



REVIEW ARTICLE

# Challenges and Opportunities in Systemic Biomarkers for Age-related Macular Degeneration

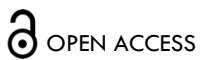
Rogil Jose de Almeida Torres <sup>1</sup>, Andrea Luchini <sup>2</sup>, Mariana Cozer Siviero <sup>3</sup>, Mebaliah Luchini de Almeida Torres <sup>4</sup>, Ana Lucia Anjos Ferreira <sup>1</sup>

<sup>1</sup> Universidade Estadual Paulista (UNESP), Medical School, Botucatu 18618-687, SP, Brazil.

<sup>2</sup> Centro Oftalmológico de Curitiba, Curitiba, PR, Brazil

<sup>3</sup> Pontifícia Universidade Católica do Paraná (PUC-PR), Curitiba, Brasil

<sup>4</sup> University South Florida, Tampa, Florida, USA



OPEN ACCESS

## PUBLISHED

31 December 2024

## CITATION

Torres, RJA., Luchini, A., et al., 2024. Challenges and Opportunities in Systemic Biomarkers for Age-related Macular Degeneration. Medical Research Archives, [online] 12(12). <https://doi.org/10.18103/mra.v12i12.6129>

## COPYRIGHT

© 2024 European Society of Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## DOI

<https://doi.org/10.18103/mra.v12i12.6129>

## ISSN

2375-1924

## ABSTRACT

Blood biomarkers have been widely used in medicine for the prevention, diagnosis, and treatment follow-up of many diseases. Age-related macular degeneration, the main cause of irreversible blindness in old age, does not benefit from these important indicators. In recent decades, technological advances have given us optical coherence tomography. Until a few years ago, this exam was used for the diagnosis, and treatment evaluation of age-related macular degeneration. Currently, as part of multimodal fundus imaging, it has also been used in the prognosis of age-related macular degeneration. However, these exams are still not capable of predicting when individuals may trigger the degenerative macular disease, and consequently adopt preventive measures, such as changes in lifestyle and consumption of antioxidants. In this regard, this article aims to address the various blood biomarkers that may be useful in the early investigation of age-related macular degeneration, even before the appearance of drusen and retinal pigment epithelium changes in the macular region, the first ophthalmoscopic manifestations of age-related macular degeneration. Among these biomarkers analyzed, the blood count, lipid profile, some enzymatic and non-enzymatic antioxidants, as well as the main inflammatory biomarkers, stand out.

## Introduction

Studies have correlated age-related macular degeneration (AMD), the main cause of irreversible blindness in old age<sup>1</sup>, with an increase in the mortality rate in patients with Covid-19 and acquired immunodeficiency syndrome<sup>2-5</sup>. Diseases such as stroke and cardiovascular diseases (CVDs) also have a worse prognosis in patients with AMD<sup>6-7</sup>. The poor evolution of these systemic diseases may be associated to the increase in systemic inflammatory markers detected in patients with AMD<sup>2-5</sup>. It is known that systemic inflammation is associated with aging and an increased risk of many chronic diseases<sup>8</sup>, contributing to the onset and exacerbation of diseases such as obesity, type 2 diabetes, and atherosclerosis<sup>9-11</sup>. As well as systemic diseases, the high systemic levels of cytokines and chemokines may also be involved in the triggering and progression of AMD<sup>12-24</sup>. Consequently, the detection of an increase in these inflammatory markers may allow more effective preventive measures, nullifying or attenuating the progression not only of systemic diseases, but also of AMD<sup>25</sup>.

In 2014, Klein and colleagues published the relationships among serum markers of inflammation, oxidative stress and endothelial dysfunction, with the cumulative incidence of early AMD over 20 years. Twenty-three percent of the 975 people who participated in the initial examination in 1988-1990 developed early AMD. This fact was verified in the 1993-1995, 1998-2000, 2003-2005 and 2008-2010 follow-ups. The study revealed modest evidence of relationships between incidence of early AMD and high-sensitivity serum C-reactive protein, tumor necrosis factor- $\alpha$  receptor 2, interleukin-6, and soluble vascular cell adhesion molecule-1<sup>26</sup>. However, recent studies have been suggested that systemic inflammation may contribute to an increased risk of AMD, even knowing that AMD is a local disease with links to local inflammatory events<sup>24,27</sup>.

Genetic, nutritional, environmental and cardiovascular factors, among others, are involved in the genesis of AMD<sup>28-29</sup>, frequently making it difficult to control. Neovascular age-related macular degeneration (nAMD), one of late forms of AMD, is currently treated with intraocular antiangiogenic injections, which are uncomfortable, periodic, continuous and expensive<sup>30-33</sup>. Furthermore, it does not prevent, in most cases, the progression of the disease and consequent loss of central vision<sup>34</sup>. Additionally, about a third of patients do not obtain the expected effects from anti-vascular endothelial growth factor (VEGF) therapy due to fibrosis or macular atrophy, which makes this disease a poor prognosis<sup>35</sup>. For geographic atrophy (GA), another late form of AMD, Food and Drug Administration (FDA) approved Pegcetacoplan intravitreal injection (complement C3-cleavage inhibitor). It was observed that monthly intravitreal injections of 15 mg Pegcetacoplan for 12 months significantly reduced the growth of lesions in GA<sup>36-37</sup>. In addition to not promoting improved vision, the treatment is also periodic, expensive and not completely free from complications<sup>36</sup>. In order to mitigate this bad evolution, a preventive approach must be initiated as early as possible.

Currently, an evaluation with retinography, fluorescein angiography, indocyanine green and optical coherence tomography (OCT), alone or combined as in multimodal ocular evaluation (multimodal fundus imaging), are capable to identifying the onset and progression of AMD, providing a more appropriate classification, detecting its activity and serving as a guide to evaluate the treatment effectiveness. Additionally, important AMD biomarkers such as macular pigment optical density (MPOD), drusen volume and pigmentary abnormalities can be measured by those exams<sup>38-39</sup>. Other risk factors for disease progression to advanced stages, such as reticular pseudodrusen, hyperreflective foci and drusen subphenotypes, can be identified by combining those tests<sup>40-41</sup>. Other structural biomarkers such as the reflectivity of the ellipsoid zone and the characteristics of the choriocapillaris flow, analyzed by multimodal fundus imaging, can also contribute to a better understanding of AMD pathogenesis and prognosis<sup>42</sup>.

However, as it is a chronic disease, often with decades of evolution, the success of preventive measures in AMD, with the use of antioxidants and lifestyle changes, are not very significant, as they are introduced after the appearance of the initial changes of the disease, such as drusen or pigmentary changes of the retinal pigment epithelium (RPE) in the foveolar region<sup>43-44</sup>. It is worth noting that these initial changes occur in elderly people, often presenting systemic inflammatory components (inflammaging)<sup>45</sup>, changes in serum cholesterol levels<sup>46</sup>, smokers<sup>47</sup>, unhealthy diets<sup>48</sup>, obesity<sup>49</sup>, sedentary life<sup>50</sup>, cardiovascular diseases<sup>51</sup>, among other comorbidities<sup>29,52</sup>. As it is a tissue with a very high metabolism, the retina is drastically impacted by such chronic changes, frustrating, in most cases, any preventive and/or treatment measures<sup>53-54</sup>. Therefore, the objective of this review is to define the main blood biomarkers that have the potential to suggest the adoption of preventive measures, even before the appearance of initial changes in AMD.

## Blood Biomarkers

### BLOOD COUNT

Considering that inflammatory and immunological factors participate in AMD pathogenesis<sup>25,55-59</sup>, changes in the blood count may indicate the beginning of systemic imbalance that may induce degenerative macular disease. Even though some studies showed conflicting results regarding white blood cells (monocytes and neutrophils), platelets and mean platelet volume (MPV)<sup>60-62</sup>, the increase in the neutrophil-lymphocyte ratio (NLR) associated with neovascular AMD has been demonstrated<sup>63</sup>. Corroborating this finding, another study demonstrated that in addition to the increase in NLR, there was also a significant increase in the platelet/lymphocyte ratio (PLR) in patients with neovascular AMD. This study also demonstrated that NLR and PLR levels were inversely proportional to best corrected visual acuity and directly proportional to central macular thickness<sup>64</sup>. Finally, a cross-sectional study with 7,719 participants detected the association of a higher peripheral monocyte count with a higher prevalence of AMD. Individuals with a monocyte count  $\geq 0.5 \times 10^9/L$  compared to participants with monocytes of  $0.1-0.4 \times 10^9/L$  had a 1.45-fold increased risk of

any AMD (early, intermediate, and late) and 1.58-fold increase in the risk of intermediate/late AMD<sup>65</sup>.

#### LIPID PROFILE

The lipids are organic compounds with essential functions for the human body which include energy storage, membrane integration and structure (phospholipid bilayer), in the biosynthesis process of substances such as prostaglandins, and as an enzyme cofactor<sup>66,67</sup>. The lipids represent the main target for the reactive oxygen species (ROS), especially the glycolipids, phospholipids, and cholesterol<sup>68</sup>. Cross-sectional and cohort studies have not reported significant associations between serum lipoprotein profiles and AMD<sup>69,70</sup>. Although one study associated higher serum levels of total cholesterol (TC), low-density lipoprotein (LDL), and triglycerides (TG) with a decreased risk of AMD<sup>71</sup>, another study demonstrated that male participants who had levels higher high-density lipoprotein (HDL) and lower TG levels were associated with greater odds of having early AMD, whereas female participants who had higher TC and LDL levels were associated with an increased risk of developing early AMD<sup>72</sup>. Another study demonstrated that elevated TC in early middle age may play a role in the early development of AMD<sup>73</sup>. In addition, it was found that alterations in serum lipid profiles, as a reflection of systemic dyslipidemia, have been associated with AMD<sup>74</sup>. Corroborating the association between serum lipoprotein levels and AMD, a systematic review and meta-analysis as well as other population studies identified a significantly increased risk of AMD associated with higher serum HDL levels<sup>46,71,75-80</sup>.

#### ANTIOXIDANT ENZYMATIC AND NON-ENZYMATIC SYSTEM

The variation in the activity of antioxidant enzymatic system suggests a response to oxidative stress which is involved in the pathogenesis of AMD<sup>81-82</sup>. The enzymatic system have the function of counterbalancing the damage caused by ROS and reactive nitrogen species (RNS) in the biomolecules of lipids, proteins, and DNA contained in the sensory retina and in the RPE, maintaining the retinal homeostasis and attenuating the installation and AMD progression<sup>83-85</sup>.

#### **Glutathione Redox cycle and enzymes**

Glutathione, or L-γ-glutamyl-L-cysteinyl-glycine, is ubiquitous tripeptide that participates in biological processes such as protein and DNA synthesis, transport, enzymatic activity, metabolism and cellular protection. Glutathione is synthesized intra-cellularly and can be found in the body in its reduced (GSH) and oxidized (GSSG) forms<sup>86</sup>. Free GSH is present mainly in its reduced form, being considered a potent ROS scavenger. Glutathione may be converted into the GSSG by the glutathione peroxidase (GPx) during oxidative stress. The oxidized form (GSSG), in turn, may be reverted into its reduced form by glutathione reductase (GR)<sup>87</sup>. While the concentration of GSH in cells varies from 1 to 10 mM<sup>88</sup>, the level of GSH within extracellular fluids and blood plasma reaches only several μM<sup>89</sup>. It is important to point out that a high level of GSH can be explained by an enhanced GSH biosynthesis and a higher conversion of GSSG into GSH by GR. On the other hand, under conditions of marked toxicity or oxidative stress, intracellular GSSG increases substantially<sup>87</sup>.

#### **Glutathione**

Glutathione, the major water-soluble antioxidant, acts primarily in the cytoplasm and mitochondria and is considered the most important antioxidant in the eye<sup>86,90</sup>. With age, the efficiency of the GSH redox system decreases, predisposing the RPE to increased damage mediated by oxidative stress<sup>90-93</sup>. Some studies have not demonstrated changes in GSH concentrations in the plasma of patients with AMD<sup>81,94</sup>. On the other hand, a study demonstrated higher serum GSH concentrations in AMD patients in relation to control group<sup>95</sup>, while most studies showed decreased GSH levels in AMD patients as normal control<sup>96-99</sup>. Consequently, GSH has been considered a non-specific parameter when analyzing the development of AMD<sup>90</sup>.

#### **Glutathione peroxidase**

Glutathione peroxidase controls hydrogen peroxide and lipid hydroperoxide levels, resulting from an attack of ROS<sup>86</sup>. This enzyme is present in a number of tissues, including the inner layers of the retina and RPE<sup>100-102</sup>. Its concentration is greatest in the posterior pole, which is constantly exposed to light<sup>101</sup>. Selenium (Se), an essential nutrient, is part of the composition of GPx<sup>100</sup>, and Se-deficient animals have markedly decreased GPx activity<sup>103</sup>. In addition to GPx protecting RPE cells in models of oxidative damage-induced retinal degeneration, it plays an important role in the maturation of photoreceptor cells<sup>104</sup>. A population-based, cross-sectional study on cataracts and AMD and their risk factors (2584 participants) showed that the high plasma level of GPx plasma was associated with a nine-fold increase in the prevalence of late AMD<sup>105</sup>. An increase in GPx activity in AMD patients has been associated with the activity of RPE cells, which attempt to eliminate a huge amount of H<sub>2</sub>O<sub>2</sub> produced during the course of the disease<sup>83</sup>. On the other hand, many studies have reported a significant reduction in GPx activity in patients with AMD<sup>106-110</sup>.

#### **Oxidized glutathione**

Oxidative stress promotes the conversion of GSH to GSSG by GPx enzyme<sup>87</sup>. A report showed that GSSG level is elevated in patients with early AMD as compared to those of healthy control<sup>111</sup>.

#### **Glutathione reductase**

Although glutathione reductase is not directly an antioxidant, its function is essential for the maintenance of available reduced GSH<sup>87</sup>. This enzyme is highly expressed in the retina and RPE<sup>112-114</sup>. Several studies have demonstrated significantly lower GR activity values in the group of AMD patients as compared those of control group; being associated with a decreasing in GSH levels<sup>85,107,115</sup>. In fact, lower GR activity is associated with weaker antioxidant abilities<sup>85</sup>.

#### **Superoxide dismutase (SOD)**

Superoxide dismutase plays a key defense role against ROS. By removing superoxide (O<sub>2</sub><sup>-</sup>) and forming O<sub>2</sub> and H<sub>2</sub>O<sup>116</sup>, it plays an important role in diseases related to oxidative stress, including aging<sup>117</sup>. In eukaryotic systems, one can find SOD linked to copper and zinc (CuZn)-SOD, present mainly in the cytosol, as well as SOD-2, dependent on manganese (Mn-SOD), and found mainly in mitochondria<sup>118</sup>. Mn-SOD is present in RPE cells and the

inner rod segment<sup>119</sup>, and reduced levels are associated with AMD progression<sup>120</sup>. Some studies demonstrated that high erythrocyte SOD activity was not associated with late AMD and early signs of AMD<sup>82,105,108,121</sup>. This finding has been attributed to compensatory mechanisms against oxidative stress<sup>122</sup>. However, other studies report lower values of SOD antioxidant parameters were significantly associated with AMD<sup>85,109,110,115</sup>. Although *in vitro* studies consistently indicate the role of SOD in responses to oxidative stress, they do not clearly show its association with AMD<sup>83</sup>.

### Catalase

Catalase, a homotetrameric protein, is present in many types of cells, with the highest concentration in erythrocytes and liver<sup>123</sup>, and is found mainly in peroxisomes, mitochondria and the nucleus. This enzyme decomposes H<sub>2</sub>O<sub>2</sub> into water and molecular oxygen, an extremely important process to prevent the formation of the hydroxyl radical ( $\bullet$ OH)<sup>124</sup>. In the retina, catalase is located within RPE peroxisomes, playing its important role in preventing lipid peroxidation and inhibiting lysosomal enzymes through the removal of H<sub>2</sub>O<sub>2</sub> from the phagosome<sup>125,126</sup>. Decreased catalase activity in RPE cells has been reported from the sixth to ninth decade of human life<sup>92</sup>. Corroborating this study, a decrease in catalase immunoreactivity with age was observed in the cytoplasm and lysosomes of macular RPE cells from normal eyes and eyes affected by AMD<sup>127</sup>. Although a study (32 early-AMD patients, 25 late-AMD patients, 50 healthy controls) did not establish a correlation between serum catalase activity and AMD<sup>108</sup>, another study (39 early-AMD patients, 100 intermediate-AMD patients, 191 late-AMD patients, and 121 controls) revealed that AMD is associated with lower erythrocyte catalase activity<sup>82</sup>. Other study (240 AMD patients and 270 controls) also reported a reduction in erythrocyte catalase activity in patients with AMD<sup>110</sup>.

### Total Oxidant Status (TOS) and Total Antioxidant Status (TAS)

Total oxidant status and total antioxidant status are parameters used to evaluate the overall oxidative stress status in the body<sup>128</sup>. An imbalance between TOS and TAS has been proposed to be responsible for the increased lipid, protein and DNA damage observed in AMD patients<sup>129</sup>. Serum (or plasma) concentrations of different oxidant species can be measured in laboratories separately, but the measurements are time-consuming, labor-intensive and costly and require complicated techniques<sup>130</sup>.

Total oxidant status may also be named total peroxide (TP), serum oxidation activity, reactive oxygen metabolites or some other synonyms<sup>131</sup>. A study (156 early-AMD patients, 80 wet-late AMD, 72 dry-late AMD and 207 healthy controls) reported that a significantly increased oxidative damage was associated with AMD patients. Both early- and late-AMD patients presented higher TOS levels than healthy controls<sup>109</sup>. Corroborating these findings, other studies also observed a significant increase in TOS levels in the sera of AMD patients when compared to controls<sup>109,129,132-133</sup>.

Total antioxidant status, in turn, expresses the free radical scavenging capacity and reflects the residual

antioxidant capacity after ROS neutralization<sup>134-135</sup>. Total antioxidant status was shown to be reduced in patients with AMD compared to controls<sup>85,115,129</sup>. Another study (32 early-AMD patients, 25 late-AMD patients and 50 healthy subjects) demonstrated that low TAS is associated with AMD and that the combined values of GPx activity and TAS are significant determinants of AMD status<sup>108</sup>.

### Protein carbonyl (PC)

Protein carbonyls are indicators of the amount of protein that has been oxidized by highly reactive free radicals and are the most studied protein oxidation markers<sup>135,136</sup>. Reactive carbonyl species are important cytotoxic mediators produced from oxidative damage of biomolecules (lipids and sugars), leading to alterations in the cell signaling mechanisms to the nucleus, positively regulating redox-sensitive transcription factors, and inducing irreversible structural modification in important molecules [proteins, peptides (cysteine, lysine, histidine), lipids, DNA]<sup>137,138</sup>. Studies have demonstrated that the values of protein carbonyl groups were higher in patients with exudative AMD than in the control group<sup>85,129</sup>. In another study, both patients with late AMD and those with early AMD had higher levels of PC when compared to healthy controls<sup>109</sup>.

## INFLAMMATORY BIOMARKERS

### Pro-inflammatory cytokines and chemokines

Several studies have demonstrated inflammation and dysregulation of inflammatory responses play an important role in the development and progression of AMD to the final stages, which include choroidal neovascularization (CNV) and geographic atrophy (GA)<sup>27,139, 140</sup>. Systemically, studies have demonstrated elevated plasma levels of these inflammatory markers in patients with AMD as compared to those without AMD<sup>13-14, 16-22,141-142</sup>. Considering these facts, inflammatory cytokines and chemokines gain relevance from a pathogenesis and therapeutic perspective in AMD.

Cytokines are water-soluble, extracellular polypeptides or glycoproteins, ranging from 8 to 30 kDa, generally produced in response to antigenic stimulation, functioning as a chemical messenger to regulate the adaptive and innate immune system. These proteins are produced by all cells involved in antigen response and presentation. They are synthesized when needed or when a cell in the immune system is "activated"<sup>143-144</sup>. Chemokines, in turn, are a large family of small cytokines and their molecular weight varies from 7 to 15kDa. Chemokines play a central role in the physiology of leukocytes and other inflammatory cells, by controlling basal and inflammatory trafficking. There are two major subfamilies of chemokines based on the position of cysteine residues: CXC and CC. As a general rule, members of the CXC chemokine family are chemotactic of neutrophils, and CC chemokines are chemotactic of monocytes and lymphocyte subtypes<sup>144</sup>.

### Interleukin-6 (IL-6)

Interleukin-6 is a cytokine that, among its multiple functions, mediates inflammation and the immune response, acting on several cells, including RPE cells<sup>145-147</sup>. IL-6 levels in adults are not expected to exceed 20 pg/mL<sup>148</sup>. Several studies have reported that IL-6 is an

important regulator of CNV and has correlated this cytokine with VEGF expression<sup>149-152</sup>. While one study found no significant association between plasma IL-6 levels and AMD<sup>26</sup>, another study considered this interleukin an important marker for the progression of AMD<sup>16</sup>. Recently, a prospective study (42 GA patients, 41 nAMD patients, and 27 healthy controls) demonstrated that plasma IL-6 has predictive capacity for progression and constitutes the first known plasma biomarker of disease activity in GA. These findings highlight its important role of chronic inflammation in the pathogenesis of this disease<sup>17</sup>. Finally, a meta-analysis involving 3,586 individuals (1,865 controls and 1,721 with AMD) suggests that an increase in systemic IL-6 in patients with AMD may be a phenomenon more closely related to late AMD subtypes<sup>153</sup>.

### **Interleukin-1β (IL-1β)**

Interleukin-1β, produced primarily by monocytes and macrophages, has been associated with mediating acute and chronic inflammation<sup>154-155</sup>. Interleukin-1β is secreted as an inactive form and requires proteolytic cleavage by the enzyme caspase-1 to be released into an active form<sup>156</sup>. The caspase-1 activation platform, known as the inflammasome, has been associated with the pathophysiology of AMD<sup>157-158</sup>. Additionally, IL-1β is capable of inducing reactive oxygen species (ROS) in RPE cells<sup>159</sup>. Analysis of plasma inflammation markers in patients with polypoidal choroidal vasculopathy (PCV), patients with neovascular AMD and a healthy control group showed a significant increase in plasma IL-1β in patients with neovascular AMD as compared to those of healthy controls<sup>17</sup>. Furthermore, this chemokine was found to be elevated in patients who progressed from the intermediate to the advanced stage of AMD<sup>24</sup>.

### **C-reactive protein (CRP)**

C-reactive protein is predominantly produced in the liver, although, under certain conditions, it can also be secreted by smooth muscle cells and endothelial cells<sup>160,161</sup>. The Centers for Disease Control and Prevention and the American Heart Association have estimated cardiovascular risk in healthy individuals as follows: low-, medium-, and high-risk values defined as < 10, 10 – 30, and > 30 mg/L<sup>162</sup>.

CRP is released into circulation upon stimulation by IL-6 and other cytokines<sup>163</sup>, and a link with AMD has been suggested<sup>16,115, 164-167</sup>. However, a meta-analysis involving 53 studies with 60,598 participants (10,392 patients and 38,901 controls) revealed that early age-related macular degeneration was not associated with systemic C-reactive protein, whereas late AMD was associated with a small-to-moderate increase in systemic C-reactive protein<sup>168</sup>. On the other hand, another meta-analysis (41,690 participants) showed that high serum CRP levels (> 3 mg/L) were associated with a twofold greater likelihood of late AMD, compared with low levels (< 1 mg/L)<sup>169</sup>. Furthermore, evidence that elevated high-sensitivity C-reactive protein (hsCRP) levels predict future AMD risk was demonstrated in pooled analysis of prospective case-control data<sup>20</sup>. The association of CRP with AMD was also found in a study that analyzed inflammation plasma markers from patients with PCV and neovascular AMD<sup>19</sup>, as well as in GA<sup>17</sup>.

### **Tumor Necrosis Factor Alpha (TNF-α)**

Tumor necrosis factor alpha is a low molecular weight protein, produced predominantly by activated macrophages, potentially involved in the production and expression of VEGF<sup>170-172</sup>. Interestingly this cytokine also has the potential to inhibit the formation of neovessels<sup>173-174</sup>. This fact may be linked to its receptors, a member of the tumor necrosis factor receptor 1A (Tnfrsf1a) superfamily and a member of the tumor necrosis factor receptor 1B (Tnfrsf1b) superfamily, which act antagonistically by inhibiting endothelial migration or promoting its activation<sup>173-175</sup>. Additionally, it was observed that pre-exposure of TNFα in the primary RPE and ARPE19 induces to increased complement activation and membrane attack complex (MAC) deposition, which may represent an early event in the pathogenesis that leads to the development of AMD<sup>176</sup>. A case-control study showed significantly increased plasma levels of soluble receptor for tumor necrosis factor type II (sTNFRII) in patients with early or nAMD and was considered a significant predictor for the prevalence of AMD<sup>177</sup>. Another study that analyzed serum levels of CRP, pro-inflammatory cytokines (TNF-α, IL-1β, IL-2, IL-6 and IL-8) and complement pathway activity in the clinical response to growth factor inhibition vascular endothelial cell in nAMD demonstrated that only lower serum levels of TNF-α were associated with an increase in visual acuity after anti-VEGF therapy<sup>178</sup>. In a prospectively study (42 GA patients, 41 nAMD patients and 27 healthy controls) was observed that patients with GA showed an increase in the pro-inflammatory plasma marker (TNF receptor 2) as compared to those of healthy controls<sup>17</sup>. Investigating the role of TNF-α on AMD, a recent systematic review (24 studies) showed measurement of systemic and local levels of TNF-α has not produce consistent results on the role of anti-TNF-α agents in the remission of symptoms caused by the disease. That review suggested the role of TNF-α in nAMD is not clear and not all anti-TNF-α agents were considered safe<sup>179</sup>. Corroborating this study, no association was demonstrated between plasma levels of TNF-R2 among patients with PCV, patients with nAMD and a healthy control group<sup>19</sup>.

### **Interleukin-8 (IL-8)**

Interleukin-8 or CXCL8 is a member of the CXC chemokine family originally identified as a chemotactic factor for neutrophils, being released by phagocytes and a wide variety of tissue cells after exposure to inflammatory stimuli<sup>180-181</sup>. In addition to activating neutrophils, IL-8 also increases the expression of adhesion molecules by endothelial cells<sup>182</sup>. Some studies have not demonstrated a correlation between plasma IL-8 levels and nAMD and/or PVC<sup>19,183</sup>. On the other hand, a significant increase in the IL-8 secretion profile of peripheral blood mononuclear cells and serum from patients with nAMD was demonstrated<sup>181,184</sup>. Additionally, this chemokine was found to be elevated in patients who progressed from the intermediate to the advanced stage of AMD<sup>24</sup>. Finally, a recent meta-analysis suggested that the IL-8 +781 C/T polymorphism affects the predisposition to dry AMD and wet AMD. Furthermore, patients with dryAMD and nAMD also have elevated levels of IL-8<sup>185</sup>, which could be considered a new genomic biomarker of AMD susceptibility<sup>186</sup>.

**Monocytic chemotactic protein 1 (MCP-1) or Chemokine C-C motif ligand 2 and its receptor CCR2**

Monocytic chemotactic protein 1, also referred to as chemokine C-C motif ligand 2 (CCL2), is encoded by the CCL2 gene which is located on chromosome 17q11.2<sup>187</sup>. CCL2 is a low molecular weight (810 kDa) chemotactic cytokine that is involved in the recruitment of monocytes to the inflammation site<sup>188-190</sup>. Chemokine C-C motif ligand 2 can be secreted by numerous cell types, including endothelial cells, activated monocytes, and EPR cells<sup>191-193</sup>. Several studies have correlated CCL2 with AMD. Firstly, oxidized lipids in the retina, arising from phagocytosis of the outer segments of photoreceptor cells, stimulate the expression of chemotactic factors, such as IL-8 and CCL2 by RPE cells, justifying the increase in intraocular CCL2 concentrations in nAMD eyes<sup>194</sup>. Consequently, the recruitment of macrophages promoted by CCL2 would help explain the presence of these cells at sites of CNV<sup>195-198</sup>. Additionally, macrophages express proangiogenic VEGF, which contributes to CNV formation<sup>199-200</sup>. It was demonstrated that intravitreal injections with a CCR2 antagonist reduce the size of laser-induced CNV in mice, as well as in the number of macrophages infiltrating the choroid and a decrease in VEGF expression<sup>201</sup>. However, CCL2 is not only involved in the pathogenesis of nAMD. Experimental models have shown that CCR2 and its ligand CCL2 are involved in drusen formation and in RPE changes seen in the early stages of AMD<sup>202-203</sup>. Chemokine C-C motif ligand 2 activated monocytes are also involved with apoptosis in the RPE, contributing to the progression of the disease<sup>204</sup>. Furthermore, the level of CCL2 and CCR2 + inflammatory infiltrating monocytes are increased in patients with GA<sup>205</sup>. However, controversial results have been found regarding serum CCL2 levels in patients with AMD. A study involving 150 participants found no association between CCL2 plasma levels and AMD<sup>206</sup>. A cross-sectional study (16 nAMD patients, 18 PCV patients, and 50 controls) did not observe significant differences in CCL2 in plasma samples between cases and controls, corroborating the previous study<sup>183</sup>. On the other hand, analysis of peripheral blood mononuclear cells, in particular monocytes from patients with nAMD, revealed that higher levels of CCL2 may be associated with the development of CNV<sup>181</sup>. Corroborating with these findings, an increase in serum CCL2 levels in patients with nAMD as compared to those of healthy participants has been observed<sup>207</sup>. Another study demonstrated that plasma CCL2 concentrations are not only elevated in nAMD, but also in PCV<sup>208</sup>. An increase in CCL2 plasma concentrations was also observed in patients with intermediate age-related macular degeneration (iAMD)<sup>209</sup>, and early AMD<sup>210</sup>. Similarly, the expression of CCR-2 levels was increased on a sub-type of monocytes in peripheral blood in patients with neovascular AMD<sup>211</sup>. Additionally, it was observed that patients with nAMD had a significantly increased proportion of non-classical CCR2+ monocytes, whereas PCV type 1 was associated with a significant increase in CCR2+ in all monocyte subsets when compared to PCV type 2<sup>212</sup>. A prospective observational study (41 GA patients, 51 nAMD patients, and 30 healthy control) demonstrated that GA was associated with greater monocytic CCR2 expression compared to nAMD<sup>21</sup>. Finally, a significant increase in urinary CCL2 levels was observed in patients with early AMD and GA as

compared of those controls. This study concluded that analysis of this urinary biomarker could provide a practical tool for detecting early AMD, monitoring progression and evaluating treatment efficacy<sup>22</sup>.

**Interferon (IFN)**

Interferons are a group of glycoproteins synthesized and secreted by almost all mammalian cells upon stimulation by specific antigens<sup>213-214</sup>. They have antiviral, antiproliferative and immunomodulatory properties that play important roles in host defense mechanisms and in the maintenance of homeostasis<sup>213-214</sup>. The three types of IFN (I, II and III) are classified by their receptor specificity and sequence homology. Type I IFNs include IFN- $\alpha$ ,  $\beta$ ,  $\epsilon$ ,  $\kappa$ , and  $\omega$ . Type II IFNs refer to IFN- $\gamma$ , and Type III IFNs include IFN $\lambda$ 1, IFN $\lambda$ 2, IFN $\lambda$ 3<sup>215</sup>. The type I Interferon exerts antiproliferative and antiangiogenic effects, modulating the activity of various immune cells<sup>216-219</sup>. On the other hand, type 2 interferon (IFN- $\gamma$ ) is classically considered a pro-inflammatory factor, but in recent years, several studies have found that IFN- $\gamma$  mediates an immunomodulatory and protective function as well<sup>220-221</sup>. In relation to AMD, IFN- $\gamma$  can exert important effects on RPE cells, positively regulating the expression of complement factor H (CFH), mediating the positive regulation of VEGF, promoting the activation of the RPE, significantly increasing the expression and secretion of IL-6<sup>147, 222-223</sup>. In the primary RPE and ARPE19, in addition to promoting increased complement activation, IFN- $\gamma$  promotes the deposition of the membrane attack complex, which may represent an early event in the development of AMD<sup>176</sup>. Hence, IFN- $\gamma$  induces pro-inflammatory responses by activating pro-inflammatory cytokines and chemokines, thus recruiting immune cells such as macrophages and T cells<sup>224-226</sup>. Despite the important role of IFN- $\gamma$  in the pathogenesis of AMD, several studies do not associate the increase in plasma levels of this cytokine with degenerative macular disease<sup>18,177,183,227</sup>. On the other hand, analysis of whole-blood samples collected from humans (27 nAMD patients, 33 PCV patients and 18 healthy individuals) demonstrated that the levels of IFN- $\gamma$  in the supernatants of cultured peripheral blood mononuclear cells in patients with PCV and nAMD were markedly elevated compared to those of controls. These results suggest that the IFN- $\gamma$ -related inflammatory pathway may be involved in the pathogenesis of PCV and nAMD<sup>228</sup>. Another study corroborated these results, demonstrating significantly high levels of IFN- $\gamma$  expression by CD4+ T cells in patients with AMD<sup>13</sup>.

**Homocysteine (Hcy)**

Homocysteine is an amino acid produced from the demethylation of methionine<sup>229</sup>. Mutations in methylenetetrahydrofolate reductase or cysteine by cystathionine-beta-synthase are related to increased plasma homocysteine (HHcy)<sup>230</sup>. Additionally, deficiency of B complex vitamins, such as folic acid, riboflavin (B2), pyridoxine hydrochloride (B6) and cyanocobalamin (B12) is related to an increase in total Hcy concentration<sup>229, 231-233</sup>. Adult total homocysteine values of 10  $\mu$ mol/L or less are probably safe, whereas values of 11  $\mu$ mol/L or above may warrant intervention<sup>234</sup>. Hyperhomocysteinemia has been linked to the

development of Alzheimer's disease<sup>235</sup>, diabetic retinopathy<sup>236</sup>, as well as an increase in the cardiovascular mortality rate<sup>237-238</sup>. Many experimental studies have suggested an association between HHcy and AMD. Hyperhomocysteinemia caused activation of microglial cells, increased the expression of pro-inflammatory cytokines such as IL-1 $\beta$  and TNF $\alpha$ , as well as downregulated anti-inflammatory cytokines in key cells that constitute the inner outer blood–retinal barriers, human retinal endothelial cells and RPE<sup>239-241</sup>. Blood-retinal barrier dysfunction caused by increased Hcy leads to RPE changes like AMD, including inducing CNV<sup>242-244</sup>. Furthermore, HHcy has been shown to promote the activation of hypoxia-inducible factor (HIF)-1 $\alpha$ <sup>244</sup>, retinal hypoxia<sup>245</sup>, and increase in the expression of VEGF in RPE cells<sup>244</sup>. A meta-analysis indicated weak evidence that increased tHcy may be associated with nAMD; however, this result must be interpreted with caution due to a marked heterogeneity among studies<sup>246</sup>. Another meta-analysis, which evaluated eleven studies (including 1072 cases and 1202 controls), demonstrated that the plasma tHcy level among AMD cases was 2.67  $\mu$ mol/L higher than controls. Studies involving vitamin B12 and folic acid were also analyzed (including 152 cases and 98 controls) and it was found that the level of vitamin B12 among AMD cases was 64.16 pg/mL lower than controls. Subgroup analyzes showed that the folic acid level was 1.66 ng/mL lower in the wet type. Together, the results demonstrated that AMD is associated with elevated tHcy levels and decreased vitamin B12 levels<sup>247</sup>. In this regard, a prospective study demonstrated that an increase in total vitamin B-12 intake was associated with a 24% reduction in the risk of incidence of any AMD in 10 years<sup>248</sup>, just as well as high folate intake is associated with a reduced risk of progression to GA<sup>249</sup>. A randomized, double-blind, placebo-controlled trial including 5442 female health care professionals 40 years or older with CVDs or CVDs

risk factors showed that supplementation with folic acid (2.5mg), B12 (1 mg) and B6 (50 mg) led to a 34% reduced risk of any AMD and a 41% reduced risk of visually significant AMD over a 7.3-year period<sup>250</sup>.

### Conclusion

Several studies point to changes in blood biomarkers in AMD, especially in the late stages of the disease. The increase in the neutrophil-lymphocyte ratio (NLR) and the platelet/lymphocyte ratio (PLR), as well as the higher peripheral monocyte count, suggest the onset of systemic changes that can potentially interfere with the course of AMD. The correlation between AMD and increased HDL, TOS, IL-6, CRP, IL-8, MCP-1 and homocysteine has been observed in many studies, as has a reduction in TAS. Most of the biomarkers analyzed were compared between normal elderly people and elderly people with AMD. There are few long-term studies in the literature analyzing the correlation of these markers with the onset of AMD. Therefore, long-term studies should be encouraged in the search for answers regarding the usefulness of such markers in identifying those people who tend to develop AMD, even before the appearance of any phenotypic changes in the retina, with the aim of predicting the risk, preventing it and monitor progression.

### Conflicts of Interest Statement:

The authors have no conflicts of interest to declare.

### Funding Statement:

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for profit sectors.

### Acknowledgments

Lorraine Luchini de Almeida Torres

## References

1. Wong WL, Su X, Li X, et al. Global prevalence of age-related macular degeneration and disease burden projection for 2020 and 2040: A systematic review and meta-analysis. *Lancet Glob Health*. 2014;2:e106-116
2. Ramlall V, Thangaraj PM, Meydan C, et al. Immune complement and coagulation dysfunction in adverse outcomes of SARS-CoV-2 infection. *Nat Med*. 2020;26:1609-1615.
3. Yang JM, Moon SY, Lee JY, Agalliu D, Yon DK, Lee SW. COVID-19 Morbidity and Severity in Patients With Age-Related Macular Degeneration: A Korean Nationwide Cohort Study. *Am J Ophthalmol*. 2022 Jul;239:159-169.
4. Torres RJ. Insights into COVID-19 in age-related macular degeneration. *Pan Am J Ophthalmol*. 2023;5:18
5. Jabs DA, Van Natta ML, Trang G, et al. Association of age-related macular degeneration with mortality in patients with acquired immunodeficiency syndrome; role of systemic inflammation. *Am J Ophthalmol*. 2019;199:230-237.
6. Gopinath B, Liew G, Burlutsky G, Mitchell P. Age-related macular degeneration and risk of total and cause-specific mortality over 15 years. *Maturitas*. 2016 Feb;84:63-67.
7. Wang J, Xue Y, Thapa S, Wang L, Tang J, Ji K. Relation between Age-Related Macular Degeneration and Cardiovascular Events and Mortality: A Systematic Review and Meta-Analysis. *Biomed Res Int*. 2016;2016:8212063.
8. Furman D, Campisi J, Verdin E, et al. Chronic inflammation in the etiology of disease across the life span. *Nat Med*. 2019;25(12):1822-1832.
9. Li C, Yuan J, Zhu YF, et al. Imbalance of Th17/Treg in Different Subtypes of Autoimmune Thyroid Diseases. *Cell Physiol Biochem*. 2016; 40: 245-252.
10. Sokolove J, Lepus CM: Role of inflammation in the pathogenesis of osteoarthritis: latest findings and interpretations. *Ther Adv Musculoskelet Dis*. 2013; 5: 77-94.
11. Lahoute C, Herbin O, Mallat Z, Tedgui A. Adaptive immunity in atherosclerosis: mechanisms and future therapeutic targets. *Nat Rev Cardiol*. 2011; 8: 348-358.
12. Cheung CMG, Wong TY. Is age-related macular degeneration a manifestation of systemic disease? New prospects for early intervention and treatment. *J Intern Med*. 2014;276(2):140-153.
13. Chen J, Wang W, Li Q. Increased Th1/Th17 responses contribute to low-grade inflammation in age-related macular degeneration. *Cell Physiol Biochem*. 2017;44:357-367.
14. Nassar K, Grisanti S, Elfar E, Lüke J, Lüke M, Grisanti S. Serum cytokines as biomarkers for age-related macular degeneration. *Graefes Arch Clin Exp Ophthalmol*. 2015 May;253(5):699-704.
15. Falk MK, Singh A, Faber C, Nissen MH, Hviid T, Sørensen TL. Dysregulation of CXCR3 expression on peripheral blood leukocytes in patients with neovascular age-related macular degeneration. *Invest Ophthalmol Vis Sci*. 2014 May 8;55(7):4050-6.
16. Seddon JM, George S, Rosner B, Rifai N. Progression of age-related macular degeneration: Prospective assessment of C-reactive protein, interleukin 6, and other cardiovascular biomarkers. *Arch Ophthalmol*. 2005;123:774-782.
17. Krogh Nielsen M, Subhi Y, Molbech CR, Falk MK, Nissen MH, Sørensen TL. Systemic Levels of Interleukin-6 Correlate With Progression Rate of Geographic Atrophy Secondary to Age-Related Macular Degeneration. *Invest Ophthalmol Vis Sci*. 2019 Jan 2;60(1):202-208.
18. Afarid M, Azimi A, Malekzadeh M. Evaluation of serum interferons in patients with age-related macular degeneration. *J Res Med Sci*. 2019;24:24.
19. Subhi Y, Krogh Nielsen M, Molbech CR, et al. Plasma markers of chronic low-grade inflammation in polypoidal choroidal vasculopathy and neovascular age-related macular degeneration. *Acta Ophthalmol*. 2019;97:99-106.
20. Mitta VP, Christen WG, Glynn RJ, et al. C-reactive protein and the incidence of macular degeneration: Pooled analysis of 5 cohorts. *JAMA Ophthalmol*. 2013;131:507-513.
21. Krogh Nielsen M, Subhi Y, Molbech CR, Falk MK, Nissen MH, Sørensen TL. Chemokine Profile and the Alterations in CCR5-CCL5 Axis in Geographic Atrophy Secondary to Age-Related Macular Degeneration. *Invest Ophthalmol Vis Sci*. 2020 Apr 9;61(4):28.
22. Guymer RH, Tao LW, Goh JK, et al. Identification of urinary biomarkers for age-related macular degeneration. *Invest Ophthalmol Vis Sci*. 2011;52:4639-4644.
23. Tuo J, Grob S, Zhang K, Chan C-C: Genetics of Immunological and Inflammatory Components in Age-related Macular Degeneration. *Ocul Immunol Inflamm*. 2012; 20: 27-36.
24. Wagner BD, Patnaik JL, Palestine AG, et al. Association of Systemic Inflammatory Factors with Progression to Advanced Age-related Macular Degeneration. *Ophthalmic Epidemiol*. 2022 Apr;29(2):139-148.
25. Frederick PA, Kleinman ME. The Immune System and AMD. *Curr Ophthalmol Rep*. 2014 Mar 1;2(1):14-19.
26. Klein R, Myers CE, Cruickshanks KJ, et al. Markers of inflammation, oxidative stress, and endothelial dysfunction and the 20-year cumulative incidence of early age-related macular degeneration: the Beaver Dam Eye Study. *JAMA Ophthalmol*. 2014 Apr 1;132(4):446-55.
27. Tan W, Zou J, Yoshida S, Jiang B, Zhou Y. The Role of Inflammation in Age-Related Macular Degeneration. *Int J Biol Sci*. 2020; 16(15):2989-3001.
28. Wong T, Chakravarthy U, Klein R, et al. The Natural History and Prognosis of Neovascular Age-Related Macular Degeneration. *Ophthalmology*. 2008;115: 116-126.
29. Hyttinen JMT, Blasiak J, Kaarniranta K. Non-Coding RNAs Regulating Mitochondrial Functions and the Oxidative Stress Response as Putative Targets against Age-Related Macular Degeneration (AMD). *Int J Mol Sci*. 2023 Jan 30;24(3):2636.



30. Wong TY, Liew G, Mitchell P. Clinical update: new treatments for age-related macular degeneration. *Lancet*. 2007; 370: 204-206.
31. CATT Research Group, Martin DF, Maguire MG, et al. Ranibizumab and Bevacizumab for Neovascular Age-Related Macular Degeneration. *N Engl J Med*. 2011; 364: 1897-1908.
32. Hussain RM, Ciulla TA. Emerging vascular endothelial growth factor antagonists to treat neovascular age-related macular degeneration. *Expert Opin Emerg Drugs*. 2017; 22: 235-246.
33. Yang S, Zhao J, Sun X. Resistance to anti-VEGF therapy in neovascular age-related macular degeneration: a comprehensive review. *Drug Des Devel Ther*. 2016; 10: 1857-1867.
34. Rofagha S, Bhisitkul RB, Boyer DS, Sadda SR, Zhang K; SEVEN-UP Study Group. Seven-year outcomes in ranibizumab-treated patients in ANCHOR, MARINA, and HORIZON: a multicenter cohort study (SEVEN-UP). *Ophthalmology*. 2013 Nov;120(11):2292-9.
35. Little K, Ma JH, Yang N, et al. Myofibroblasts in macular fibrosis secondary to neovascular age-related macular degeneration - the potential sources and molecular cues for their recruitment and activation. *EBioMedicine*. 2018; 38: 283-291.
36. Heier JS, Lad EM, Holz FG, et al. Pegcetacoplan for the treatment of geographic atrophy secondary to age-related macular degeneration (OAKS and DERBY): Two multicentre, randomised, double-masked, sham-controlled, phase 3 trials. *Lancet*. 2023; 402:1434-1448.
37. Liao DS, Grossi FV, El Mehdi D, et al. Complement C3 inhibitor pegcetacoplan for geographic atrophy secondary to age-related macular degeneration: A randomized phase 2 trial. *Ophthalmology*. 2020;127:186-195.
38. Nathoo NA, Or C, Young M, et al. Optical coherence tomography-based measurement of drusen load predicts development of advanced age-related macular degeneration. *Am J Ophthalmol*. 2014;158:757-761.e1.
39. Folgar FA, Yuan EL, Sevilla MB, et al. Drusen volume and retinal pigment epithelium abnormal thinning volume predict 2-year progression of age-related macular degeneration. *Ophthalmology*. 2016;123:139-50.e1.
40. Lad EM, Finger RP, Guymer R. Biomarkers for the Progression of Intermediate Age-Related Macular Degeneration. *Ophthalmol Ther*. 2023 Dec;12(6):2917-2941.
41. Trinh M, Cheung R, Duong A, Nivison-Smith L, Ly A. OCT Prognostic Biomarkers for Progression to Late Age-related Macular Degeneration: A Systematic Review and Meta-analysis. *Ophthalmol Retina*. 2023 Dec 27:S2468-6530(23)00668-1.
42. Damian I, Nicoară SD. SD-OCT Biomarkers and the Current Status of Artificial Intelligence in Predicting Progression from Intermediate to Advanced AMD. *Life (Basel)*. 2022 Mar 19;12(3):454.
43. Curcio CA, Millican CL. Basal linear deposit and large drusen are specific for early age-related maculopathy. *Arch Ophthalmol*. 1999;117:329-339.
44. Bressler NM, Silva JC, Bressler SB, Green WR. Clinicopathologic correlation of drusen and retinal pigment epithelial abnormalities in age-related macular degeneration. *Retina*. 1994;14:130-142
45. Khan AH, Chowers I, Lotery AJ. Beyond the Complement Cascade: Insights into Systemic Immunosenescence and Inflammaging in Age-Related Macular Degeneration and Current Barriers to Treatment. *Cells*. 2023 Jun 23;12(13):1708.
46. Fan Q, Maranville JC, Fritsche L, et al. HDL-cholesterol levels and risk of age-related macular degeneration: a multiethnic genetic study using Mendelian randomization. *Int J Epidemiol*. 2017 Dec 1;46(6):1891-1902.
47. Velilla S, García-Medina JJ, García-Layana A, et al. Smoking and age-related macular degeneration: review and update. *J Ophthalmol*. 2013;2013:895147.
48. Dighe S, Zhao J, Steffen L, Mares JA, et al. Diet patterns and the incidence of age-related macular degeneration in the Atherosclerosis Risk in Communities (ARIC) study. *Br J Ophthalmol*. 2020 Aug;104(8):1070-1076.
49. Zhang QY, Tie LJ, Wu SS, et al. Overweight, Obesity, and Risk of Age-Related Macular Degeneration. *Invest Ophthalmol Vis Sci*. 2016 Mar;57(3):1276-1283.
50. Mauschitz MM, Schmitz MT, Verzijden T, et al. Physical Activity, Incidence, and Progression of Age-Related Macular Degeneration: A Multicohort Study. *Am J Ophthalmol*. 2022 Apr;236:99-106.
51. Feng J, Xie F, Wu Z, Wu Y. Age-related macular degeneration and cardiovascular disease in US population: an observational study. *Acta Cardiologica*. 2023; 1-7. <https://doi.org/10.1080/00015385.2023.2295103>
52. Schnabolk G. Systemic inflammatory disease and AMD comorbidity. *Adv Exp Med Biol*. 2019;1185: 27-31.
53. Kaur C, Foulds WS, Ling EA. Hypoxia-ischemia and retinal ganglion cell damage. *Clin Ophthalmol*. 2008 Dec;2(4):879-889.
54. McBean GJ, Aslan M, Griffiths HR, Torrão RC. Thiol redox homeostasis in neurodegenerative disease. *Redox Biol*. 2015;5:186-194.
55. Johnson LV, Ozaki S, Staples MK, Erickson PA, Anderson DH. A potential role for immune complex pathogenesis in drusen formation. *Exp Eye Res*. 2000; 70(4):441-449
56. Johnson LV, Leitner WP, Staples MK, Anderson DH. Complement activation and inflammatory processes in Drusen formation and age related macular degeneration. *Exp Eye Res*. 2001; 73(6):887-896.
57. Handerson DH, Mullins RF, Hageman GS, Johnson LV. A role for local inflammation in the formation of drusen in the aging eye. *Am J Ophthalmol*. 2002;134(3):411-431.
58. Hollyfield JG, Bonilha VL, Rayborn ME, et al. Oxidative damage-induced inflammation initiates age-related macular degeneration. *Nat Med*. 2008 Feb;14(2):194-198.
59. Heloterä H, Kaarniranta K. A Linkage between Angiogenesis and Inflammation in Neovascular Age-Related Macular Degeneration. *Cells*. 2022 Nov 1;11(21):3453.
60. Pinna A, Porcu T, D'Amico-Ricci G, et al. Complete Blood Cell Count-Derived Inflammation Biomarkers in

- Men with Age-Related Macular Degeneration. *Ocul Immunol Inflamm.* 2019;27(6):932-936.
61. Shankar A, Mitchell P, Rojchana E, Tan J, Wang JJ. Association between circulating white blood cell count and long-term incidence of age-related macular degeneration: the Blue Mountains Eye Study. *Am J Epidemiol.* 2007 Feb 15;165(4):375-382.
  62. Naif S, Majed R, Mohieldin E, Hanan A, Lamis A, Maha A. Neutrophil-Lymphocyte Ratios in Dry Age-Related Macular Degeneration. *Ocul Immunol Inflamm.* 2022 Jul 13:1-6.
  63. Kurtul BE, Ozer PA. The Relationship between Neutrophil-to-lymphocyte Ratio and Age-related Macular Degeneration. *Korean J Ophthalmol.* 2016 Oct;30(5):377-381.
  64. Sengul EA, Artunay O, Kockar A, et al. Correlation of neutrophil/lymphocyte and platelet/lymphocyte ratio with visual acuity and macular thickness in age-related macular degeneration. *Int J Ophthalmol.* 2017 May 18;10(5):754-759.
  65. Xue CC, Cui J, Gao LQ, et al. Peripheral Monocyte Count and Age-Related Macular Degeneration. The Tongren Health Care Study. *Am J Ophthalmol.* 2021 Jul;227:143-153.
  66. Dąbrowska M, Zielińska A, Nowak I. Lipid oxidation products as a potential health and analytical problem. *Chemik.* 2015; 69: 89-94.
  67. Yin H, Xu L, Porter NA. Free radical lipid peroxidation: mechanisms and analysis. *Chem Rev.* 2011; 111: 5944-5972.
  68. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid Med Cell Longev.* 2014: 360438.
  69. Lüdtko L, Jürgens C, Ittermann T, Völzke H, Tost F. Age-related macular degeneration and associated risk factors in the population-based study of health in pomerania (SHIP-Trend) *Med Sci Monit.* 2019;25:6383-6390.
  70. Mao F, Yang X, Yang K, et al. Six-year incidence and risk factors for age-related macular degeneration in a rural Chinese population: the Handan Eye study. *Investig Ophthalmol Vis Sci.* 2019;60:4966-4971.
  71. Wang Y, Wang M, Zhang X, et al. The association between the lipids levels in blood and risk of age-related macular degeneration. *Nutrients.* 2016;8:663.
  72. Sasaki M, Harada S, Kawasaki Y, et al. Gender-specific association of early age-related macular degeneration with systemic and genetic factors in a Japanese population. *Sci Rep.* 2018;8:785.
  73. Kananen F, Strandberg T, Loukovaara S, Immonen I. Early middle age cholesterol levels and the association with age-related macular degeneration. *Acta Ophthalmol.* 2021 Nov;99(7):e1063-e1069. doi: 10.1111/aos.14774. Epub 2021 Feb 3.
  74. Lin JB, Halawa OA, Husain D, Miller JW, Vavvas DG. Dyslipidemia in age-related macular degeneration. *Eye (Lond).* 2022 Feb;36(2):312-318.
  75. Colijn JM, den Hollander AI, Demirkan A, et al. Increased high-density lipoprotein levels associated with age-related macular degeneration: evidence from the EYE-RISK and European Eye Epidemiology Consortia. *Ophthalmology.* 2019;126:393-406
  76. Yip JLY, Khawaja AP, Chan MPY, et al. Cross sectional and longitudinal associations between cardiovascular risk factors and age related macular degeneration in the EPIC-Norfolk Eye Study. *PLoS ONE.* 2015;10:e0132565.
  77. Burgess S, Davey, Smith G. Mendelian randomization implicates high-density lipoprotein cholesterol-associated mechanisms in etiology of age-related macular degeneration. *Ophthalmology.* 2017;124:1165-1174.
  78. Han X, Ong J-S, Hewitt AW, Gharahkhani P, MacGregor S. The effects of eight serum lipid biomarkers on age-related macular degeneration risk: a Mendelian randomization study. *Int J Epidemiol.* 2021;50:325-336
  79. Nordestgaard LT, Tybjaerg-Hansen A, Frikke-Schmidt R, Nordestgaard BG. Elevated apolipoprotein A1 and HDL cholesterol associated with age-related macular degeneration: 2 population cohorts. *J Clin Endocrinol Metab.* 2021;106(7):e2749-e2758.
  80. Hwang S, Kang SW, Choi J, et al. Lipid profile and future risk of exudative age-related macular degeneration development: a nationwide cohort study from South Korea. *Sci Rep.* 2022 Nov 5;12(1):18777.
  81. Yildirim Z, Ucgun NI, Yildirim F. The role of oxidative stress and antioxidants in the pathogenesis of age-related macular degeneration. *Clinics (Sao Paulo Braz.).* 2011; 66:743-746.
  82. Ułańczyk Z, Grabowicz A, Cecerska-Heryć E, Śleboda-Taront D, et al. Dietary and Lifestyle Factors Modulate the Activity of the Endogenous Antioxidant System in Patients with Age-Related Macular Degeneration: Correlations with Disease Severity. *Antioxidants (Basel).* 2020 Oct 5;9(10):954.
  83. Tokarz P, Kaarniranta K, Blasiak J. Role of antioxidant enzymes and small molecular weight antioxidants in the pathogenesis of age-related macular degeneration (AMD). *Biogerontology.* 2013 Oct;14(5):461-482.
  84. Torres RJA, Torres RJA, Luchini A, Ferreira ALDA. The nuclear factor E2-related factor 2 and age-related macular degeneration. *Arq Bras Oftalmol* 2023 Mar-Apr;86(2):178-187.
  85. Zafrilla P, Losada M, Perez A, Caravaca G, Mulero J. Biomarkers of oxidative stress in patients with wet age related macular degeneration. *J Nutr Health Aging.* 2013; 17:219-222.
  86. Meister A, Anderson ME. Glutathione. *Annu Rev Biochem.* 1983; 52(1):711-760.
  87. Sies H, Gerstenecker C, Menzel H, Flohé L. Oxidation in the NADP system and release of GSSG from hemoglobin-free perfused rat liver during peroxidative oxidation of glutathione by hydroperoxides. *FEBS.* 1972; 27(1):171-175.
  88. Lu SC. Glutathione synthesis. *Biochim Biophys Acta.* 2013;1830:3143-3153.
  89. Cantin AM, North SL, Hubbard RC, Crystal RG. Normal alveolar epithelial lining fluid contains high levels of glutathione. *J Appl Physiol.* 1987;63:152-157.
  90. Brodzka S, Baszyński J, Rektor K, et al. The Role of Glutathione in Age-Related Macular Degeneration (AMD). *Int J Mol Sci.* 2024 Apr 9;25(8):4158.

91. Tate DJ Jr, Newsome DA, Oliver PD. Metallothionein shows an age-related decrease in human macular retinal pigment epithelium. *Invest Ophthalmol Vis Sci.* 1993;34(7):2348-2351.
92. Liles MR, Newsome DA, Oliver PD. Antioxidant enzymes in the aging human retinal pigment epithelium. *Arch Ophthalmol.* 1991; 109(9):1285-1288.
93. Sun Y, Zheng Y, Wang C, Liu Y. Glutathione depletion induces ferroptosis, autophagy, and premature cell senescence in retinal pigment epithelial cells. *Cell Death Dis.* 2018; 9:753.
94. Brantley MA, Osborn MP, Sanders BJ, et al. Plasma biomarkers of oxidative stress and genetic variants in age-related macular degeneration. *Am J Ophthalmol.* 2012;153:460-467.
95. Nowak M, Swietochowska E, Wielkoszyński T, et al. Changes in blood antioxidants and several lipid peroxidation products in women with age-related macular degeneration. *Eur J Ophthalmol.* 2003;13:281-286.
96. Samiec PS, Drews-Botsch C, Flagg EW, et al. Glutathione in human plasma: Decline in association with aging, age-related macular degeneration, and diabetes. *Free Radic Biol Med.* 1998;24:699-704.
97. Coral K, Raman R, Rathi S, et al. Plasma homocysteine and total thiol content in patients with exudative age-related macular degeneration. *Eye.* 2006; 20:203-207.
98. Ates O, Alp HH, Mumcu U, et al. The effect of L-carnitine treatment on levels of malondialdehyde and glutathione in patients with age related macular degeneration. *Eur J Med.* 2008; 40:1-5.
99. Javadzadeh A, Ghorbanihaghjo A, Bahreini E, Rashtchizadeh N, Argani H, Alizadeh S. Plasma oxidized LDL and thiol-containing molecules in patients with exudative age-related macular degeneration. *Mol Vis* 2010; 16: 2578-2584.
100. Flohé L. The glutathione peroxidase reaction: molecular basis of the antioxidant function of selenium in mammals. *Curr Top Cell Regul.* 1985;27:473-478
101. Ohira A, Tanito M, Kaidzu S, Kondo T. Glutathione peroxidase induced in rat retinas to counteract photic injury. *Invest Ophthalmol Vis Sci.* 2003;44(3):1230-6.
102. Lu L, Oveson BC, Jo YJ, Lauer TW, et al. Increased expression of glutathione peroxidase 4 strongly protects retina from oxidative damage. *Antioxid Redox Signal.* 2009;11(4):715-724.
103. Zakowski JJ, Forstrom JW, Condell RA, Tappel AL. Attachment of selenocysteine in the catalytic site of glutathione peroxidase. *Biochem Biophys Res Commun.* 1978;84(1):248-253.
104. Ueta T, Inoue T, Furukawa T, et al. Glutathione peroxidase 4 is required for maturation of photoreceptor cells. *J Biol Chem.* 2012; 287:7675-7682.
105. Delcourt C, Cristol JP, Léger CL, Descomps B, Papoz L. Associations of antioxidant enzymes with cataract and age-related macular degeneration. The POLA Study. *Pathologies Oculaires Liées à l'Age. Ophthalmology.* 1999;106(2):215-222.
106. Prashar S, Pandav SS, Gupta A, Nath R. Antioxidant enzymes in RBCs as a biological index of age-related macular degeneration. *Acta Ophthalmol (Copenh).* 1993;71(2):214-218.
107. Cohen SM, Olin KL, Feuer WJ, Hjelmeland L, Keen CL, Morse LS. Low glutathione reductase and peroxidase activity in age-related macular degeneration. *Br J Ophthalmol.* 1994;78(10):791-794.
108. Plestina-Borjan I, Katusic D, Medvidovic-Grubisic M, S et al. Association of age-related macular degeneration with erythrocyte antioxidant enzymes activity and serum total antioxidant status. *Oxid Med Cell Longev.* 2015;2015:804054.
109. Venza I, Visalli M, Cucinotta M, Teti D, Venza M. Association between oxidative stress and macromolecular damage in elderly patients with age-related macular degeneration. *Aging Clin Exp Res.* 2012 Feb;24(1):21-27.
110. Mrowicka M, Mrowicki J, Szaflik JP, et al. Analysis of antioxidative factors related to AMD risk development in the polish patients. *Acta Ophthalmol.* 2017 Aug;95(5):530-536.
111. Qin L, Mroczkowska SA, Ekart A, Patel SR, Gibson JM, Gherghel D. Patients with early age-related macular degeneration exhibit signs of macro- and micro-vascular disease and abnormal blood glutathione levels. *Graefes Arch Clin Exp Ophthalmol.* 2014 Jan;252(1):23-30.
112. Fujii T, Mori K, Takahashi Y, et al. Immunohistochemical study of glutathione reductase in rat ocular tissues at different developmental stages. *Histochem J.* 2001;33(5):267-272.
113. Saxena M, Singhal SS, Awasthi YC. A specific, sensitive, and rapid method for the determination of glutathione and its application in ocular tissues. *Exp Eye Res.* 1992;55(3):461-468.
114. Huster D, Hjelle OP, Haug FM, Nagelhus EA, Reichelt W, Ottersen OP. Subcellular compartmentation of glutathione and glutathione precursors. A high resolution immunogold analysis of the outer retina of guinea pig. *Anat Embryol (Berl).* 1998;198(4):277-287.
115. Čolak E, Majkić-Singh N, Žoric L, Radosavljević A, Kosanović-Jaković N. The impact of inflammation to the antioxidant defense parameters in AMD patients. *Aging Clin Exp Res.* 2012 Dec;24(6):588-594.
116. McCord JM, Fridovich I. Superoxide dismutase. An enzymic function for erythrocyte (hemocuprein). *J Biol Chem.* 1969;244(22):6049-6055.
117. Indo HP, Yen HC, Nakanishi I, et al. A mitochondrial superoxide theory for oxidative stress diseases and aging. *J Clin Biochem Nutr.* 2015;56(1):1-7.
118. Yu BP. Cellular defenses against damage from reactive oxygen species. *Physiol Rev.* 1994;74(1):139-162
119. Yamamoto M, Lidia K, Gong H, Onitsuka S, Kotani T, Ohira A. Changes in manganese superoxide dismutase expression after exposure of the retina to intense light. *Histochem J.* 1999;31(2):81-87.
120. Sachdeva MM, Cano M, Handa JT. Nrf2 signaling is impaired in the aging RPE given an oxidative insult. *Exp Eye Res.* 2014;119:111-114.
121. Jia L, Dong Y, Yang H, Pan X, Fan R, Zhai L. Serum superoxide dismutase and malondialdehyde levels in a group of Chinese patients with age-related

- macular degeneration. *Aging Clin Exp Res.* 2011; 23:264–267.
122. Rex TS, Tsui I, Hahn P, et al. Adenovirus-mediated delivery of catalase to retinal pigment epithelial cells protects neighboring photoreceptors from photo-oxidative stress. *Hum Gene Ther.* 2004; 15:960–967.
  123. Masters C, Pegg M, Crane D. On the multiplicity of the enzyme catalase in mammalian liver. *Mol Cell Biochem.* 1986;70(2):113–120.
  124. Gaetani GF, Galiano S, Canepa L, Ferraris AM, Kirkman HN. Catalase and glutathione peroxidase are equally active in detoxification of hydrogen peroxide in human erythrocytes. *Blood.* 1989;73(1):334–339
  125. Robison WG Jr, Kuwabara T. Vitamin A storage and peroxisomes in retinal pigment epithelium and liver. *Invest Ophthalmol Vis Sci.* 1977;16(12):1110–1117.
  126. Beard ME, Davies T, Holloway M, Holtzman E. Peroxisomes in pigment epithelium and Müller cells of amphibian retina possess D-amino acid oxidase as well as catalase. *Exp Eye Res.* 1988;47(6):795–806.
  127. Frank RN, Amin RH, Puklin JE. Antioxidant enzymes in the macular retinal pigment epithelium of eyes with neovascular age-related macular degeneration. *Am J Ophthalmol.* 1999;127(6):694–709
  128. Wu R, Feng J, Yang Y, et al. Significance of Serum Total Oxidant/Antioxidant Status in Patients with Colorectal Cancer. *PLoS One.* 2017 Jan 19;12(1):e0170003.
  129. Totan Y, Yağci R, Bardak Y, et al. Oxidative macromolecular damage in age-related macular degeneration. *Curr Eye Res.* 2009; 34:1089-1093.
  130. Tarpey MM, Wink DA, Grisham MB. Methods for detection of reactive metabolites of oxygen and nitrogen: in vitro and in vivo considerations. *Am J Physiol, Regul Integr Comp Physiol.* 2004;286(3):R431–444.
  131. Erel O. A new automated colorimetric method for measuring total oxidant status. *Clin Biochem.* 2005 Dec;38(12):1103-1111.
  132. Uğurlu N, Aşık MD, Yülek F, Neselioglu S, Cagil N. Oxidative stress and anti-oxidative defence in patients with age-related macular degeneration. *Curr Eye Res.* 2013 Apr;38(4):497-502.
  133. Elbay A, Ozer OF, Akkan JCU, et al. Comparison of serum thiol-disulphide homeostasis and total antioxidant-oxidant levels between exudative age-related macular degeneration patients and healthy subjects. *Int Ophthalmol.* 2017 Oct;37(5):1095-1101.
  134. Crews H, Alink G, Andersen R, et al. A critical assessment of some biomarker approaches linked with dietary intake. *Br J Nutr.* 2001 Aug;86 Suppl 1:S5-35. doi: 10.1079/bjn2001337.
  135. Dalle-Donne I, Giustarini D, Colombo R, Rossi R, Milzani A. Protein carbonylation in human diseases. *Trends Mol Med.* 2003; 9:169-176.
  136. Levine RL. Carbonyl modified proteins in cellular regulation, aging, and disease. *Free Radic Biol Med.* 2002; 32:790-796.
  137. Regazzoni L, de Courten B, Garzon D, et al. A carnosine intervention study in overweight human volunteers: bioavailability and reactive carbonyl species sequestering effect. *Sci Rep.* 2016; 6:27224
  138. de Almeida Torres RJ, Moreto F, Luchini A, et al. Carnosine supplementation and retinal oxidative parameters in a high-calorie diet rat model. *BMC Ophthalmol.* 2023 Dec 8;23(1):502.
  139. Xu H, Chen M, Forrester JV. Para-inflammation in the aging retina. *Prog Retin Eye Res.* 2009; 28:348–368.
  140. Ambati J, Ambati BK, Yoo SH, Ianchulev S, Adamis AP. Age-related macular degeneration: etiology, pathogenesis, and therapeutic strategies. *Surv Ophthalmol.* 2003; 48:257–293.
  141. Torres RJDA, Torres R, Luchini A, Ferreira ALDA. Transcription Factor NF- $\kappa$ B and Molecules Derived from its Activation in Age-Related Macular Degeneration. *Integr J Med Sci [Internet].* 2021 Mar. 30 [cited 2023 Oct. 1];8. Available from: <https://mbmj.org/index.php/ijms/article/view/393>
  142. de Almeida Torres RJ, de Almeida Torres RJ, Luchini A, Anjos Ferreira AL. The oxidative and inflammatory nature of age-related macular degeneration. *J Clin Ophthalmol Res.* 2022;10:3-8.
  143. Akdis M, Burgler S, Cramer R, et al. Interleukins, from 1 to 37, and interferon- $\gamma$ : receptors, functions, and roles in diseases. *J Allergy Clin Immunol.* 2011 Mar;127(3):701-21.e1-70.
  144. Murphy PM, Baggiolini M, Charo IF, et al. International union of pharmacology. XXII. Nomenclature for chemokine receptors. *Pharmacol Rev.* 2000 Mar;52(1):145-176.
  145. Kopf M, Baumann H, Freer G, et al. Impaired immune and acute-phase responses in interleukin-6-deficient mice. *Nature.* 1994;368:339-342.
  146. Elnor VM, Scales W, Elnor SG, Danforth J, Kunkel SL, Strieter RM. Interleukin-6 (IL-6) gene expression and secretion by cytokine-stimulated human retinal pigment epithelial cells. *Exp Eye Res.* 1992;54:361-368.
  147. Nagineni CN, Detrick B, Hooks JJ. Synergistic effects of gamma interferon on inflammatory mediators that induce interleukin-6 gene expression and secretion by human retinal pigment epithelial cells. *Clin Diagn Lab Immunol.* 1994;1:569–577.
  148. Corbi P, Rahmati M, Delwail A, et al. Circulating soluble gp130, soluble IL-6R, and IL-6 in patients undergoing cardiac surgery, with or without extracorporeal circulation. *Eur J Cardiothorac Surg.* 2000;18:98-103.
  149. Van Snick J. Interleukin-6: an overview. *Annu Rev Immunol.* 1990; 8:253-278.
  150. Koto T, Nagai N, Mochimaru H, et al. Eicosapentaenoic acid is anti-inflammatory in preventing choroidal neovascularization in mice. *Invest Ophthalmol Vis Sci.* 2007;48:4328-4334.
  151. Paimela T, Ryhänen T, Mannermaa E, et al. The effect of 17 $\beta$ -estradiol on IL-6 secretion and NF-kappaB DNA-binding activity in human retinal

- pigment epithelial cells. *Immunol Lett.* 2007;110:139-144.
152. Cohen T, Nahari D, Cerem LW, Gera N, Levi B. Interleukin-6 induces the expression of vascular endothelial growth factor. *J Biol Chem.* 1996; 271:736-741
  153. Nahavandipour A, Krogh Nielsen M, Sørensen TL, Subhi Y. Systemic levels of interleukin-6 in patients with age-related macular degeneration: a systematic review and meta-analysis. *Acta Ophthalmol.* 2020 Aug;98(5):434-444.
  154. Rock KL, Lai JJ, Kono H. Innate and adaptive immune responses to cell death. *Immunological reviews.* 2011; 243(1):191-205.
  155. Bird S, Zou J, Wang T, Munday B, Cunningham C, Secombes CJ. Evolution of interleukin-1beta. *Cytokine Growth Factor Rev.* 2002 Dec;13(6):483-502.
  156. Miao EA, Rajan JV, Aderem A. Caspase-1-induced pyroptotic cell death. *Immunological reviews.* 2011; 243(1): 206-214.
  157. Doyle SL, Campbell M, Ozaki E, et al. NLRP3 has a protective role in age-related macular degeneration through the induction of IL-18 by drusen components. *Nat Med.* 2012;18(5):791-798.
  158. Anderson OA, Finkelstein A, Shima DT. A2E induces IL-1ss production in retinal pigment epithelial cells via the NLRP3 inflammasome. *PLoS One.* 2013; 8(6): e67263
  159. Yang D, Elner SG, Bian ZM, et al. Pro-inflammatory cytokines increase reactive oxygen species through mitochondria and NADPH oxidase in cultured RPE cells. *Exp Eye Res.* 2007;85:462-472.
  160. Calabro P, Willerson JT, Yeh ET. Inflammatory cytokines stimulated C-reactive protein production by human coronary artery smooth muscle cells. *Circulation.* 2003; 108:1930-1932.
  161. Venugopal SK, Devaraj S, Jialall. Macrophage conditioned medium induces the expression of C-reactive protein in human aortic endothelial cells: potential for paracrine/autocrine effects. *Am J Pathol.* 2005; 166:1265-1271.
  162. Pearson TA, Mensah GA, Hong Y, Smith SC Jr, CDC; AHA. CDC/AHA Workshop on Markers of Inflammation and Cardiovascular Disease: Application to Clinical and Public Health Practice: overview. *Circulation.* 2004 Dec 21;110(25):e543-544.
  163. Ganter U, Arcone R, Toniatti C, Morrone G, Ciliberto G. Dual control of C-reactive protein gene expression by interleukin-1 and interleukin-6. *EMBO J.* 1989 Dec 1; 8(12):3773-3779
  164. Hogg RE, Woodside JV, Gilchrist SE, et al. Cardiovascular disease and hypertension are strong risk factors for choroidal neovascularization. *Ophthalmology.* 2008 Jun;115(6):1046-1052.e2.
  165. Boekhoorn SS, Vingerling JR, Witteman JC, Hofman A, de Jong PT. C-reactive protein level and risk of aging macula disorder: The Rotterdam Study. *Arch Ophthalmol.* 2007;125(10):1396-1401.
  166. Schaumberg DA, Christen WG, Buring JE, Glynn RJ, Rifai N, Ridker PM. High-sensitivity C-reactive protein, other markers of inflammation, and the incidence of macular degeneration in women. *Arch Ophthalmol.* 2007;125(3):300-305.
  167. Čolak E, Kosanović-Jaković N, Žorić L, Radosavljević A, Stanković S, Majkić-Singh N. The association of lipoprotein parameters and C-reactive protein in patients with age-related macular degeneration. *Ophthalmic Res.* 2011;46:125-132.
  168. Feng C, Krogh Nielsen M, Sørensen TL, Subhi Y. Systemic levels of C-reactive protein in patients with age-related macular degeneration: A systematic review with meta-analyses. *Mech Ageing Dev.* 2020 Oct;191:111353.
  169. Hong T, Tan AG, Mithchell P, Wang JJ. A review and meta-analysis of the association between C-reactive protein and age-related macular degeneration. *Surv Ophthalmol.* 2011;56:184-194.
  170. Ryuto M, Ono M, Izumi H, et al. Induction of vascular endothelial growth factor by tumor necrosis factor alpha in human glioma cells. Possible roles of SP-1. *J Biol Chem.* 1996;271:28220-28228.
  171. Patterson C, Perrella MA, Endege WO, Yoshizumi M, Lee ME, Haber E. Downregulation of vascular endothelial growth factor receptors by tumor necrosis factor-alpha in cultured human vascular endothelial cells. *J Clin Invest.* 1996;98:490-496.
  172. Oh H, Takagi H, Takagi C, et al. The potential angiogenic role of macrophages in the formation of choroidal neovascular membranes. *Invest Ophthalmol Vis Sci.* 1999 Aug;40(9):1891-1898.
  173. Hsu H, Shu HB, Pan MG, Goeddel DV. TRADD-TRAF2 and TRADD-FADD interactions define two distinct TNF receptor 1 signal transduction pathways. *Cell.* 1996; 84:299-308.
  174. Semkova I, Muether PS, Kuebbeler M, Meyer KL, Kociok N, Joussen AM. Recruitment of Blood-Derived Inflammatory Cells Mediated via Tumor Necrosis Factor-Receptor 1b Exacerbates Choroidal Neovascularization. *Invest Ophthalmol Vis Sci.* 2011; 52:6101-6108.
  175. Luo D, Luo Y, He Y, et al. Differential functions of tumor necrosis factor receptor 1 and 2 signaling in ischemia-mediated arteriogenesis and angiogenesis. *Am J Pathol.* 2006; 169:1886-1898
  176. Udsen M, Tagmose C, Garred P, Nissen MH, Faber C. Complement activation by RPE cells preexposed to TNF $\alpha$  and IFN $\gamma$ . *Exp Eye Res.* 2022 May;218:108982.
  177. Faber C, Jehs T, Juel HB, et al. Early and exudative age-related macular degeneration is associated with increased plasma levels of soluble TNF receptor II. *Acta Ophthalmol.* 2015 May;93(3):242-247.
  178. Khan AH, Pierce CO, De Salvo G, et al. The effect of systemic levels of TNF-alpha and complement pathway activity on outcomes of VEGF inhibition in neovascular AMD. *Eye (Lond).* 2022 Nov;36(11):2192-2199.
  179. Papadopoulos Z. The role of the cytokine TNF- $\alpha$  in choroidal neovascularization: a systematic review. *Eye (Lond).* 2024 Jan;38(1):25-32.
  180. Baggiolini M, Walz A, Kunkel SL. Neutrophil-activating peptide-1/interleukin 8, a novel

- cytokine that activates neutrophils. *J Clin Invest*. 1989; 84:1045-1049
181. Lechner J, Chen M, Hogg RE, et al. Peripheral blood mononuclear cells from neovascular age-related macular degeneration patients produce higher levels of chemokines CCL2 (MCP-1) and CXCL8 (IL-8). *J Neuroinflammation*. 2017 Feb 23;14(1):42.
  182. Zwahlen R, Walz A, Rot A. In vitro and in vivo activity and pathophysiology of human interleukin-8 and related peptides. *Int Rev Exp Pathol*. 1993;34:27-42.
  183. Agrawal R, Balne PK, Wei X, et al. Cytokine Profiling in Patients With Exudative Age-Related Macular Degeneration and Polypoidal Choroidal Vasculopathy. *Invest Ophthalmol Vis Sci*. 2019 Jan 2;60(1):376-382.
  184. Liukkonen MPK, Paterno JJ, Kivinen N, Siintamo L, Koskela AKJ, Kaarniranta K. Epithelial-mesenchymal transition-related serum markers ET-1, IL-8 and TGF- $\beta$ 2 are elevated in a Finnish wet age-related macular degeneration cohort. *Acta Ophthalmol*. 2022 Aug;100(5):e1153-e1162.
  185. Roshanipour N, Shahriyari E, Ghaffari Laleh M, et al. Associations of TLR4 and IL-8 genes polymorphisms with age-related macular degeneration (AMD): a systematic review and meta-analysis. *Ophthalmic Genet*. 2021 Dec;42(6):641-649.
  186. Ricci F, Staurengi G, Lepre T, et al. Haplotypes in IL-8 Gene Are Associated to Age-Related Macular Degeneration: A Case-Control Study. *PLoS ONE*. 2013; 8(6): e66978.
  187. Zhao C, Wu M, Zeng N, et al. Cancer-associated adipocytes: Emerging supporters in breast cancer. *J Exp Clin Cancer Res*. 2020;39(1):156.
  188. Sallusto F, Mackay CR, Lanzavecchia A. The role of chemokine receptors in primary, effector, and memory immune responses. *Annu Rev Immunol*. 2000;18:593-620.
  189. Weber C, Schober A, Zernecke A. Chemokines: key regulators of mononuclear cell recruitment in atherosclerotic vascular disease. *Arterioscler Thromb Vasc Biol* 2004; 24(11):1997-2008. (Luster AD. Chemokines--chemotactic cytokines that mediate inflammation. *N Engl J Med*. 1998;338(7):436-445.
  190. Peterson PK, Hu S, Salak-Johnson J, Molitor TW, Chao CC. Differential Production of and Migratory Response to Beta Chemokines by Human Microglia and Astrocytes. *J Infect Dis*. 1997;175(2):478-481
  191. Prodjosudjadi W, Gerritsma JS, Klar-Mohamad Ngaisah, et al. Production and cytokine-mediated regulation of monocyte chemoattractant protein-1 by human proximal tubular epithelial cells. *Kidney Int*. 1995;48(5):1477-1486.
  192. Grassia G, Maddaluno M, Guglielmotti A, et al. The anti-inflammatory agent bindarit inhibits neointima formation in both rats and hyperlipidaemic mice. *Cardiovasc Res*. 2009;84(3):485-493.
  193. Pons M, Marin-Castano ME. Cigarette smoke-related hydroquinone dysregulates MCP-1, VEGF and PEDF expression in retinal pigment epithelium in vitro and in vivo. *PLoS One*. 2011;6:e16722.
  194. Jonas JB, Tao Y, Neumaier M, Findeisen P. Monocyte chemoattractant protein 1, intercellular adhesion molecule 1, and vascular cell adhesion molecule 1 in exudative age-related macular degeneration. *Arch Ophthalmol*. 2010;128:1281-1286.
  195. Mata NL, Weng J, Travis GH. Biosynthesis of a major lipofuscin fluorophore in mice and humans with ABCR-mediated retinal and macular degeneration. *Proc Natl Acad Sci U.S.A.* 2000;97:1549.
  196. Higgins GT, Wang JH, Dockery P, Cleary PE, Redmond HP. Induction of angiogenic cytokine expression in cultured RPE by ingestion of oxidized photoreceptor outer segments. *Invest Ophthalmol Vis Sci*. 2003;44:1775-82.
  197. Killingsworth MC, Sarks JP, Sarks SH. Macrophages related to Bruch's membrane in age-related macular degeneration. *Eye (Lond)*. 1990;4 (pt 4): 613-621.
  198. Sarks JP, Sarks SH, Killingsworth MC. Morphology of early choroidal neovascularisation in age-related macular degeneration: correlation with activity. *Eye (Lond)*. 1997; 11 (Pt 4): 515-522.
  199. Lopez PF, Lambert HM, Grossniklaus HE, Sternberg PJr. Well-defined subfoveal choroidal neovascular membranes in age-related macular degeneration. *Ophthalmology*. 1993;100: 415-422.
  200. Yamada K, Sakurai E, Itaya M. Inhibition of laser-induced choroidal neovascularization by atorvastatin by downregulation of monocyte chemotactic protein-1 synthesis in mice. *Invest Ophthalmol Vis Sci*. 2007;48:1839-1843
  201. Xie P, Kamei M, Suzuki M, et al. Suppression and regression of choroidal neovascularization in mice by a novel CCR2 antagonist, INCB3344. *PLoS One*. 2011;6:e28933.
  202. Luhmann UF, Robbie S, Munro PM, et al. The drusenlike phenotype in aging Ccl2-knockout mice is caused by an accelerated accumulation of swollen autofluorescent subretinal macrophages. *Invest Ophthalmol Vis Sci*. 2009 Dec;50(12):5934-5943.
  203. Ambati J, Anand A, Fernandez S, et al. An animal model of age-related macular degeneration in senescent Ccl-2- or Ccr-2-deficient mice. *Nat Med*. 2003 Nov;9(11):1390-1397.
  204. Yang D, Elnor SG, Chen X. MCP-1-activated monocytes induce apoptosis in human retinal pigment epithelium. *Invest Ophthalmol Vis Sci*. 2011;52:6026-6034.
  205. Sennlaub F, Auvynet C, Calippe B, et al. CCR2(+) monocytes infiltrate atrophic lesions in age-related macular disease and mediate photoreceptor degeneration in experimental subretinal inflammation in Cx3cr1 deficient mice. *EMBO Mol Med*. 2013 Nov;5(11):1775-1793.
  206. Falk MK, Singh A, Faber C, Nissen MH, Hviid T, Sørensen TL. CX3CL1/CX3CR1 and CCL2/CCR2 chemokine/chemokine receptor complex in patients with AMD. *PLoS One*. 2014 Dec 15;9(12):e112473.
  207. Zor RK, Erşan S, Küçük E, Yıldırım G, Sarı İ. Serum malondialdehyde, monocyte chemoattractant protein-1, and vitamin C levels in wet type age-related macular degeneration patients. *Ther Adv*

- Ophthalmol. 2020 Sep 30;12:2515841420951682.
208. Zhou H, Zhao X, Chen Y. Plasma Cytokine Profiles in Patients With Polypoidal Choroidal Vasculopathy and Neovascular Age-Related Macular Degeneration. *Asia Pac J Ophthalmol (Phila)*. 2022 Nov 1;11(6):536-542.
  209. Palestine AG, Wagner BD, Patnaik JL, et al. Plasma C-C Chemokine Concentrations in Intermediate Age-Related Macular Degeneration. *Front Med (Lausanne)*. 2021 Nov 18;8:710595.
  210. Gudauskiene G, Vilkeviciute A, Gedvilaite G, Liutkeviciene R, Zaliuniene D. CCL2, CCR2 Gene Variants and CCL2, CCR2 Serum Levels Association with Age-Related Macular Degeneration. *Life (Basel)*. 2022 Jul 12;12(7):1038.
  211. Grunin M, Burstyn-Cohen T, Hagbi-Levi S, Peled A, Chowers I. Chemokine receptor expression in peripheral blood monocytes from patients with neovascular age-related macular degeneration. *Invest Ophthalmol Vis Sci*. 2012 Aug 7;53(9):5292-300.
  212. Subhi Y, Krogh Nielsen M, Molbech CR, Sørensen TL. Altered proportion of CCR2<sup>+</sup> and CX3CR1<sup>+</sup> circulating monocytes in neovascular age-related macular degeneration and polypoidal choroidal vasculopathy. *Clin Exp Ophthalmol*. 2018 Aug;46(6):661-669.
  213. Schoggins JW. Interferon-stimulated genes: roles in viral pathogenesis. *Curr Opin Virol*. 2014;6:40-46.
  214. Reyes-Vázquez C, Prieto-Gómez B, Dafny N. Interferon modulates central nervous system function. *Brain Res*. 2012;1442:76-89.
  215. Chow KT, Gale M Jr. SnapShot: Interferon Signaling. *Cell*. 2015 Dec 17;163(7):1808-1808.e1. doi: 10.1016/j.cell.2015.12.008. PMID: 26687364.
  216. Lückoff A, Caramoy A, Scholz R, Prinz M, Kalinke U, Langmann T. Interferon-beta signaling in retinal mononuclear phagocytes attenuates pathological neovascularization. *EMBO Mol Med*. 2016 Jun 1;8(6):670-678.
  217. Li R, Maminishkis A, Wang FE, Miller SS. PDGF-C and -D induced proliferation/migration of human RPE is abolished by inflammatory cytokines. *Invest Ophthalmol Vis Sci*. 2007;48:5722-5732.
  218. Borden EC, Hogan TF, Voelkel JG. Comparative antiproliferative activity in vitro of natural interferons alpha and beta for diploid and transformed human cells. *Cancer Res*. 1982;42:4948-4953.
  219. Indraccolo S. Interferon-alpha as angiogenesis inhibitor: learning from tumor models. *Autoimmunity*. 2010;43:244-247.
  220. Battle TE, Lynch RA, Frank DA. Signal transducer and activator of transcription 1 activation in endothelial cells is a negative regulator of angiogenesis. *Cancer Res*. 2006;66:3649-3657.
  221. Kawano Y, Matsui N, Kamihigashi S, Narahara H, Miyakawa I. Effects of interferon-gamma on secretion of vascular endothelial growth factor by endometrial stromal cells. *Am J Reprod Immunol*. 2000;43:47-52.
  222. Wu Z, Lauer TW, Sick A, Hackett SF, Campochiaro PA. Oxidative stress modulates complement factor H expression in retinal pigmented epithelial cells by acetylation of FOXO3. *J Biol Chem*. 2007;282:22414-22425
  223. Liu B, Faia L, Hu M, Nussenblatt RB. Pro-angiogenic effect of IFN gamma is dependent on the PI3K/mTOR/translational pathway in human retinal pigmented epithelial cells. *Mol Vis*. 2010;16:184-193.
  224. Jiang K, Cao S, Cui JZ, Matsubara JA. Immunomodulatory Effect of IFN-gamma in AMD and its Role as a Possible Target for Therapy. *J Clin Exp Ophthalmol*. 2013 Feb 26;Suppl 2:0071-76.
  225. Cousins SW, Espinosa-Heidmann DG, Miller DM, et al. Macrophage activation associated with chronic murine cytomegalovirus infection results in more severe experimental choroidal neovascularization. *PLoS Pathog*. 2012;8:e1002671.
  226. Brown J, Wallet MA, Krastins B, Sarracino D, Goodenow MM. Proteome bioprofiles distinguish between M1 priming and activation states in human macrophages. *J Leukoc Biol*. 2010;87:655-662.
  227. Supanji S, Perdamaian ABI, Wardhana FS, et al. Circulating levels of Interferon-Gamma in patients with neovascular age-related macular degeneration in Yogyakarta. *Med J Malaysia*. 2022 Jul;77(Suppl 1):62-65.
  228. Yu Y, Ren XR, Wen F, Chen H, Su SB. T-helper-associated cytokines expression by peripheral blood mononuclear cells in patients with polypoidal choroidal vasculopathy and age-related macular degeneration. *BMC Ophthalmol*. 2016 Jun 7;16:80.
  229. Fonseca V, Guba SC, Fink LM. Hyperhomocysteinemia and the endocrine system: implications for Atherosclerosis and thrombosis. *Endocr Rev*. 1999;20:738-759.
  230. Lee YJ, Ke CY, Tien N, Lin PK. Hyperhomocysteinemia Causes Chorioretinal Angiogenesis with Placental Growth Factor Upregulation. *Sci Rep*. 2018;8:15755.
  231. Clark R. Lowering blood homocysteine with folic acid based supplements: Meta-analysis of randomised trials. *BMJ*. 1998;316(7135):894-898.
  232. Graham IM, Daly LE, Refsum HM, et al. Plasma homocysteine as a risk factor for vascular disease. The European Concerted Action Project. *JAMA*. 1997 Jun 11;277(22):1775-1781.
  233. Hustad S, Ueland PM, Vollset SE, Zhang Y, Bjorke-Monsen AL, Schneede J. Riboflavin as a determinant of plasma total homocysteine: effect modification by the methylenetetrahydrofolate reductase C677T polymorphism. *Clin Chem*. 2000; 46:1065-1071.
  234. Smith AD, Refsum H. Homocysteine - from disease biomarker to disease prevention. *J Intern Med*. 2021 Oct;290(4):826-854.
  235. Smith AD, Refsum H, Bottiglieri T, et al. Homocysteine and Dementia: An International Consensus Statement. *J Alzheimers Dis*. 2018;62:561-570.
  236. Fotiou P, Raptis A, Apergis G, Dimitriadis G, Vergados I, Theodossiadis P. Vitamin status as a determinant of serum homocysteine concentration in type 2 diabetic retinopathy. *J Diabetes Res*. 2014;2014:807209.

237. Anderson JL, Muhlestein JB, Horne BD, et al. Plasma homocysteine predicts mortality independently of traditional risk factors and C-reactive protein in patients with angiographically defined coronary artery disease. *Circulation*. 2000 Sep 12;102(11):1227-1232.
238. Nygård O, Nordrehaug JE, Refsum H, Ueland PM, Farstad M, Vollset SE. Plasma homocysteine levels and mortality in patients with coronary artery disease. *N Engl J Med*. 1997 Jul 24;337(4):230-236.
239. Elsherbiny NM, Sharma I, Kira D, et al. Homocysteine Induces Inflammation in Retina and Brain. *Biomolecules*. 2020 Mar 3;10(3):393.
240. Nowak M, Swietochowska E, Wielkoszynski T, et al. Homocysteine, vitamin B12, and folic acid in age-related macular degeneration. *Eur J Ophthalmol*. 2005;15:764–767
241. Kamburoglu G, Gumus K, Kadayifcilar S, Eldem B. Plasma homocysteine, vitamin B12 and folate levels in age-related macular degeneration. *Graefes Arch. Clin Exp Ophthalmol*. 2006;244:565–569.
242. Tawfik A, Samra YA, Elsherbiny NM, Al-Shabrawey M. Implication of Hyperhomocysteinemia in Blood Retinal Barrier (BRB) Dysfunction. *Biomolecules*. 2020 Jul 29;10(8):1119.
243. Ibrahim AS, Mander S, Hussein KA, et al. Hyperhomocysteinemia disrupts retinal pigment epithelial structure and function with features of age-related macular degeneration. *Oncotarget*. 2016;7:8532–8545.
244. Samra YA, Kira D, Rajpurohit P, et al. Implication of N-Methyl-d-Aspartate Receptor in Homocysteine-Induced Age-Related Macular Degeneration. *Int J Mol Sci*. 2021;22:9356.
245. Tawfik A, Al-Shabrawey M, Roon P, et al. Alterations of retinal vasculature in cystathionine-beta-synthase mutant mice, a model of hyperhomocysteinemia. *Investig Ophthalmol Vis Sci*. 2013;54: 939–949.
246. Pinna A, Zaccheddu F, Boscia F, Carru C, Solinas G. Homocysteine and risk of age-related macular degeneration: a systematic review and meta-analysis. *Acta Ophthalmol*. 2018 May;96(3):e269-e276.
247. Huang P, Wang F, Sah BK, et al. Homocysteine and the risk of age-related macular degeneration: a systematic review and meta-analysis. *Sci Rep*. 2015 Jul 21;5:10585.
248. Gopinath B, Flood VM, Rochtchina E, Wang JJ, Mitchell P. Homocysteine, folate, vitamin B-12, and 10-y incidence of age-related macular degeneration. *Am J Clin Nutr*. 2013; 98: 129–135.
249. Merle BM, Silver RE, Rosner B, Seddon JM. Dietary folate, B vitamins, genetic susceptibility and progression to advanced nonexudative age-related macular degeneration with geographic atrophy: a prospective cohort study. *Am J Clin Nutr*. 2016; 103: 1135–1144.
250. Christen WG, Glynn RJ, Chew EY, Albert CM, Manson JE. Folic acid, pyridoxine, and cyanocobalamin combination treatment and age-related macular degeneration in women: the Women's Antioxidant and Folic Acid Cardiovascular Study. *Arch Intern Med*. 2009; 169: 335–341