁶. Male factor is responsible for 40-50% of all infertility in Nigeria although it varies from one region to another, and the causes also vary from place to place. A study in the midwestern Nigeria showed that about 50% of the 780 couples evaluated were observed to have varied causes of infertility ⁶.

In the southwest, male factor was reported to be responsible for 42.4% infertility cases, while in Maiduguri, North-Eastern Nigeria, infertility is the reason for about 40% of all gynecological consultations. In Kano, 40.8% prevalence was reported, 46% in lle-lfe and 55–93% was observed in Enugu, Eastern Nigeria for male factor infertility ⁶. Similarly, some countries, most notably Kenya, Gabon, Botswana, Zimbabwe and many other African countries, have shown a pattern toward lower fertility ⁵.

Aim and scope

The aim of this paper is to investigate the impact of estrogen beta receptor (ER β) polymorphism on male infertility. The review aims to explore the role of ER β in male fertility by examining its expression in the testis, its effects on sperm motility and fertilizing capacity, and the potential relationship between ER β mutation and male infertility. The paper seeks to provide a comprehensive understanding of the functions of estrogen receptors, particularly ER β , in male reproductive health and to explore the potential implications of ER β polymorphism in contributing to male infertility.

The scope of the paper includes a thorough review of literature on ER β and its role in male infertility, encompassing its impact on sperm function, its association with idiopathic infertility, and the prevalence of male infertility in different regions, particularly in African countries. Additionally, the paper aims to shed light on the potential genetic, environmental, and physiological factors contributing to male factor infertility and emphasizes the need for further molecular studies to understand the multifactorial implications of ERB mutation in male infertility. Overall, the paper aims to contribute to the existing body of knowledge on male infertility, with a particular focus on the role of ERB polymorphism, and to provide insights that may guide future research and potential therapeutic interventions.

This is a systematic review conducted by first of all using PubMed, Google scholar, research gate with the following headings; "male infertility", "functions of estrogen receptor", spermatogenesis", and "beta receptor polymorphism" to select the articles that are in tandem with the review objectives from over a one hundred down to just about 60. Using metaanalysis to resolve the protocol, research questions and idea validation.

Why are the results of this paper important?

The results of this study are important for several reasons:

1. Understanding Male Infertility: The study sheds light on the complex factors contributing to male infertility, including the role of estrogen beta receptor polymorphism. By highlighting the impact of $ER\beta$ on sperm motility, longevity, and fertilizing capacity, the study provides valuable insights into the molecular and genetic mechanisms underlying male infertility.

- Clinical and Research Implications: The findings have implications for clinical practice, as they emphasize the need for comprehensive genetic evaluations and personalized treatment modalities to address male factor infertility. Furthermore, the study underscores the importance of further molecular research to elucidate the multifactorial implications of ERβ mutation in male infertility.
- 3. Public Health Significance: Male infertility is a significant public health issue globally, and the study's findings contribute to raising awareness about the prevalence of male factor infertility, particularly in African countries. By recognizing the high percentage of idiopathic infertility cases and the varying prevalence rates in different regions, the study underscores the importance of addressing male infertility as a crucial reproductive health concern.
- 4. Lifestyle and Environmental Counseling: The study also highlights the importance of promoting healthy lifestyle choices and the avoidance of factors such as tobacco smoking, marijuana, and alcohol use, which may contribute to sperm DNA damage and male factor infertility. This emphasizes the need for counseling and education to promote male reproductive health and fertility.

In summary, the results of this study are important as they contribute to advancing the understanding of male infertility, highlighting the role of ER β polymorphism, and emphasizing the need for comprehensive approaches to address male factor infertility and promote reproductive health. The findings have implications for clinical practice, research, and public health interventions aimed at addressing male infertility globally.

Oestrogen

Oestrogens play vital roles in the development and maintenance of male reproductive function and fertility. Oestrogen activity is mediated through two receptors (ESR), ESR1 and ESR2 (otherwise called ERa and ERß respectively) that belong to the nuclear receptor family of a transcription factors. ERa is known to be strongly expressed in the epididymis, efferent ductules and Leydig cells, whereas ERß is predominantly expressed in germ cells, especially in the primary spermatocytes and round spermatids of the human testis. The ESRß gene is situated on chromosome 14q22-24, comprising 8 exons that cover about 40 kb. ESR β codes for a 530 amino acid protein ⁷. Quite a lot of sequence variants of the ESR β gene have also been identified which include 2 silent G/A polymorphisms, Rsal (rs1256049) and Alul (rs4986938) ⁷.

In human testis, ER β has also been found in ejaculated spermatozoa ⁷. It also affects motility and fertilizing capacity. In males, ER β polymorphism was suggested to have an effect on azoospermia or idiopathic severe oligospermia.

The relationship between Rsal polymorphism (rs1256049) with male infertility has been documented in Caucasian patients. It is located in exon 5 of ER β . Existing data suggest that ER β may also play an important role in male infertility ⁸.

Oestrogen and male fertility

Approximately 15% of male infertile cases are related to genetic factors, including chromosomal aberrations and single gene mutations, which may result in spermatogenic failure and sperm dysfunction ⁹. The traditional understanding of oestradiol as the 'female' hormone and of testosterone as the 'male' hormone has been challenged due to the increased interest in elucidating the role of oestrogen in males. Oestrogens are produced in the male reproductive system by Sertoli cells, Leydia cells, and aerm cells. In addition, studies have shown that oestrogens reduce testosterone production from Leydig cells and reduce Sertoli cell numbers in adult when they are given during development ¹⁰. The oestrogens can also disrupt fetal Leydig cell development, inhibit apoptosis of human post meiotic germ cells, and increase spermatogonia number per testis.

Oxidative DNA damage may be involved in oestrogen induced effects on male reproduction. Oestrogen excess during the adulthood can deteriorate sperm production and maturation. Oestrogen acts both peripherally and in the central nervous system. The physiological responses to oestrogens are modulated by the oestrogen receptors (ER1 and ER2) genes ¹¹.

Oestrogen has been reported as a survival factor for germ cells, involving in the induction of oxidative DNA damage, and the aberrant level of oestrogen may lead to impaired sperm production. It has been shown that free radicals inhibit steriodogenesis by interfering with cholesterol transport to the mitochondria and/or the catalytic function of P450 enzymes, which leads to an increase in lipid per oxidation and decline in the antioxidant barrier ¹². Moreover, oestrogens can regulate mitochondrial function by increasing nuclear respiratory factor-1 (NRF-1) expression. Specifically, oestradiol stimulates mitochondrial function through a genomic mechanism of ER action involving direct ER α and ER β interaction with an oestrogen response element in the NRF-1 promoter.

In vivo knockdown experiments have indicated that oestradiol stimulates NRF-1 transcription and consequently increases mitochondrial biogenesis through ER α activity but not through ER β activity in MCF-7 breast cells. This experiment indicates that ER α polymorphisms can increase mitochondrial activity via NRF-1 transcription in human ejaculated spermatozoa, presenting them with high motility ¹⁰.

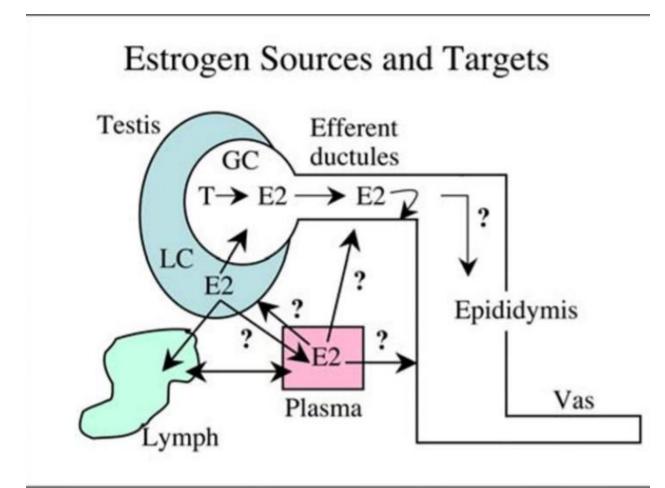


Figure 1: Estrogen sources and targets in the male reproductive tract ¹³.

Estrogen sources and targets in the male reproductive tract. Estradiol 176 (E2) is produced in peripheral tissues and delivered via the plasma, but is also synthesized by Leydig cells (LC) in the testicular interstitium.

Oestrogen receptors

The physical functions of oestrogens were involved in the oestrogen receptors (ERs). Moreover, ERs are members of the nuclear receptor (NR) superfamily that mediates the pleiotropic effects of oestrogen in a diverse range of developmental and physiological processes, playing an important role in mediating oestrogen action on target tissues.

ERs have been identified to be two subtypes of ER α and ER β . ER α is a 595-amino acid protein encoded by the ERs1 gene on chromosome 6q25, and ER β is a 530-amino acid protein which encoded by the ERs2 gene on chromosome 14q22-24. Genetic screening for the ER α gene locus has revealed several polymorphic sites, and two polymorphisms located in ER α intron 1(T/C transition, rs2234693) and in 50 bp downstream of the former one (G/A transition, rs9340799) have been widely concerned. In addition, the ER β genes have been described with two silent G/A polymorphisms (rs1256049 and rs4986938). ¹⁰.

Oestrogen receptors and male fertility

Both ER α and ER β receptors are expressed in the testis and in the epididymis. In human testis, ER α and ER β have also been found in ejaculated spermatozoa. It has been shown that, the absence

of ER β leads to decreased epididymal sperm content, decreased sperm motility and fertilizing capacity ⁸. Association between male infertility and polymorphisms in ER α and ER β genes has been shown in few studies. Polymorphisms in ER α gene (Xbal and Pvull) have been shown to be associated with azoospermia or severe oligozoospermia. Overall, studies regarding the effect of ER genes on male fertility have produced conflicting evidence ¹¹.

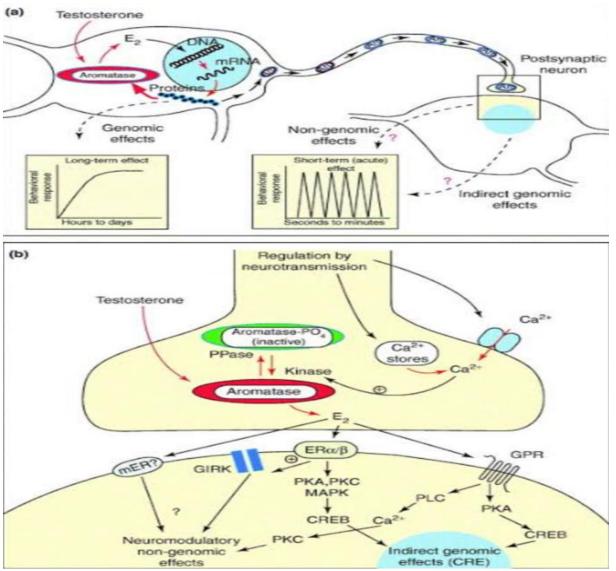
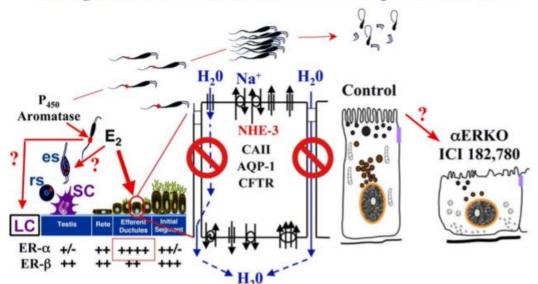


Figure 1: Effect of Estrogen Beta Receptor in Male Reproductive Tract.²

Oestrogen beta receptor polymorphisms in male infertility

The mechanisms behind altered ERs function in subjects with polymorphisms remain unclear. The polymorphism rs1256049 located at the splice acceptor site just prior to exon 8 in ER β and may potentially affect the splicing of this exon, leading to proteins with different properties than the wild-type ER β^{14} . In addition, studies have reported the polymorphism could also have a direct effect

through changing the nucleotide sequence and thereby the secondary structure of the ER β mRNA, possibly leading to changes in the function of mRNA ¹⁵. It has been reported that ER β gene polymorphisms (rs2234693 and rs9340799) may modulate the effect of oestradiol on CYP19, which encodes aromatase expression, disrupting the gene causes a decline in sperm numbers and loss of male infertility ¹¹.



Estrogen and Its Inhibition in the Male Reproductive Tract

Figure 3: Estrogen and its inhibition in the male reproductive tract¹³

In adult males (Figure 3), germ cells, as well as Leydig cells (LC) contain P450 aromatase and actively synthesize estrogen (E2), which produces a relatively high concentration in rete testis fluid. This luminal estrogen targets estrogen receptors that are abundant throughout the male reproductive tract, but particularly ER α that is localized in the efferent ductile epithelium, where its expression is more abundant than even the female reproductive tract. In the testis, E2 may also feedback to influence the function of LC and spermatids, either round spermatids or elongated spermatids. Estrogen's primary function in the male tract is the regulation of fluid reabsorption in the efferent ductile via $ER\alpha$, which increases the concentration of sperm prior to entering the epididymis. Disruption of $ER\alpha$, either in the knockout (α ERKO) or by treatment with a pure antiestrogen ICI 182,780, results in a decrease in Na+ transport from lumen to interstitial and thus a decrease in water (H2O) and fluid reabsorption¹³.

The precise role of oestrogen receptors in male fertility status is understood. Some findings suggest that specific polymorphisms of the ER α , and ER β genes which confer a lower sex hormone binding globulin (SHBG) and thus a stronger unbound oestrogen effect, may adversely affect human spermatogenesis. SHBG is involved in both delivering reproductive hormones to target tissues and controlling the concentration of androgens and oestrogens in the serum and tissues. It has been established that infertile men with severe oligozoospermia had significantly lower T (testosterone) and higher E2 (oestradiol)

concentrations than fertile men, resulting in an elevated T/E2 ratio 11 .

Oestrogen beta receptors and spermatozoa

In human sperm, ERs were found differently located being both receptors in the midpiece, while ER β continues to be expressed in the flagellum. Sperm are able to synthesize oestrogen raising the possibility that they not only will be exposed to oestrogens in female genital tract but provide themselves a persisting local source of oestrogen ¹⁶. This autocrine loop E2/ERs may modulate the extraejaculation sperm acquisition of fertilizing ability. In ejaculated sperm, E2 stimulates various sperm functions including motility, longevity, capacitation, and acrosome reaction ¹⁶.

In the male reproductive tract, a key action of oestrogens was observed in the regulation of the spermatogenesis, spermiogenesis, and gamete functional maturation ¹⁶.

The highly polarization in the structure and function of spermatozoa compartmentalize specific signaling pathways to the regions where they are needed. The massive presence of ER β in the flagellum may indicate an important role in the sperm motility. In this regard, it was previously reported that oestrogens affect sperm motility ¹⁷. In sperm, flagellum serves dual functions: in addition to being a basic engine for locomotion it is involved in physiological functions such as capacitation and acrosome reaction. Besides, most of the glycolytic enzymes have been localized to the flagellum and the central role of mitochondria in energy production is well known. The concomitant location of the two ERs in the flagellum and midpiece, where mitochondria are present, led us to hypothesize that these receptors could be involved in sperm metabolism ¹⁶.

Oestrogen beta receptor and the testis

Both ERs are expressed more extensively in adult testes compared to the juvenile ones. However, this expression pattern is not the same for all testicular cell types. In the case of Sertoli cells, the expression level of ER α declines, while the expression of ER β increases with age. This switch between ER α and ER β expression allows E2 to mediate its effects in distinct directions and regulate proliferation of immature Sertoli cells, while later the same signal pushes these cells towards differentiation ¹⁸.

Sensitivity to oestrogen hormones also depends on availability of oestrogen receptor co-regulators. E2 treatment leads to an increase in the recruitment of ERB and its co-repressor NCoR1 (nuclear receptor co-repressor 1) to the ERE of Arpc1b (actin related protein 2/3 complex subunit 1B) gene, thus causing downregulation of Arpc1b transcription in testes¹⁹. In contrast, the recruitment of $ER\beta$ and its coactivator Src1 (steroid receptor co-activator 1) is decreased in Evl (Ena-vasodilator stimulated phosphoprotein) ERE after E2 treatment, thus causing down regulation of Evl. Both, Arpc1b and Evl are involved in actin remodeling during spermiation, and they are responsive to E2 but not to dihydrotestosterone. Src1, Src2 (also named TIF2, GRIP1), and Src3 (also named p/CIP, RAC3, ACTR, AIB1) belong to the p160 family of nuclear receptor co-activators. Even though the Src1 null male mouse mutants are fertile, they have a smaller but histologically normal testis. The effect is more pronounced in the TIF2-/- (Src2-/-) male mice, which have impaired fertility with defects in spermiogenesis and age-dependent testicular degeneration²⁰. It is appropriate to mention that oestrogen receptor co-regulators may also be recruited by other nuclear receptors, and thus the effect of inactivation of these co-regulators may not only reflect the role of E2 signaling, but rather it may be more widespread ¹⁸.

Treatment options

With recent advancement in technology and methods, there are numerous treatment options for male infertility. Depending on the cause of infertility, treatments may include:

Medication: Hormone therapy to increase the number of sperm production.

Lifestyle changes; Maintaining a healthy body weight. Quit smoking, stop drinking, avoid using marijuana²¹.

Surgery: Vasectomy reversal; This is a common outpatient surgical procedure. The surgeon reconnects the vas deferens which is the tube in the scrotum through which the sperm passes. Viewing the vas deferens through a high-power surgical microscope, the surgeon will carefully sew the ends back together²².

Vasoepididymostomy: Blockages in the vas deferens are repaired with a similar technique of vasectomy reversal. The vas deferens is surgically split, the blockage is removed and the ends of the tube are reconnected²¹.

Sperm retrieval: in some severe cases, a biopsy of the testicle is required

Intracytoplasmic sperm injection: Artificial techniques of reproduction have advanced to the point where a single sperm can be physically injected into an egg. This procedure, called intracytoplasmic sperm injection (ICSI) has dramatically changed the treatment available for even the most severe male factor infertility. Because of this technique, 90% of all infertile males have the potential to conceive their own genetic child²³.

Invitro fertilization; Couples dealing with male infertility, invitro fertilization (IVF) is the treatment of choice. During the IVF process, the ovaries are stimulated with injectable fertility medications to cause multiple eggs to mature. When the eggs are ready, they are collected in a minor procedure²³. Fertilization is accomplished by exposing the eggs to sperm in a culture dish, or by directly injecting a single sperm into each mature egg, a process called intracytoplasmic sperm injection. After fertilization, embryo development is monitored over the next three to five days, and two to three embryos are then placed into the uterus by way of a small catheter inserted through the cervix²².

Conclusion

Estrogen beta receptors are strongly expressed in the testis and in the epididymis. In human testis, ER β have also been found in ejaculated spermatozoa which plays significant role in the motility, longevity and fertilizing capacity of the sperm. It has been shown that, the absence of ER β leads to decreased epididymal sperm content, decreased sperm motility and fertilizing capacity which in turn leads to male infertility.

Recommendations

1. In the diagnosis of male infertility, it is recommended that genetic evaluation should be carried out for assessing the presence and level of $ER\beta$ receptors.

- 2. Antioxidant's vitamins and mineral should be included as part of treatment regimen in order to prevent oxidative DNA damage which may be involved in oestrogen induced effects on male reproduction.
- 3. Healthy lifestyle is recommended. Individuals

should avoid tobacco smoking, marijuana and alcohol use as it damages sperm DNA.

Conflict of Interest: The authors have no conflict of interest to declare.

References

- Adewoyin M, Ibrahim M, Roszaman R, et al. Male Infertility: The Effect of Natural Antioxidants and Phytocompounds on Seminal Oxidative Stress. *Diseases*. 2017;5(1):9. doi:10.3390/diseases5010009
- Sikka, S.C.; Hellstrom, W.J.G.; Naz RKP. Pentoxifylline: Role in management of male infertility/mechanisms of action. *Mol Androl.* 2017;5(5):220-231.
- Cao XW, Lin K, Li CY, Yuan CW. [A review of WHO Laboratory Manual for the Examination and Processing of Human Semen (5th edition)]. Zhonghua Nan Ke Xue. 2017;17(12):1059-1063.
- Ahmed A, Bello A, Mbibu N, Maitama H, Kalayi G. Epidemiological and aetiological factors of male infertility in Northern Nigeria. *Niger J Clin Pract.* 2010;13(2):205-209.
- Abarikwu SO. Causes and Risk Factors for Male-Factor Infertility in Nigeria : A Review. Afr J Reprod Health. 2018;17(4):150-166.
- Uadia PO, Emokpae AM. Male infertility in Nigeria: A neglected reproductive health issue requiring attention. J Basic Clin Reprod Sci. 2015;4(2):45-53.
- Liaqat S, Hasnain S, Muzammil S, Hayat S. Polymorphism analysis in estrogen receptors alpha and beta genes and their association with infertile population in Pakistan. *EXCLI J.* 2015;14:1085-1094.

doi:10.17179/excli2015-559

- Younes AH, Hamed HB, Mohamed EM, Makki MAE, Gaber N, Mohamed HM. Oestrogen receptors beta genotype in infertile Egyptian men with nonobstructive azoospermia. *First Int J Androl.* 2016;1:1-5. doi:10.1111/and.12575
- Ferlin A, Raicu F, Gatta V, Zuccarello D, Palka G, Foresta C. Male infertility: Role of genetic background. Reprod Biomed Online. 2017;14(6):734-745. doi:10.1016/S1472-6483(10)60677-3
- Li TF, Wu QY, Zhang C, et al. Polymorphisms in estrogen receptors predict the risk of male infertility: A meta-analysis. Reprod Biol Endocrinol. 2014;12(1). doi:10.1186/1477-7827-12-79
- 11. Safarinejad MR, Shafiei N, Safarinejad S. Association of polymorphisms in the estrogen receptors alpha, and beta (ESR1, ESR2) with the occurrence of male infertility and semen parameters. J Steroid Biochem Mol Biol. 2018;122(4):193-203.

doi:10.1016/j.jsbmb.2010.06.011

12. Makker K, Agarwal A, Sharma R. Oxidative

stress & male infertility. Indian J Med Resaerch. 2014;12(9):357-367.

- Hess. RA. No Title Estrogen in the adult male reproductive tract: A review. Reprod Biol Endocrinol. 2019;1:52.
- Bordin BM, Moura KKVO. Association between Rsal polymorphism in estrogen receptor β gene and male infertility. Genet Mol Res. 2015;14(3):10954-10960. doi:10.4238/2015.September.21.7
- Leavy M, Trottmann M, Liedl B, et al. Effects of Elevated β-Estradiol Levels on the Functional Morphology of the Testis - New Insights. Sci Rep. 2017;7(2):1-11. doi:10.1038/srep39931
- 16. 16. Guido C, Perrotta I, Panza S, et al. Human Sperm Physiology: Estrogen Receptor Alpha (Erα) and Estrogen Receptor Beta (Erβ) Influence Sperm Metabolism and May Be Involved in the Pathophysiology Of Varicocele-Associated Male Infertility. J Cell Physiol. 2011;226(12):3403-3412. doi:10.1002/jcp.22703
- Dumasia K, Kumar A, Deshpande S, Balasinor NH. Estrogen, through estrogen receptor 1, regulates histone modifications and chromatin remodeling during spermatogenesis in adult rats. *Epigenetics*. 2017;12(11):953-963. doi:10.1080/15592294.2017.1382786
- Dostalova P, Zatecka E, Dvorakova-Hortova K. Of oestrogens and sperm: A review of the roles of oestrogens and oestrogen receptors in male reproduction. Int J Mol Sci. 2017;18(5):55-59. doi:10.3390/ijms18050904
- Couse JF, Mahato D, Eddy EM, Korach KS. Molecular mechanism of estrogen action in the male: Insights from the estrogen receptor null mice. Reprod Fertil Dev. 2021;13(4):211-219. doi:10.1071/RD00128
- Aschim EL, Giwercman A, Ståhl O, et al. The Rsal polymorphism in the estrogen receptor-β gene is associated with male infertility. J Clin Endocrinol Metab. 2005;90(9):5343-5348. doi:10.1210/jc.2005-0263
- Okonofua F, Ivanov I. A case-control study of risk factors for male infertility in Nigeria. Published online 2015. doi:10.1111/j.1745-7262.2005.00046.x
- 22. Olooto W. Infertility in Male; Risk Factors, Causes and Management- A Review. J Microbiol Biotechnol Res. 2012;2(4):641-645.
- 23. Izabel A, Balbin S, Netherton J, Baker MA. From Past to Present: The Link Between Reactive Oxygen Species in Sperm and Male Infertility. J Antioxidants. 2019;8(16):1-19.