



REVIEW ARTICLE

Pentosan Polysulfate, An Anti-Viral Heparinoid, Prevents Severe Acute Respiratory Syndrome Corona Virus-2 Infection and Treats Symptoms of Long Coronavirus Disease

Margaret M. Smith¹, James Melrose^{1,2,3*}

¹Raymond Purves Bone and Joint Research Laboratory, Kolling Institute, St. Leonards, NSW 2065, Australia,

²School of Medical Sciences, Faculty of Medicine and Health, The University of Sydney at Royal North Shore Hospital, St. Leonards, NSW 2065, Australia,

³Graduate School of Biomedical Engineering, University of New South Wales, Sydney, NSW 2052, Australia.

james.melrose@sydney.edu.au

OPEN ACCESS

PUBLISHED

31 August 2025

CITATION

Smith, MM., Melrose, J., 2025. Pentosan Polysulfate, An Anti-Viral Heparinoid, Prevents Severe Acute Respiratory Syndrome Corona Virus-2 Infection and Treats Symptoms of Long Coronavirus Disease. Medical Research Archives, [online] 13(8).

<https://doi.org/10.18103/mra.v13.18.6735>

COPYRIGHT

© 2025 European Society of Medicine. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

DOI

<https://doi.org/10.18103/mra.v13.18.6735>

ISSN

2375-1924

ABSTRACT

This study highlights the roles of pentosan polysulfate as a decoy anti-viral prophylactic that prevents severe acute respiratory syndrome coronavirus-2 infection. PPS also has multifunctional cell and tissue protective properties relevant to the treatment of the symptoms produced by long COVID disease. PPS has heparan sulfate (HS)-like properties, a key functional component of the lung glycocalyx. The glycocalyx is also rich in hyaluronan which has important cell shielding and cell regulatory properties. A healthy glycocalyx prevents access of viral particles to cell surface heparan sulfate-proteoglycans (syndecan, glypican) which act as viral receptors. Pentosan polysulfate promotes hyaluronan synthesis by many cell types, ensuring cells are surrounded by a healthy protective glycocalyx. Hyaluronan, however, has a relatively short biological half-life and is susceptible to degradation by hyaluronidases that are upregulated by inflammatory cytokines in acute respiratory distress syndrome in COVID-19 disease. This results in the glycocalyx becoming degraded and endothelial cells dysfunctional in COVID-19 disease. Prevention of viral interaction with the host cell surface intercepted by pentosan polysulfate, a decoy viral binding prophylactic agent, blocks viral interaction with cell-surface heparan sulfate, preventing viral interactions with other cell surface receptors such as neuropilin-1 and angiotensin-converting enzyme 2. Co-operation between heparan sulfate, neuropilin-1 and angiotensin-converting enzyme 2 facilitates the infection of host cells with severe acute respiratory syndrome coronavirus 2, thus if the initial interaction with heparan sulfate is blocked this prevents the subsequent viral interactive stages. Pentosan polysulfate also has multifunctional cell and tissue protective properties, broad anti-oxidant and anti-inflammatory properties and inhibits cytokine production in acute respiratory disease syndrome. Pentosan polysulfate inhibits p38 mitogen-activated protein kinase and nuclear factor- κ B activation, reducing the production of pro-inflammatory cytokines such as tumor necrosis factor- α , interleukin-1 β and interleukin-6. Furthermore, pentosan polysulfate is processed by enzymes of the gut microbiome into prebiotic xylo-oligosaccharides that preserve gut health and combat gut dysbiosis seen in COVID-19 disease. Studies are thus warranted to fully assess pentosan polysulfate as an anti-severe acute respiratory syndrome coronavirus-2 prophylactic agent and its multifunctional cell and tissue protective properties. Furthermore, from a practical and economic point of view, treatment with pentosan polysulfate would offer substantial cost-benefit advantages over conventional vaccine and antibiotic treatments and could also be used in an adjunctive capacity with existing therapies, offering flexibility in its use.

Keywords: Pentosan polysulfate; SP54; Neuropilin-1; angiotensin-converting enzyme-2; Heparan sulfate; SARS-CoV-2; HIV; Herpes simplex; Dengue virus; Papillomavirus.

1. Introduction

The aim of this study was to highlight the roles of pentosan polysulfate (PPS), a semisynthetic heparinoid, as a decoy anti-viral prophylactic in the prevention of SARS-CoV-2 infection of host cells. The multifunctional cell and tissue protective properties of PPS are also described illustrating how PPS may be employed to treat the multi-parameter symptomatology that characterises long COVID disease (Figure 1).

1.1 COVID-19 IS A PANDEMIC VIRAL DISORDER OF GLOBAL IMPACT

COVID-19 is a pandemic disease that emerged in Wuhan, China in late 2019 with the emergence of severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2), a single strand RNA virus that is 96.2% identical in genomic sequence to the bat CoV RaTG13 virus. SARS CoV-2 is highly transmissible through aerosols, droplets, fomite affected surfaces or direct skin contact¹. Historically, SARS-CoV-2 has demonstrated an unprecedented infectious global profile and has undergone rapid evolutionary mutational changes as part of its natural life cycle into several variants which avoid immune detection. Some of these SARS-CoV-2 variants bind more efficiently and with greater rapidity to respiratory epithelial cells, rendering these viral forms significantly more infectious^{2,3}. The Omicron variant is currently a dominant global variant, 99% of all variants circulating in the U.S. are mutations of Omicron, most commonly EG.5 (24% of all SARS CoV-2 strains) and FL 1.5.1 (14%) (<https://covid.cdc.gov/covid-data-tracker/#variant-proportions>)⁴. A distinctive feature of these Omicron variants is a change in their infective profiles displaying more effective infection of the nose and throat rather than the original Wuhan and Delta variants which primarily infected the lungs⁵. Respiratory distress is a prominent feature of SARS CoV-2 infections, but other organ systems can also be affected including the brain, liver, heart, and kidney⁶⁻⁸. Symptoms of SARS CoV-2 infection include fever, anosmia, ageusia, dry cough, fatigue, breathlessness, hair-

loss and so-called brain-fogging with a decline in problem solving capability, cognition, ability to concentrate, negative neurological impact and an increase in long-term anxiety^{9,10}. These symptoms can be mild, moderate or severe and a fatality rate of 1 in 100 is reported depending on the comorbidities that patients display; these can significantly impact the severity of COVID-19. A global systematic review of 76 studies that examined a total of 17,860,001 patients across 14 countries showed that age >75 years, male sex, severe obesity, lymphopenia, and cancer increased the impact of SARS CoV-2 infection on health and well-being¹¹.

Viral recombination is a normal part of the viral life cycle but results in an extremely wide spread in epitope presentations and these continually undergo changes in structure. This makes it problematic to raise vaccines or antibodies to current infectious viral strains and these require continual updating putting enormous strain on viral treatment resources. In the present study we have shown how PPS can inhibit all viral classes by blocking the interaction of virion particles with cell surface HS and we propose that this has considerable merit as a preventative approach to treatment of potential infections by SARS CoV-2 but is also applicable to other viral classes. Furthermore, PPS is a pleotropic cell and tissue protective agent^{12,13} as we have outlined in this review and is suitable for the treatment of many facets of the varied symptomatology encountered in COVID-19 infected tissues throughout the human body. This is a further strength of PPS as a therapeutic agent for viral conditions.

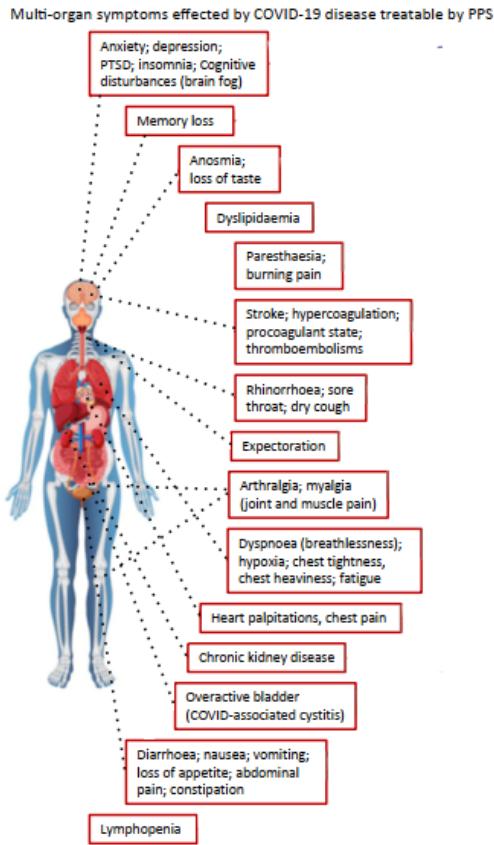


Figure 1. The multifunctional properties of pentosan polysulfate. Multi-organ systems affected by COVID-19 treatable with pentosan polysulfate.

1.2 PENTOSAN POLYSULFATE, A PROPHYLACTIC ANTI-VIRAL PLEOTROPIC CELL AND TISSUE PROTECTIVE AGENT

Pentosan polysulfate is a semi-synthetic sulfated xylan biomimetic heparinoid that has been categorized as a disease modifying anti-arthritis drug (DMOAD). It has a smaller molecular weight

than heparan sulfate (HS) or heparin but has a higher charge density and has many properties that mimic HS found on cell surfaces and in extracellular matrix heparan sulfate proteoglycans (HSPGs) (Figure 2). This provides PPS with a multifunctional cell and tissue protective profile that is discussed more fully later in this review^{12,13}.

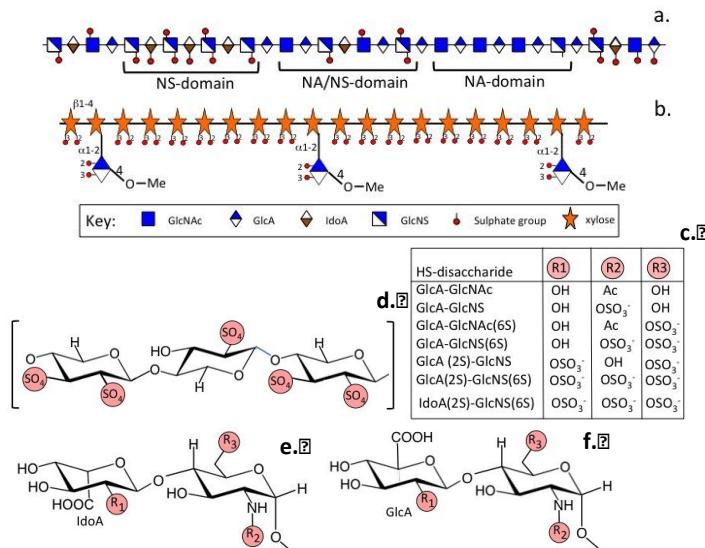


Figure 2. Sulfation patterns of heparan sulfate and pentosan polysulfate. Structural organization of a putative HS chain (a) and pentosan polysulfate (PPS) with its 4-O-methyl-glucuronic acid monosaccharide side chain α 1-2 linked to every tenth xylose residue (b), disaccharides HS is assembled from (c), saccharide structure of PPS (d), and two examples of HS disaccharides (e, f). Heparan sulfate contains three substitution sites in the D-glucuronic acid 1-4 linked N-acetyl glucosamine repeat disaccharide, marked R1, R2, R3 on HS can be occupied by combinations of hydroxyl, acetyl, and sulfate groups.

2. The impact of Coronaviruses on human health and well-being

Coronaviruses (CoVs) are enveloped viruses of the *Nidovirales* order, *Coronaviridae* family¹⁴. Bats, dogs, cats and humans can all be infected with these viruses¹⁴. Seven species of CoVs have so far been identified, four of these produce relatively mild symptoms of the common cold¹⁵ but severe acute respiratory syndrome (SARS-CoV), Middle East respiratory syndrome (MERS-CoV) and SARS-CoV-2 have a higher impact and can be life-threatening diseases¹⁶. The SARS-CoV pandemic of 2002–2003 resulted in 774 deaths and 8098 cases of infection distributed over 26 countries¹⁷. Middle East respiratory syndrome coronavirus (MERS-CoV) emerged ten years later as the sixth coronavirus and resulted in infections across 27 countries in the Middle East, Asia, North Africa and Europe which resulted in 2040 infections and 712 deaths. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the seventh coronavirus (CoV) which has recently emerged resulting in a global health pandemic. Severe acute respiratory syndrome coronavirus 2 is closely related to Severe acute respiratory syndrome coronavirus (SARS-CoV) but has a far more infectious profile and thus has a significantly greater impact on global human health. As of 19th Sept 2024 704 million confirmed cases of SARS-CoV-2 infection in 223 countries and territories, 7 million deaths and 675 million cases of recovery from infection have been reported by Worldometer [<https://www.worldometers.info/coronavirus/> accessed on 19-9-2024]¹⁸

2.1 PREVENTION OF VIRAL INFECTION OF HOST CELLS BY PENTOSAN POLYSULFATE

To infect human cells, viruses must pass a dense layer of carbohydrate (glycocalyx) attached to the cell surface. Several viruses, including Herpes, HIV and other coronaviruses, bind to HS during this infection phase. HS and ACE2 are necessary for SARS-CoV-2 infection and Nrp-1 has additional roles to play as a co-receptor in this infective process¹⁹⁻²⁴. Molecular modeling, atomic force

measurements and X-ray crystallography have shown that SARS-CoV-2 HS binds the Spike S1 receptor binding domain (RBD). Infection can be prevented by enzymatic removal of HS from the cell surface, demonstrating the importance of HS in the initial viral attachment phase. Exogenous heparin, LMW heparin and PPS block coronavirus infections in lab-grown cells by binding to the isolated SARS-CoV-2 viral particles dispersed in biological fluids and this prevents them from entering and infecting host cells. A GAG-binding site in the N-terminal domain (NTD) of Spike protein in residues 241–246²⁵ binds HS. Prophylactic administration of HS oligosaccharides also bind to this site and prevent productive associations between Spike protein and ACE2 showing how PPS blocks SARS CoV-2 infection²⁵. Molecular dynamic simulations of the Spike trimer interaction with HS dodecasaccharides (and PPS) indicate that when attached to this HS binding site these GAG components span the RRAR (CendR) furin cleavage site interfering with Spike protein interactions with cell surface receptors essential for SARS-CoV-2 infection²⁶. Use of heparin in-vitro can block this infective process in epithelial cells²⁷. Polysulfates block SARS-CoV-2 uptake into cells showing electrostatic interactions are important in this process²⁸ explaining why PPS inhibits SARS-CoV-2 infection of host cells. Biochemical, biophysical, and genetic studies show HS induces an open conformation in S protein required for binding to ACE2²⁹, a high degree of coordination between host cell HS and S protein asparagine-linked glycans enables ACE2 binding and host cell infection²⁹. Prophylactic use of PPS disrupts this interaction with cell surface HS and prevents host cell infection. SARS-CoV-2 Spike protein is proteolytically processed by transmembrane protease, serine 2 (TMPRSS2) and furin produced by host cells to prime the S1 domain for binding to ACE 2^{21,24,30-35}. Transmembrane protease, serine 2 (TMPRSS2), a respiratory and gastrointestinal membrane anchored protease, plays a crucial role in the activation of SARS-CoV-2 spike protein. Endogenous serine protease inhibitory proteins in

tissues may have a protective role to play in the prevention of this protease mediated remodeling of SARS CoV-2 Spike protein required to facilitate its interaction with ACE2. Small drug inhibitors (Camostat, Nafamostat, and Bromhexine) have been re-purposed from anti-tumor applications to inhibit TMPRSS2 mediated SARS-CoV-2 S protein priming³⁶. Entry of SARS-CoV-2 into host cells via the receptor binding domain (RBD) of S protein after the S1 and S2 subunits dissociate from each other by the action of TMPRSS2 allow conformational rearrangement and prime it for interaction with the host cell³⁷⁻³⁹. Furin, a type 1 membrane protease, also cleaves between S1 and S2 in SARS-CoV-2 S protein to facilitate binding to ACE2 and viral membrane fusion with the host cell plasma membrane, effecting internalisation of SARS-CoV-2³⁶. Transmembrane protease, serine 2 is part of a mucous secretory network highly upregulated in inflammation by interleukin-13. Interleukin-13 (IL13) and viral infection also mediate effects on ACE2 expression in the airway epithelium with interferon mediated responses to respiratory viruses highly upregulating ACE2 expression⁴⁰. Moreover, some viruses synthesize their own TMPRSS2⁴¹ and this also has roles in viral activation^{42,43}. While ACE2 is considered to be the primary host receptor in SARS-CoV-2 and SARS-CoV infections these related viruses have vastly different infection rates, suggesting the involvement of factors in addition to ACE2 that promote SARS-CoV-2 infection. Severe acute respiratory syndrome coronavirus-1 (SARS CoV-1) and SARS CoV-2 both bind to the ACE-2 receptor on host cells, however the latter is considerably more infectious, utilising multiple factors to achieve this higher infection rate (Table 1). Severe acute respiratory syndrome coronavirus-2 is particularly good at infecting cells, in the upper respiratory tract, and deeper in the lungs⁴⁴. Neuropilin-1 (NRP-1) is another host cell co-receptor that SARS-CoV-2 also uses for cellular attachment³⁵. Furin generates a C-end rule motif (CendR) in the SARS-CoV-2 spike protein and this

interacts with a CendR receptor in Nrp-1 promoting the internalisation of CoV-2 viral particles by endocytosis^{37-39,45} (Table I). The greater infective efficiency of the Omicron CoV-2 variant suggests it utilises these additional cell surface motifs to infect host cells. Cell surface HS is used by many viruses as a docking module in host cell infection (Table 2). Heparan sulfate proteoglycans (HSPGs) are endocytic receptors that viruses use for cell entry⁴⁶⁻⁴⁸. NRP-1 is also an endocytic receptor⁴⁹. Herpes simplex, hepatitis, papilloma, flaviviruses and respiratory syncytial virus all utilize multiple cell surface receptors as part of their internalization strategy to infect host cells⁵⁰⁻⁵³. Viruses do not bind to the non-sulfated HA component of the glycocalyx, and this acts as a barrier to viral penetration⁵⁴⁻⁵⁸. It has been proposed that COVID-19 is an endothelial disease⁵⁹ brought on by the cytokine storm of ARDS that produces destructive changes in the endothelial cell glycocalyx^{60,61}. Significantly, binding of viral particles to PPS in biological fluids prevents them from interacting with the HS chains of syndecan (SDC) or glycan (GPC) to gain access to the Nrp-1 or ACE2 receptors on host cells. As shown in Table II, many human and animal viruses utilize cell surface HS as a docking module to facilitate infection of host cells.

Table I ACE2, cell surface HSPGs and Nrp-1 interact with SARS CoV-2 Spike protein facilitating viral entry to host cells.

Receptor	Physiological properties	Evidence for roles as a SARS CoV-2 receptor	Ref
HS	Cell-ECM signaling Cell adhesion Cell growth factor and cytokine interactions	Direct interaction of HS with S glycoprotein in ECM, GAG microarray, co-precipitation experiments. Enzymatic removal of HS or HS knockdown results in reduced SARS CoV-2 infection levels.	62-64
ACE2	Regulation of blood pressure	Cryo EM images/ X ray crystallography demonstrate ACE2 bound to S RBD. ACE2 over-expression in cells results in enhanced CoV-2 infection. Human ACE2 over-expression in mice results in enhanced CoV-2 infection. Inhibition of SARS CoV-2 infection is evident in ACE2 knockout cells	65-70
Nrp-1	Regulation of neural network development and angiogenesis in tissue development	Demonstration of binding of Nrp-1 to Furin generated C-end rule (CendR) motif in Spike protein. Overexpression of Nrp-1 in cells results in enhanced SARS CoV-2 infection. Nrp-1 KO results in a reduced SARS CoV-2 infection	37,39,71,72

Viruses utilize cell surface syndecan and glycan HS-proteoglycan as docking structures as part of the infective process of prospective host (Table II).

These proteoglycans have a ubiquitous cellular distribution.

Table II Cell surface HS Proteoglycans that act as viral receptors

HSPG receptor	Viruses	Ref
Syndecan-1	Hepatitis C virus	73
Syndecan-2	Hepatitis B virus, Dengue virus strain DEN2 16681	74,75
Syndecan-3	HIV-1	76
Syndecan-4	Adeno-Associated Virus 9, Porcine reproductive and respiratory syndrome virus	77,78
Glycan-5	Hepatitis B and D viruses	79
Syndecans and glycans	Porcine hemagglutinating encephalomyelitis virus, Papilloma viruses.	80-82
Syndecan	SARS-CoV-2	83
Syndecans	HIV-1	84-86

2.2 CELL SURFACE GLYCOSAMINOGLYCANS AND VIRAL INFECTION OF HOST CELLS.

2.2.1 Anionic anti-viral compounds

Anionic polysulfate GAGs have inhibitory effects on host cell infection with multiple viruses including SARS-CoV-2⁸⁷ and AIDS; PPS has been proposed as a drug for the prevention of infection with AIDS

and SARS-CoV-2⁸⁸. It has been proposed that these compounds should be administered as aerosols inhaled into lung tissues to increase their potency⁸⁹. Administration of sulfated hyaluronan derivatives delivered by aerosol prolong the survival of K18 ACE2 mice infected with a lethal dose of SARS-CoV-2⁹⁰. PPS (SP 54), a low molecular weight sulfated polysaccharide is one of the most

active *in vitro* inhibitors of retrovirus-specific reverse transcriptase⁹⁰ and is a selective anti-HIV and anti-SARS-CoV-2 agent *in vitro*^{91,92}. Polysulfated polyxyran (HOE/BAY 946) completely inhibited syncytium formation induced by HIV infection of T-lymphocytes as well as viral replication and inhibited HIV reverse transcriptase. Furthermore, a drastic decrease in the release of viral particles in HIV infected U937 pro-monocytic cells was also elicited by HOE/BAY 946⁹³, this increases membrane hydrophobicity of human lymphocytes and specifically suppresses HIV-protein synthesis⁹⁴, and also inhibits HIV replication in human monocytes/macrophages⁹⁵. The pharmaco-kinetics of intravenous HOE/BAY 946 has been examined in HIV patients⁹⁶. Sulfated polysaccharides have also been shown to inhibit lymphocyte-to-epithelial transmission by HIV-1⁹⁷. Chemically oversulfated galactosaminoglycan sulfates inhibit the enveloped viruses HIV-1, HSV-1 and HCMV⁹⁸. Chondroitin polysulfate displays anti-HIV-1 activity *in vitro*⁹⁹. A synthetic polysulfonated naphthalene polymer (PRO 2000) binds to HIV-1 gp120 glycoprotein and interferes with viral binding to CD4⁺ T cells but also interacts with CD4 and CXCR4, a G-protein coupled chemokine receptor that can induce expression of selective chemokines with potential anti-viral activity^{100,101}, it also inhibits infection of host cells with HIV and SARS-CoV-2¹⁰⁰. Pentosan polysulfate also ameliorates the symptoms of human T lymphotropic virus type I (HTLV-I)-associated myelopathy/tropical spastic paraparesis (HAM/TSP) which is characterized by lower extremity motor dysfunction¹⁰².

2.3 SEVERE ACUTE RESPIRATORY SYNDROME VIRUS-2 VARIANTS

A highly virulent Delta SARS-CoV-2 variant (B.1.617.2) emerged in India in 2020 becoming the dominant global strain. On 24 November 2021, a further highly infectious SARS-CoV-2 variant (B.1.1.529/BA.1) was reported, this has also had a significant global impact displacing the delta variant as the dominant SARS Cov-2 strain¹⁰³. The World Health Organization Technical Advisory Group on SARS-CoV-2 Viral Evolution designated

this emergent CoV variant as B.1.1.529, the fifth Coronavirus variant and named it Omicron¹⁰⁴. Several Omicron variants have emerged since then with the BA-4 and BA-5 variants becoming firmly established. Vaccines raised to the original Wuhan strain of SARS CoV-2 offer incomplete coverage of these variants and multiple COVID-19 re-infections two or three times have been reported. This emphasizes the need to develop alternative preventative strategies to prevent COVID-19 infections rather than vaccines or antibodies that treat the symptoms. It is not known to what extent all of the symptoms of long COVID disease are treatable or whether full recovery is possible, however besides acting as a viral anti-infective agent, PPS also treats COVID-19 disease symptomatology¹⁰⁵.

The BA-4 and BA-5 Omicron variants are the most infectious forms of SARS-CoV-2 and are of major concern; their greater infectivity is related to 32 mutations in their S protein compared to the original Wuhan CoV-2 strain, 15 of these mutations specifically affect the CoV-2 RBD of S protein¹⁰⁶ (Figure 2). The high infective rate of the Omicron variants suggest these utilise a more effective range of cell surface binding motifs in addition to the ACE2 receptor and Nrp-1. A further Omicron sub-variant, a so-called second generation sub-variant, BA.2.75 has emerged in India, unofficially named Centaurus¹⁰⁷⁻¹¹⁰ and has been detected in Germany, The Netherlands, Japan, UK, US, Australia and New Zealand.

Two mutations in the BA.2.75 variant (G446S and R493Q) allow it to escape immune detection and to bind more strongly to the ACE2 receptor, it is predicted that this increases its infectivity. Prior COVID-19 immunizations may limit the infectiousness of this new sub-variant however it is not known how effective pre-existing antibody preparations will be against this new Omicron variant.

2.4 MUTATIONS IN S1 SPIKE GLYCOPROTEIN IN SEVERE ACUTE RESPIRATORY SYNDROME VIRUS-2 VARIANTS

Examination of amino acid sequences in the S1 glycoprotein of SARS-CoV-2 variants demonstrates

significant substitutions of native SARS-CoV-2 sequence which partly explains the waning effectiveness of vaccines and therapeutic antibodies in the treatment of COVID-19 disease and the

evasion of immune detection of these variant forms of SARS-CoV-2. Of the viral strains of SARS-CoV-2 so far identified, the Omicron strain has the highest number of S1 RBD substitutions (Figure 3).

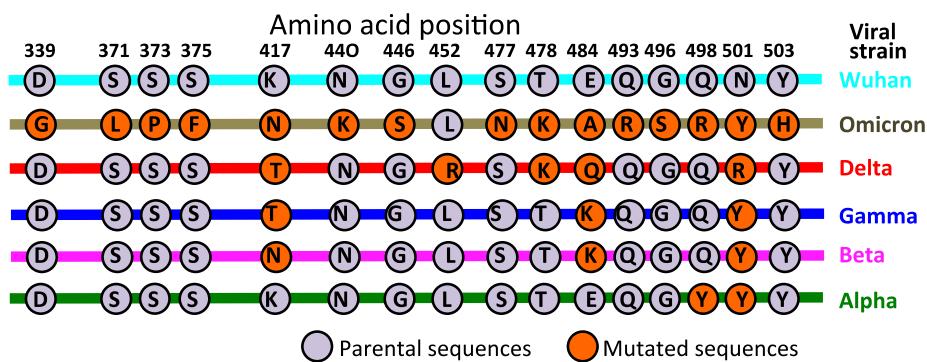


Figure 3. Point mutations in the receptor binding domain of spike protein in coronavirus variants. Amino acid sequences in the receptor binding domain of the SARS CoV-2 Spike protein in the original Wuhan strain and the mutations in its variant forms. Figure from¹¹¹.

Table III. A. Viruses that gain access to cells through interaction with cell surface HS and B. antiviral sulfated polysaccharides that block such viral interactions

Virus	Docking module	Ref
Viruses that utilize HS or related GAGs for infection of host cells A		
Adeno-associated virus 2	HS	112-114
Adeno associated virus serotype 3B	HS	115
Akabane and Schmallenberg Viruses.	Sulfated GAGs	116
Chikungunya Virus Strains	N- and 6-O-sulfated HS	117-119
Coxsackievirus B3 variant , Coxsackievirus A16, B4	HS	120-122
Dengue Viruses	HS	123
Duck Tembus virus	HS	124
Ebola virus	HS	125
Echovirus 5	HS	126
Enterovirus A71	HS	118,126,127
Filovirus	HS	128
Henipavirus	HS	129
Hepatitis B virus	HSPG	130
Hepatitis delta virus	HSPG	131
Hepatitis C	HS, HS-proteoglycans	132,133
Human herpes virus 8	HS	134
Herpes simplex virus type 1	HS	135
Human meta pneumo virus	HS	136
Herpes simplex virus type 1	Syndecan-1	137,138
HIV	HS	139-141
Human, monkey, rodent Foamy virus	HS	142,143
Human papillomavirus 16 virus	HS	144
Human respiratory syncytial virus	HS	145
Human meta pneumovirus	HS	146
Human Parechovirus	HS	117
Japanese encephalitis virus	HS	147

Virus	Docking module	Ref
Merkel cell polyomavirus	HS, sialylated glycans	148
Murine leukemia virus	HS	149
Murine herpes virus 68	HS	150
Moloney Murine leukemia virus	HS	140
Porcine epidemic diarrhea virus	HS	151
Pseudorabies virus	HS	152
Rabies virus	HS	153
Respiratory syncytial virus G	Heparin	154,155
SARS CoV-2	HS, HSPGs	21,24,28,33,64,156
Swine vesicular disease virus	HS	157
Sindbis virus	HS	158
Swine fever virus	HS	159
Vaccinia virus	HS	160
Zika virus	HS	161
Inhibition of viral attachment to host cells using sulfated polysaccharides B		
Virus	Blocking polysaccharide	Ref
African swine fever virus	PPS and Sulfated polysaccharides	162
Herpes simplex, Cytomegalovirus, Vesicular stomatis virus, Sindbis virus HIV	PPS and Sulfated polysaccharides	163,164
T cell leukemis virus type-1	PPS	165
Chikungunya virus	PPS	166
Ross river virus	PPS	166,167
SARS-CoV-2	PPS, Polysulfates, heparin, enoxaparin	24,28,64,156,168

3. Anti-inflammatory and tissue protective properties of pentosan polysulfate

Pentosan polysulfate has anti-inflammatory properties in knee OA, reducing joint swelling and pain¹⁶⁹ and has reno-protective effects in kidney injury, nephrectomy and diabetic nephropathy¹⁷⁰. Pentosan polysulfate is also effective against arthritogenic alphaviruses such as Ross River virus (RRV) and chikungunya virus (CHIKV) which cause cartilage destruction, crippling pain and joint inflammation¹⁶⁶. Pentosan polysulfate increases production of the anti-inflammatory cytokine IL-10 and reduces production of proinflammatory cytokines, modulates growth factor signaling and lymphocyte activation and reduces inflammatory infiltrates in joint fluids in chikungunya infected mice¹⁷¹. Pentosan polysulfate has systemic and local anti-inflammatory activity in post-acute pulmonary inflammation in an influenza virus A induced pulmonary inflammation model¹⁷². The beneficial

effects of PPS are due to a combination of its anti-viral and anti-inflammatory properties¹⁷³. Pentosan polysulfate also supports tissue repair processes in the degenerate IVD¹³, representing part of its pleotropic tissue and cell protective properties¹⁰⁵.

3.1 THE HYPERCOAGULATIVE STATE OF COVID-19 IMPAIRS PLATELET FUNCTION AND TISSUE REPAIR RESPONSES, WEAKENING NORMAL LUNG FUNCTION

Corona virus-2 infected patients that develop a severe pro-inflammatory state are also frequently associated with a procoagulant endothelial phenotype¹⁷⁴ that produces an elevation in fibrinogen and D-dimer/fibrin(ogen) degradation products associated with systemic hypercoagulability¹⁷⁵. Fibrinogen D-dimer levels positively correlate with mortality rates in COVID-19 patients and lead to arterial thrombotic events including stroke, ischemia and microvascular thrombotic events in the pulmonary vascular beds¹⁷⁶. Heparan sulfate is a critical regulator of the

immunoreceptor tyrosine-based inhibition motif (ITIM) receptor G6b-B-R that regulates platelet production and activation¹⁷⁷. Binding of G6b-B-R to the HS side chains of perlecan and multivalent heparin inhibits platelet and megakaryocyte function by inducing downstream signaling via the protein tyrosine phosphatases Shp1 and Shp2. SARS-CoV-2 initiates programmed cell death in platelets¹⁷⁸ thus G6b-B-R has important roles to play maintaining platelet levels in wound healing responses¹⁷⁹. Perlecan's interaction with G6b and G6b-R regulates fibrotic changes in tissues produced by excessive levels of platelet activation¹⁸⁰. Perlecan HS also regulates cell adhesion, proliferation and growth factor signaling in tissue repair responses in tissue homeostasis and optimal tissue function, features mimicked by PPS^{180,181}.

3.2 A DYNAMIC BALANCE BETWEEN THE FIBRINOLYTIC AND COAGULATION SYSTEMS IS CRITICAL TO NORMAL LUNG FUNCTION AND HOMEOSTASIS.

The fibrinolytic and coagulation system are interconnected however in COVID-19 can be overwhelmed by a hypercoagulative state that prevails. Plasmin is a major clot dissolving fibrinolytic enzyme produced with elevated levels of tissue plasminogen activator (tPA) which in turn is regulated by plasminogen activator inhibitors-1 and -2 (PAI-1, PAI-2). Autopsies of COVID-19 fatalities show thrombosis, micro-angiopathy, haemorrhage and alveolar damage. The dyslipidemia displayed by COVID-19 patients results in abnormally high levels of low density lipoproteins (LDLs) and low levels of high density lipoproteins (HDLs) in serum.

3.3 LUNG HEPARAN SULFATE PROTEOGLYCANS AND THEIR CELL REGULATORY PROPERTIES

Cell surface HSPGs in the lung are growth factor coreceptors binding these through HS and core protein interactions¹⁸². Instructive interactions with growth factors, morphogens, chemokines and ECM components, regulate cell adhesion, proliferation, migration, and differentiation, regulating pathophysiological processes in tissue

development and repair, inflammation, infection, and tumor development^{182,183}. HS-proteoglycans in the lung have instructive roles critical to regulation of tissue development, organ structure, and the control of resident cell populations¹⁸⁴⁻¹⁸⁸. Pikachurin, agrin, perlecan are HSPG components of the lung interactome with essential roles in lung development, homeostasis and function and roles in tissue fibrosis in lung disease^{179,189}. Fragmentation of lung ECM components due to endogenous protease activity or by proteases produced by an influx of inflammatory cells in lung disease leads to the release of bioactive protein fragments (matricryptins, matrikines) which can regulate cell metabolism. Matrikines have been identified with tissue repair properties^{190,191}. While ACE2 is the primary receptor for SARS CoV-2 entry other cell surface and ECM proteins may also bind to the SARS CoV-2 spike RBD such as perlecan LG3 and may potentially enhance RBD-ACE2 interactions representing a potential therapeutic target¹⁹². Proteoglycans embedded in the vascular endothelial glycocalyx, regulate the activity of cytokines and inflammatory responses but are proteolytically cleaved in inflammatory diseases and modulate pathological inflammatory responses. Soluble forms of SDC-1, SDC-3 and BGN are anti-inflammatory, suppress proinflammatory cytokine expression and leukocyte migration, and induce autophagy of proinflammatory M1 macrophages. However, soluble versikine, SDC-2, mimecan and DCN are proinflammatory increasing inflammatory cytokine synthesis and leukocyte migration. This contrasts with SDC-4 and perlecan which have anti-inflammatory properties¹⁹³ promoting tissue repair¹⁹⁴. Glycans also regulate Hh and Wnt signaling in systemic inflammation. Collectively, vascular endothelial glycocalyx-derived SDC-1-4 ectodomains, BGN, versikine, mimecan, perlecan, GPC and DCN are thus of therapeutic potential in the regulation of cytokine and leukocyte responses in lung inflammatory diseases¹⁹⁵. Pentosan polysulfate down regulates the secretion of a range of inflammatory cytokines and has potent anti-oxidant

activity. Both of these properties exert protective properties on cells and preserves tissue function¹⁰⁵.

4. Depolymerisation of HA in COVID-19 disease.

4.1 CELL MIGRATION-INDUCING AND HYALURONAN-BINDING PROTEIN (CEMIP, KIAA1199) IS A DEAFNESS GENE LINKED WITH DEPOLYMERISATION OF HYALURONAN

KIAA1199 knockdown abolishes HA degradation by human skin fibroblasts, cellular transfection of KIAA1199 cDNA confers an ability to catabolize HA in an endo- β -N-acetylglucosaminidase-dependent manner¹⁹⁶. The enhanced degradation of HA that occurs in synovial fibroblasts in OA¹⁹⁷ and RA correlates with elevated KIAA1199 expression and can be abrogated by knockdown of KIAA1199¹⁹⁸.

Depolymerisation of HA in long COVID-19 disease is associated with loss of hearing. Auditory neuropathy (deafness) is caused by disruption of nerve impulses travelling from the inner ear to the brain. Viral infection with mumps, measles, meningitis, SARS-CoV-2 and cytomegalovirus can all result in hearing loss. COVID-19 does not cause a sudden hearing loss but irreversible hearing loss and tinnitus can develop as a complication of SARS-CoV-2 infection¹⁹⁹. KIAA1199, a deafness gene of unknown function, plays a central role in HA binding and depolymerization independently of CD44 and HYAL-1 and HYAL-2¹⁹⁶. Pentosan polysulfate stimulates HA production in a number of cell types²⁰⁰ and also inhibits hyaluronidase²⁰¹ helping to maintain a healthy glycocalyx.

4.2 ROLES FOR ENDOTHELIAL CELLS AND HYALURONAN IN TISSUE MORPHOGENESIS AND EXTRACELLULAR MATRIX REPAIR

Hyaluronan promotes proliferation and migration of many cell types, and has important roles in tissue morphogenesis, wound healing, inflammation, angiogenesis, and tissue repair processes²⁰². Endothelial cells are responsive to HA oligosaccharides which stimulate proliferation, migration, new vessel formation and tissue repair responses²⁰³⁻²⁰⁶. Pulmonary stromal fibroblasts and

myofibroblasts synthesise HA contributing to the deposition of HA in the endothelial glycocalyx²⁰⁷, COVID-19 has been proposed to be an endothelial cell dysfunction disease. Angiotensin converting enzyme is highly expressed by endothelial cells, ACE2 has critical roles that impact on the progression of COVID-19 disease²⁰⁸⁻²¹¹.

5. A summation of the pleotropic cell and tissue protective properties of Pentosan polysulfate

Supplementary Figure 1 summarises the major changes that have been documented in COVID-19 and studies which have utilized PPS to treat the multiple symptoms which arise from viral infection^{105,212}. Besides having the ability to prevent attachment of a large range of viruses to host cells which occur through cell surface HS interactions (Table II, Table III) PPS also has many cell and tissue protective properties. These include application in the treatment of cystitis and painful bowel disease²¹³⁻²¹⁹, as a tissue protective enzyme inhibitor²²⁰⁻²²³, promotion of cartilage and IVD repair²²⁴⁻²²⁷, healing of OA cartilage and the degenerate IVD^{221,223,228,229}. PPS has been used in bioscaffolds in tissue engineering applications²³⁰⁻²³². PPS regulates Complement activation^{233,234}, coagulation/fibrinolysis²³⁵⁻²³⁸, thrombocytopenia^{239,240} and induces HA production by many cell types^{200,241}. Pentosan polysulfate inhibits NGF production by osteocytes which reduces bone pain in OA/RA²⁴² and promotes lipid removal from subchondral blood vessels engorged with lipid in OA/RA reducing pain in these conditions²⁴³. Regulation of cytokine and inflammatory mediator production by PPS in ARDS reduces inflammation in tissues. PPS also has anti-viral activity^{162,166,167} and is an anti-tumor agent in a number of cancers²⁴⁴.

5.1 PENTOSAN POLYSULFATE AND THE GUT MICROBIOME

The gut microbiome is disturbed in COVID-19 disease, with alterations in cell populations and imbalance in beneficial symbionts and opportunistic pathogens^{245,246}. Xylan is the second most

abundant plant carbohydrate biomass found in nature. Accumulated evidence shows that xylans interact with gut microbiota in a beneficial way²⁴⁷. Humans cannot digest xylans but they act as bulking material aiding in the throughput of digested food items through the gut. The gut microbiome produce a number of xylanolytic enzymes that allow the gut microbiome to utilize xylans as a nutrient source, the generated xylo-oligosaccharides have pre-biotic properties that aid in gut homeostasis²⁴⁸ countering the gut dysbiosis that occurs in COVID-19 disease²⁴⁹. Endoxylanases produced by the gut microbiota generate xylo-oligosaccharides (xylo-oligos)^{250,251} promoting beneficial symbiont microbes such as *Bifidobacterium* and *Lactobacillus* populations in the human gut improving mucosal health and immune function²⁵² and inhibit colonization of the gut by pro-inflammatory bacteria such as *Salmonella* sp. This improves gut barrier properties, and plasma lipid levels attenuating pro-inflammatory effects of a high fat diet and decreases blood LPS levels and the damaging effects of IL-1 β and IL-13.

6. Multi-organ involvement in Severe acute respiratory syndrome coronavirus 2 infection

SARS-CoV-2 is implicated in the clinical pathology of multiple organs and organ systems (Figure 4). Severe acute respiratory syndrome coronavirus 2 canonical mediators, ACE2, and TMPRSS2 are assisted by other coronavirus-associated receptors and factors, including basigin (BSG/CD147), dipeptidyl peptidase-4 (DPP4/CD26), cathepsin B/L, furin, interferon-induced transmembrane protein (IFITM1-3) and Nrp-1. The localization of these SARS-CoV-2 receptors, proteases, and genes involved in coding proteins that drive viral pathogenesis predisposes to SARS-CoV-2 infection in a number of tissues²⁵³, COVID-19 infection thus involves the hACE2 receptor and its co-receptors Nrp-1 and DPP4/CD26 which engage with the SARS CoV-2 spike protein²⁵⁴. In-silico

development of a bispecific antibody against SARS CoV-2 spike glycoprotein and DPP4 receptors (Regdanvimab and Begelomab) has been shown to block the D614G mutated spike glycoprotein of SARS-CoV-2 variants and host DPP4 receptor, respectively. This demonstrates the co-involvement of SARS CoV-2 S protein, hACE2 and DPP4/CD26 in the infective process in multi-organ viral infection^{254,255}.

6.1 HEMOLYSIS IN COVID 19 INFECTED LUNG TISSUES

Hemolysis is a common feature of COVID-19 infected tissues²⁵⁶, fibrotic changes in tissues also occurs resulting in a reduction in tissue elastic properties and lung function²⁵⁷. Pro-coagulant activity also results in thrombus formations in tissues impairing their functional properties^{258,259}. This leads to further detrimental effects on these tissues with free heme release resulting in oxidative stress, local generation of oxygen free radicals and mitochondrial and ER distress, leukocyte recruitment, vascular permeabilization, platelet and Complement activation, thrombosis, and fibrosis leading to impaired lung function. Platelets initiate blood clotting, severely affected COVID-19 patients display a high incidence of hypercoagulation in the lungs and brain. Plasma fibrinogen levels are also elevated with advancing age, high cholesterol, being female, menopause, obesity, smoking, inactivity and stress. Most of these features are putative risk factors for COVID-19^{260,261}. Heparin treatment of COVID-19 patients displaying enhanced coagulation levels results in an improved prognosis however heparin will only prevent thrombus formation and will not dissolve existing fibrin clots, thus is palliative and not curative. Prevention of SARS-CoV-2 infection of host cells by PPS represents a more effective treatment strategy and has the added advantage of minimizing inflammatory cytokine production and exacerbation of inflammatory conditions in tissues²⁶². Heme is a prosthetic group with functional roles in a wide variety of heme proteins such as hemoglobin and the cytochromes. Release of free heme in injured lung tissues promotes

adhesion molecule expression, leukocyte recruitment, vascular permeabilization, platelet activation, complement activation and thrombosis. Heme, however, can be degraded by the anti-inflammatory enzyme heme oxygenase (HO-1) generating biliverdin/bilirubin, iron/ferritin and carbon monoxide²⁶³. Free heme promotes lung inflammation in critically ill COVID-19 patients. Heme oxygenase -1 has anti-oxidative and anti-inflammatory properties and may represent a specific means of targeting hemolysis therapeutically in COVID-19 disease²⁶⁴.

6.2 COVID AND COGNITIVE DECLINE

COVID-19 infected patients frequently exhibit neurological symptoms of anosmia and fatigue and long-term neurological deficits post-infection such as cognitive decline and brain-fogging^{168,265,266}. Positron emission tomography (PET) and SPECT (Single-photon emission computed tomography) molecular imaging techniques have been used to shed light on how COVID-19 affects human brain structure²⁶⁷. Human brain structure is affected by long COVID-19 disease even after recovery of respiratory function and has been referred to as Post COVID Syndrome^{268,269}. It is not known how long such neurological deficits will persist in cases of severe SARS CoV-2 infection following recovery of respiratory function²⁷⁰ however reports of a reduction in IQ and altered immune regulation in young children effected by even very mild COVID-19 respiratory disease are particularly concerning^{271,272}. Long-term CNS neuroinflammation following COVID-19 infection in children may deleteriously affect brain development²⁷³. Disturbing reports are emerging of learning difficulties and a decline in the educational status of 9 year olds affected by COVID-19, an effect which may be exacerbated in individuals who also display underlying neurological deficits²⁷⁴⁻²⁷⁶.

6.2.1 The impact of COVID-19 on patients suffering from neurological deficits

The COVID-19 pandemic has disproportionately impacted patients suffering from AD and dementia

who have a reduced capacity to understand and comply with pandemic health care restrictions and may represent a spreader risk for COVID-19²⁷⁷. Present day AD/dementia patient numbers of 47 million are projected to triple by 2050 and this will be further compounded by the impact of the COVID-19 pandemic. It is thus predicted that neurological disorders will likely make a greater impact on general health even in patients who have only been impacted by mild symptoms of COVID-19. Cognitive deficits have been reported in patients after recovery from COVID-19 respiratory disease. An inability to concentrate and a fogging of thought processes, impaired concentration and problem-solving capability coupled with feelings of long-term anxiety and insecurity have all been reported²⁷⁸⁻²⁸². Anecdotal reports of COVID-19 infection resulting in a reduction in IQ in children is particularly alarming. COVID-19 disease is often referred to as a mild disorder in children based on its relative impact on respiratory function however little regard is made of the potential long-term effects of COVID-19 disease on brain function. Long-term fatigue with COVID also impacts on the development of neuropsychiatric disorders²⁸³.

7. Conclusions

Use of PPS as a prophylactic that intercepts SARS Cov-2 virion particles in the glycocalyx prevents their binding to cell surface HS in all viral strains and is not impeded by point mutations arising from recombination as part of the natural viral life-cycle. SARS-CoV-2 possesses 24 spike glycoproteins per virion particle which have central roles in binding to cell surface ACE2 facilitating viral entry into host cells. This occurs through the RBD of spike protein however this is buried within the S1 domain which is exposed by a conformational change upon interaction with cell surface HS. PPS prevents such HS interactions occurring and viral infection and warrant further investigation. PPS is effective against all classes of viruses and its anti-viral properties are not diminished by viral mutations. The emergence of a further bat coronavirus, HKU5-CoV-2 related to SARS CoV2^{284,285} and of a mink

respiratory coronavirus (MRCoV)²⁸⁶ related to MERS and SARS CoV2 indicates that due diligence is essential. PPS would be expected to be an effective blocking agent for these new CoV strains, however vaccines or antibodies have yet to be developed. It may thus be a prime time to adopt PPS in preventative anti-viral strategies.

Disclosures

JM has received consultancy fees from Arthropharm-Sylvan Pharmaceutical Ltd. MMS is clinical research director at Arthropharm-Sylvan Pharmaceutical Ltd. The authors have no conflicts to report.

Funding Statement:

None.

Acknowledgements:

This study was funded by the Melrose Personal Research Fund, Sydney, Australia. The contributions of the many scientists who have characterized the properties of PPS in laboratory studies and pre-clinical trials and whose work is reviewed in this manuscript is acknowledged.

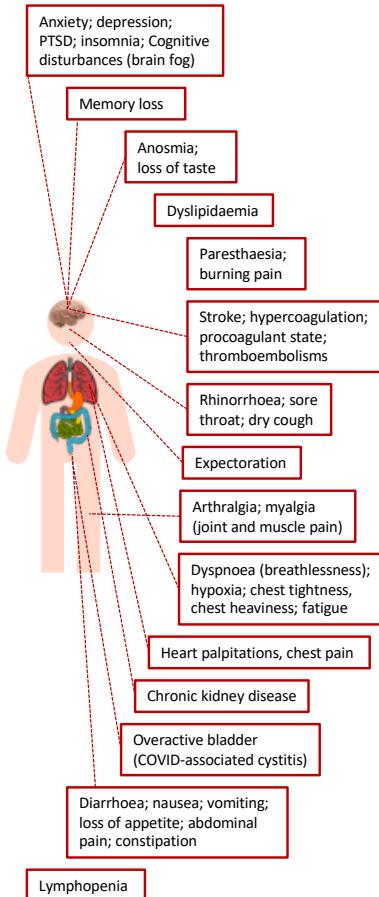
Author contributions

JM and MMS shared in conception of the study and shared in the writing and editing of this manuscript. JM and MMS both shared in the preparation of the figures. Both authors accepted the final version of the manuscript.

Supplemental Figure

Why use pentosan polysulfate (PPS) for long COVID?

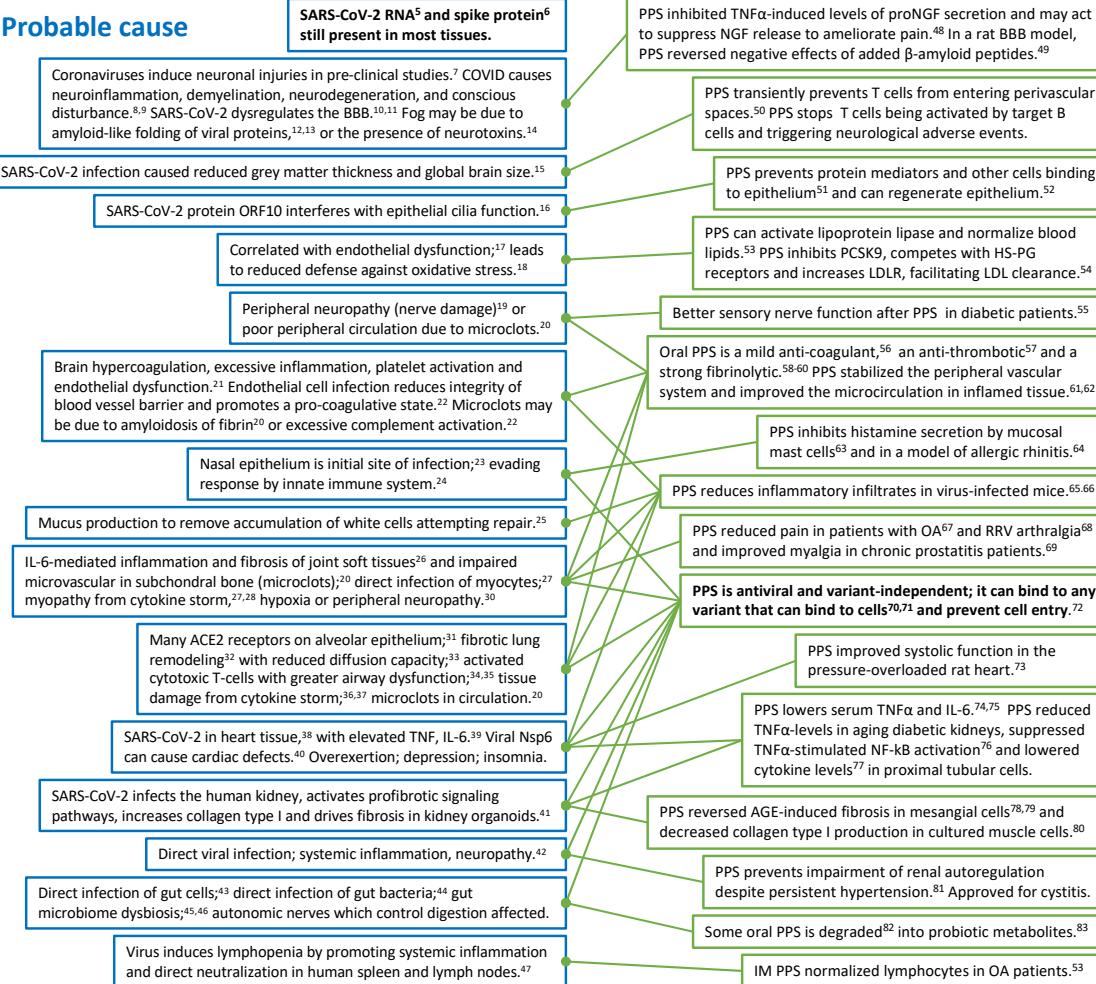
Symptom



Probable cause

PPS is a semisynthetic sulfated xylan manufactured from beechwood hemicellulose by bene-Arzneimittel GmbH in Germany.^{1,2} Approved in Australia, USA and Europe (Elmiron® capsules) for oral use as a treatment for people with interstitial cystitis. Safe and non-toxic,^{3,4} no SAE or deaths after millions of oral doses.

PPS action



Supplementary Figure 1. Illustration of how the multifunctional properties of pentosan polysulfate (PPS) can be used to treat COVID-19 infected tissues.

References

1. Degenhardt M, Benend H, Wätzig H. Quality control of pentosane polysulfate by capillary zone electrophoresis using indirect detection. *Journal of chromatography* 1998; **817**(1-2): 297-306.
2. Degenhardt M, Ghosh P, Wätzig H. Studies on the structural variations of pentosan polysulfate sodium (NaPPS) from different sources by capillary electrophoresis. *Arch Pharm (Weinheim)* 2001; **334**(1): 27-9.
3. Davis EL, El Khoudary SR, Talbott EO, Davis J, Regan LJ. Safety and efficacy of the use of intravesical and oral pentosan polysulfate sodium for interstitial cystitis: a randomized double-blind clinical trial. *J Urol* 2008; **179**: 177-85.
4. Kasyan G, Kupriyanov Y, Karasev A, Baibarin K, Pushkar D. Safety and efficacy of pentosan polysulfate in patients with bladder pain syndrome/interstitial cystitis: a multicenter, double-blind, placebo-controlled, randomized study. *Cent Eur J Urol* 2021; **74**: 201-7.
5. Chertow D, Stein S, Ramelli S, et al. SARS-CoV-2 infection and persistence throughout the human body and brain. Research Square: Research Square Platform LLC; 2021. p. unpublished.
6. Swank Z, Senussi Y, Alter G, Walt DR. Persistent circulating SARS-CoV-2 spike is associated with post-acute COVID-19 sequelae. MedRxiv: Cold Spring Harbor Laboratory; 2022.
7. Desforges M, Le Coupanec A, Dubeau P, et al. Human coronaviruses and other respiratory viruses: underestimated opportunistic pathogens of the central nervous system? *Viruses* 2019; **12**: 14.
8. Pallanti S, Grassi E, Makris N, Gasic GP, Hollander E. Neurocovid-19: a clinical neuroscience-based approach to reduce SARS-CoV-2-related mental health sequelae. *J Psychiatr Res* 2020; **130**: 215-7.
9. Fernández-Castañeda A, Lu P, Geraghty AC, et al. Mild respiratory SARS-CoV-2 infection can cause multi-lineage cellular dysregulation and myelin loss in the brain. *Cell* 2022.
10. Sasannejad C, Ely EW, Lahiri S. Long-term cognitive impairment after acute respiratory distress syndrome: a review of clinical impact and pathophysiological mechanisms. *Crit Care* 2019; **23**: 352.
11. Krasemann S, Haferkamp U, Pfefferle S, et al. The blood-brain barrier is dysregulated in COVID-19 and serves as a CNS entry route for SARS-CoV-2. *Stem Cell Rep* 2022; **17**: 307-20.
12. Nyström S, Hammarström P. Amyloidogenesis of SARS-CoV-2 spike protein. *J Am Chem Soc* 2021: in press.
13. Charnley M, Islam S, Bindra GK, et al. Neurotoxic amyloidogenic peptides in the proteome of SARS-CoV2: potential implications for neurological symptoms in COVID-19. *Nature Commun* 2022; **13**: 3387.
14. Cysique LA, Jakabek D, Bracken SG, et al. Post-acute COVID-19 cognitive impairment and decline uniquely associate with kynurenone pathway activation: a longitudinal observational study. MedRxiv: CSH Laboratory; 2022.
15. Douaud G, Lee S, Alfaro-Almagro F, et al. SARS-CoV-2 is associated with changes in brain structure in UK Biobank. *Nature* 2022; **604**: 697-707.
16. Wang L, Liu C, Yang B, et al. SARS-CoV-2 ORF10 impairs cilia by enhancing CUL2ZYG11B activity. *J Cell Biol* 2022; **221**: e202108015.
17. Charfeddine S, Ibn Hadj Amor H, Jdidi J, et al. Long COVID 19 syndrome: is it related to microcirculation and endothelial dysfunction? Insights from TUN-EndCOV study. *Front Cardiovasc Med* 2021; **8**: 745758.
18. Dei Cas M, Ottolenghi S, Morano C, et al. Link between serum lipid signature and prognostic factors in COVID-19 patients. *Sci Reports* 2021; **11**: 21633.
19. Novak P, Mukerji SS, Alabsi HS, et al. Multisystem involvement in post-acute sequelae of coronavirus disease-19. *Ann Neurol* 2022; **91**: 367-79.
20. Grobbelaar LM, Venter C, Vlok M, et al. SARS-CoV-2 spike protein S1 induces fibrin(ogen) resistant to fibrinolysis: implications for microclot formation in COVID-19. *Biosci Rep* 2021; **41**: BSR20210611.
21. Bikdeli B, Madhavan MV, Jimenez D, et al. COVID-19 and thrombotic or thromboembolic disease: implications for prevention, antithrombotic therapy, and follow-up. *J Am College Cardiol* 2020; **75**: 2950-73.
22. Jin Y, Ji W, Yang H, Chen S, Zhang W, Duan G. Endothelial activation and dysfunction in COVID-19: from basic mechanisms to potential therapeutic approaches. *Signal Transduct Target Ther* 2020; **5**: 293.
23. Diamond MS, Thirumala-Devi K. Innate immunity: the first line of defense against SARS-CoV-2. *Nature immunol* 2022; **23**: 165-76.
24. Hatton CF, Botting RA, Dueñas ME, et al. Delayed induction of type I and III interferons mediates nasal epithelial cell permissiveness to SARS-CoV-2. *Nature Commun* 2021; **12**: 7092.
25. Rendeiro AF, Ravichandran H, Bram Y, et al. The spatial landscape of lung pathology during COVID-19 progression. *Nature* 2021; **593**: 564-9.
26. Drożdżał S, Rosik J, Lechowicz K, et al. COVID-19: Pain management in patients with SARS-CoV-2 infection—molecular mechanisms, challenges, and perspectives. *Brain Sci* 2020; **10**: 465.
27. dos Santos PK, Sigoli E, Bragaña LJV, Cornachione AS. The musculoskeletal involvement after mild to moderate COVID-19 infection. *Front Physiol* 2022; **13**: 813924.
28. Pedersen SF, Ho Y-C. SARS-CoV-2: a storm is raging. *J Clin Invest* 2020; **130**: 2202-5.

29. Hejbol EK, Harbo T, Agergaard J, et al. Myopathy as a cause of fatigue in long-term post-COVID-19 symptoms: Evidence of skeletal muscle histopathology. *Eur J Neurol* 2022; in press.

30. Silva CC, Bichara CNC, Carneiro FRO, et al. Muscle dysfunction in the long coronavirus disease 2019 syndrome: Pathogenesis and clinical approach. *Rev Med Virol* 2022; **32**: e2355.

31. Zhao Y, Zhao Z, Wang Y, Zhou Y, Ma Y, Zuo W. Single-cell RNA expression profiling of ACE2, the putative receptor of Wuhan 2019-nCov. *Am J Resp Crit Care Med* 2020; **202**: 756-9.

32. Zhao Y-M, Shang Y-M, Song W-B, et al. Follow-up study of the pulmonary function and related physiological characteristics of COVID-19 survivors three months after recovery. *EClinicalMedicine* 2020; **25**: 100463.

33. Van Den Borst B, Peters JB, Brink M, et al. Comprehensive health assessment 3 months after recovery from acute coronavirus disease 2019 (COVID-19). *Clin Infect Dis* 2020; **73**: e1089-e98.

34. Vijayakumar B, Boustani K, Ogger PP, et al. Immuno-proteomic profiling reveals aberrant immune cell regulation in the airways of individuals with ongoing post-COVID-19 respiratory disease. *Immunity* 2022; **55**: 542-56.

35. Littlefield KM, Watson RO, Schneider JM, et al. SARS-CoV-2-specific T cells associate with inflammation and reduced lung function in pulmonary post-acute sequela of SARS-CoV-2. *PLOS Pathogens* 2022; **18**: e1010359.

36. Chau AS, Weber AG, Maria NI, et al. The longitudinal immune response to coronavirus disease 2019: chasing the cytokine storm. *Arthritis Rheumatol* 2021; **73**: 23-35.

37. Hadjadj J, Yatim N, Barnabei L, et al. Impaired type I interferon activity and inflammatory responses in severe COVID-19 patients. *Science* 2020; **369**: 718-24.

38. Lindner D, Fitzek A, Bräuninger H, et al. Association of cardiac infection with SARS-CoV-2 in confirmed COVID-19 autopsy cases. *JAMA Cardiol* 2020; **5**: 1281-5.

39. Bräuninger H, Stoffers B, Fitzek ADE, et al. Cardiac SARS-CoV-2 infection is associated with pro-inflammatory transcriptomic alterations within the heart. *Cardiovasc Res* 2022; **118**: 542-55.

40. Zhu J-Y, Wang G, Huang X, et al. SARS-CoV-2 Nsp6 causes cardiac defects through MGA/MAX complex-mediated increased glycolysis. Research Square Platform LLC; 2022.

41. Jansen J, Reimer KC, Nagai JS, et al. SARS-CoV-2 infects the human kidney and drives fibrosis in kidney organoids. *Cell Stem Cell* 2022; **29**: 217-31.e8.

42. Lamb LE, Timar R, Wills M, et al. Long COVID and COVID-19-associated cystitis (CAC). *Int Urol Nephrol* 2022; **54**: 17-21.

43. Liang W, Feng Z, Rao S, et al. Diarrhoea may be underestimated: a missing link in 2019 novel coronavirus. *Gut* 2020; **69**: 1141-3.

44. Brogna C, Brogna B, Bisaccia DR, et al. Could SARS-CoV-2 have bacteriophage behavior or induce the activity of other bacteriophages? *Vaccines* 2022; **10**: 708.

45. Wang B, Zhang L, Wang Y, et al. Alterations in microbiota of patients with COVID-19: potential mechanisms and therapeutic interventions. *Signal Transduct Target Ther* 2022; **7**: 143.

46. Xiao F, Tang M, Zheng X, Liu Y, Li X, Shan H. Evidence for gastrointestinal infection of SARS-CoV-2. *Gastroenterol* 2020; **158**: 1831-3.

47. Xiang Q, Feng Z, Diao B, et al. SARS-CoV-2 Induces Lymphocytopenia by Promoting Inflammation and Decimates Secondary Lymphoid Organs. *Frontiers in Immunology* 2021; **12**.

48. Stapledon CJM, Tsangari H, et al. Human osteocyte expression of Nerve Growth Factor: The effect of Pentosan Polysulphate Sodium (PPS) and implications for pain associated with knee osteoarthritis. *PLoS One* 2019; **14**: e0222602.

49. Deli MA, Veszelka S, Csizsar B, et al. Protection of the blood-brain barrier by pentosan against amyloid- β -Induced toxicity. *J Alzheimer's Dis* 2010; **22**: 777-94.

50. Klinger M, Zugmaier G, Nägele V, et al. Adhesion of T Cells to endothelial cells facilitates Blinatumomab-associated neurologic adverse events. *Cancer Res* 2020; **80**: 91-101.

51. Hueber AO, Pierres M, He HT. Sulfated glycans directly interact with mouse Thy-1 and negatively regulate Thy-1-mediated adhesion of thymocytes to thymic epithelial cells. *J Immunol* 1992; **148**: 3692-9.

52. Herbert JM, Floutard D, Paul R, Maffrand JP. Effect of pentosan polysulfate on endothelial regeneration. *Pathol Biol* 1989; **37**(7): 847-50.

53. Anderson JM, Edelman J, Ghosh P. Effects of pentosan polysulphate on peripheral blood leukocyte populations and mononuclear cell procoagulant activity in patients with osteoarthritis. *Curr Therap Res* 1997; **58**: 93-107.

54. Gustafsen C, Olsen D, Vilstrup J, et al. Heparan sulfate proteoglycans present PCSK9 to the LDL receptor. *Nature Communications* 2017; **8**: 503.

55. Laczy B, Cseh J, Mohás M, et al. Effects of pentoxifylline and pentosan polysulphate combination therapy on diabetic neuropathy in type 2 diabetes mellitus. *Acta Diabetol* 2009; **46**: 105-11.

56. Ghosh P, Smith MM, Wells C. Second-Line Agents in Osteoarthritis. In: Dixon JS, Furst DE, eds. *Second-Line Agents in the Treatment of Rheumatic Diseases: Present and Future*. New York, Basel, Hong Kong: Marcel Dekker; 1992: 383-428.

57. Marsh N, Peyser P, Gaffney P. Measurement of fibrinolysis in conscious and anesthetized rats - effects of pentosan polysulfate (SP54). *Thromb Haemost* 1985; **54**: 213.

58. Vinazzer H. Klinische untersuchungen zur fibrinolytischen wirkung von pentosanpolysulfat. *Folia Haematol* 1986; **113**: 272-7.

59. Sié P, Albareda JL, Robert M, et al. Tolerance and biological activity of pentosan polysulfate after intramuscular or subcutaneous administration for ten days in human volunteers. *Thromb Haemost* 1986; **55**: 86-9.

60. Vinazzer H. Effect of pentosan polysulphate on fibrinolysis: Basic tests and clinical application. *Semin Thromb Hemost* 1991; **17**: 375-8.

61. Kalbhen DA. Pharmacological studies on the anti-inflammatory effect of a semi-synthetic polysaccharide (pentosan polysulfate). *Pharmacol* 1973; **9**: 74-9.

62. Kalbhen DA. Biochemisch-pharmakologische Grundlagen zur antiphlogistisch-antirheumatischen Wirkung von Pentosanpolysulfat. *Wiener klin Wschr* 1978; **90**: 101-5.

63. Chiang G, Patra P, et al. Pentosanpolysulfate inhibits mast cell histamine secretion and intracellular calcium ion levels: an alternative explanation of its beneficial effect in interstitial cystitis. *J Urol* 2000; **164**: 2119-25.

64. Sanden C, Mori M, Jogdand P, et al. Broad Th2 neutralization and anti-inflammatory action of pentosan polysulfate sodium in experimental allergic rhinitis. *Immunity Inflam Dis* 2017; **5**: 300-9.

65. Herrero LJ, Foo S-S, et al. Pentosan polysulfate: a novel glycosaminoglycan-like molecule for the effective treatment of alphavirus-induced cartilage destruction and inflammatory disease. *J Virol* 2015; **89**: 8063-76.

66. Rudd PA, Lim EXY, Stapledon CJM, Krishnan R, Herrero LJ. Pentosan polysulfate sodium prevents functional decline in chikungunya infected mice by modulating growth factor signalling and lymphocyte activation. *PLoS ONE* 2021; **16**: e0255125.

67. Ghosh P, Edelman J, March L, Smith MM. Effects of pentosan polysulfate in osteoarthritis of the knee: A randomized, double-blind, placebo-controlled pilot study. *Curr Ther Res* 2005; **66**: 552-71.

68. Krishnan R, Duiker M, Rudd PA, et al. Pentosan polysulfate sodium for Ross River virus-induced arthralgia: a phase 2a, randomized, double-blind, placebo-controlled study. *BMC Musculoskeletal Disorders* 2021; **22**: 271.

69. Wedrén H. Effects of sodium pentosanpolysulphate on symptoms related to chronic non-bacterial prostatitis. *Scand J Urol Nephrol* 1987; **21**: 81-8.

70. Nie C, Pouyan P, Lauster D, et al. Polysulfates block SARS-CoV-2 uptake through electrostatic interactions. *Angew Chem Int Ed* 2021; **69**: 15870-8.

71. Ennemoser M, Rieger J, Muttenthaler E, Gerlza T, Zatloukal K, Kungl AJ. Enoxaparin and pentosan polysulfate bind to the SARS-CoV-2 spike protein and human ACE2 receptor, inhibiting Vero cell infection. *Biomedicines* 2022; **10**: 49.

72. Zhang F, He P, Rodrigues AL, et al. Potential anti-SARS-CoV-2 activity of pentosan polysulfate and mucopolysaccharide polysulfate. *Pharmaceutics* 2022; **15**: 258.

73. Vistnes M, Aronsen JM, Lunde IG, Sjaastad I, Carlson CR, Christensen G. Pentosan polysulfate decreases myocardial expression of the extracellular matrix enzyme ADAMTS4 and improves cardiac function in vivo in rats subjected to pressure overload by aortic banding. *PLoS One* 2014; **9**: e0089621.

74. Smith MM, Numata Y, Ghosh P. Effects of calcium pentosan polysulfate on joint inflammation and pouch fluid levels of leukocytes, nitric oxide, and interleukin-6 in a rat model of arthritis. *Curr Ther Res* 1999; **60**: 561-76.

75. Smith MM, Ghosh P, Numata Y, Bansal MK. The effects of orally administered calcium pentosan polysulfate on inflammation and cartilage degradation produced in rabbit joints by intraarticular injection of a hyaluronate-polylysine complex. *Arthritis Rheum* 1994; **37**: 125-36.

76. Wu J, Guan T-j, Zheng S, et al. Inhibition of inflammation by pentosan polysulfate impedes the development and progression of severe diabetic nephropathy in aging C57B6 mice. *Lab Invest* 2011; **91**: 1459-71.

77. Chen P, Yuan Y, Zhang T, Xu B, Gao Q, Guan T. Pentosan polysulfate ameliorates apoptosis and inflammation by suppressing activation of the p38 MAPK pathway in high glucosetreated HK2 cells. *Int J Mol Med* 2018; **41**: 908-14.

78. Elliot SJ, Striker LJ, Stetlers-Stevenson WG, Jacot TA, Striker GE. Pentosan polysulfate decreases proliferation and net extracellular matrix production in mouse mesangial cells. *J Am Soc Nephrol* 1999; **10**(1): 62-8.

79. Xiao L, Chen AL, Gao Q, Xu B, Guo X, Guan T. Pentosan polysulfate ameliorates fibrosis and inflammation markers in SV40 MES13 cells by suppressing activation of PI3K/AKT pathway via miR-446a-3p. *BMC nephrology* 2022; **23**: 105.

80. Elliot SJ, Zorn BH, McLeod DG, et al. Pentosan polysulfate decreases prostate smooth muscle proliferation and extracellular matrix turnover. *Prostate Cancer Prostatic Dis* 2003; **6**(2): 138-42.

81. Guan Z, Singletary ST, et al. Pentosan polysulfate preserves renal micro-vascular P2X1 receptor reactivity and autoregulatory behavior in DOCA-salt hypertensive rats. *Am J Physiol Renal Physiol* 2016; **310**: F456-F65.

82. Simon M, McClanahan RH, Shah JF, Repko T, Modi NB. Metabolism of [³H]pentosan polysulfate sodium (PPS) in healthy human volunteers. *Xenobiotica* 2005; **35**: 775-84.

83. Aachary AA, Prapulla SG. Xylooligosaccharides (XOS) as an emerging prebiotic: microbial synthesis, utilization, structural characterization, bioactive properties, and applications. *Compr Rev Food Sci Food Safety* 2011; **10**: 2-16.

References:

- 1 Johnston, C., Hughes, H, Lingard, S, Hailey, S, Healy, B. Immunity and infectivity in covid-19. *378*, e061402 (2022).
- 2 Yisimayi, A., et al., Repeated Omicron exposures override ancestral SARS-CoV-2 immune imprinting. *Nature* **625**, 148-156 (2024).
- 3 Cele, S., et al., Omicron extensively but incompletely escapes Pfizer BNT162b2 neutralization. *Nature* **602**, 654-656 (2022).
- 4 Kaku, Y., et al., Virological characteristics of the SARS-CoV-2 KP.2 variant. *Lancet Infect Dis* (2024).
- 5 Plante, J. A., et al., Spike mutation D614G alters SARS-CoV-2 fitness. *Nature* **592**, 116-121 (2021).
- 6 Lopes-Pacheco, M., Silva, PL, Cruz, FF, Battaglini, D, Robba, C, Pelosi, P, Morales, MM, Caruso Neves, C, Rocco, PRM. Pathogenesis of Multiple Organ Injury in COVID-19 and Potential Therapeutic Strategies. *Front Physiol* **12**, 593223 (2021).
- 7 Mokhtari, T., Hassani, F, Ghaffari, N, Ebrahimi, B, Yarahmadi, A, Hassanzadeh, G. COVID-19 and multiorgan failure: A narrative review on potential mechanisms. *J Mol Histo* **51**, 613-628 (2020).
- 8 Zaim, S., Chong, JH, Sankaranarayanan, V, Harky, A. COVID-19 and Multiorgan Response. *Curr Probl Cardiol* **45**, 100618 (2020).
- 9 Narayanan, S., Jamison, DA, Guarnieri, JW et al. A comprehensive SARS-CoV-2 and COVID-19 review, Part 2: host extracellular to systemic effects of SARS-CoV-2 infection. *Eur J Hum Genet* **32**, 10-20 (2024).
- 10 Tang, D., Comish ,P, Kang, R The hallmarks of COVID-19 disease. *PLoS Pathog* **16**, e1008536 (2020).
- 11 Menon, N., Mohapatra, S. The COVID-19 pandemic: Virus transmission and risk assessment. *Curr Opin Environ Sci Health* **28**, 100373 (2022).
- 12 Smith, M., Melrose, J. Pentosan Polysulfate Affords Pleotropic Protection to Multiple Cells and Tissues. *Pharmaceuticals (Basel)* **16**, 437 (2023).
- 13 Smith, M., Hayes, AJ, Melrose, J. Pentosan Polysulfate, a Semisynthetic Heparinoid Disease-Modifying Osteoarthritic Drug with Roles in Intervertebral Disc Repair Biology Emulating the Stem Cell Instructive and Tissue Reparative Properties of Heparan Sulfate. *Stem Cells Dev* **31**, 406-430. doi: 410.1089/scd.2022.0007 (2022).
- 14 Fehr, A., Perlman, S. Coronaviruses: an overview of their replication and pathogenesis. *Methoda Mol Biol* **1282**, 1-23 (2015).
- 15 Rabaan, A., Al-Ahmed, SH, Haque, S, Sah, R, Tiwari, R, Malik, YS, Dhama, K, Yatoo, MI, Bonilla-Aldana, DK, Rodriguez-Morales, AJ. SARS-CoV-2, SARS-CoV, and MERS-CoV: a comparative overview. *Infez Med* **28**, 174-184 (2020).
- 16 Rabaan, A., Al-Ahmed, SH, Haque, S, Sah, R, Tiwari, R, Malik, YS, Dhama, K, Yatoo, MI, Bonilla-Aldana, DK, Rodriguez-Morales, AJ. SARS-CoV-2, SARS-CoV, and MERS-CoV: a comparative overview. *Infez Med* **28**, 174-184 (2020).
- 17 Chafekar, A., Fielding, BC. MERS-CoV: understanding the latest human coronavirus threat. *Viruses* **10**, 93 (2018).
- 18 Worldometer. Coronavirus death toll. <https://http://www.worldometers.info/coronavirus/coronavirus-death-toll/> (2022).
- 19 Bhat, E., Khan, J, Sajjad, N, Ali, A, Aldakeel, FM, Mateen, A, Alqahtani, MS, Syed, R. SARS-CoV-2: Insight in genome structure, pathogenesis and viral receptor binding analysis - An updated review. *Int Immunopharmacol* **95**, 107493 (2021).
- 20 Guney, C., Akar, F. Epithelial and Endothelial Expressions of ACE2: SARS-CoV-2 Entry Routes. *J Pharm Pharm Sci* **24**, 84-93 (2021).
- 21 Kim, S., Jin, W, Sood, A, Montgomery, DW, Grant, OC, Fuster, MM, Fu, L, Dordick, JS, Woods, RJ, Zhang, F, Linhardt, RJ. . Characterization of heparin and severe acute respiratory syndrome-related coronavirus 2 (SARS-CoV-2) spike glycoprotein binding interactions. *Antiviral Res* **181**, 104873 (2020).
- 22 Rusnati, M., Urbinati, C, Caputo, A, Possati, L, Lortat-Jacob, H, Giacca, M, Ribatti, D, Presta, M. . Pentosan polysulfate as an inhibitor of extracellular HIV-1 Tat. *J Biol Chem* **276**, 22420-22425 (2001).
- 23 Tandon, R., Sharp, JS, Zhang, F, Pomin, VH, Ashpole, NM, Mitra, D, McCandless, MG, Jin, W, Liu, H, Sharma, P, Linhardt, RJ. Effective Inhibition

of SARS-CoV-2 Entry by Heparin and Enoxaparin Derivatives. *J Virol* **95**, e01987-01920 (2021).

24 Yu, M., Zhang, T, Zhang, W, Sun, Q, Li, H, Li, JP. Elucidating the Interactions Between Heparin/Heparan Sulfate and SARS-CoV-2-Related Proteins-An Important Strategy for Developing Novel Therapeutics for the COVID-19 Pandemic. *Front Mol Biosci* **7**, 628551 (2021).

25 Schuurs, Z., Hammond, E, Elli, S, Rudd, TR, Mycroft-West, CJ, Lima, MA, Skidmore, MA, Karlsson, R, Chen, YH, Bagdonaitė, I, Yang, Z, Ahmed, YA, Richard, DJ, Turnbull, J, Ferro, V, Coombe, DR, Gandhi, NS. . Evidence of a putative glycosaminoglycan binding site on the glycosylated SARS-CoV-2 spike protein N-terminal domain. *Comput Struct Biotechnol J* **19**, 2806-2818 (2021).

26 Clausen, T., Sandoval, DR, Spliid, CB, Pihl, J, Painter, CD, et al. SARS-CoV-2 infection depends on cellular heparan sulfate and ACE2. *Cell* **183**, 1043-1057 (2020).

27 Partridge, L., Urwin, L, Nicklin, MJH, James, DC, Green, LR, Monk, PN. ACE2-Independent Interaction of SARS-CoV-2 Spike Protein with Human Epithelial Cells Is Inhibited by Unfractionated Heparin. *Cells* **10**, 1419 (2021).

28 Nie, C., Pouyan, P, Lauster, D, Trimpert, J, Kerkhoff, Y, Szekeres, GP, Wallert, M, Block, S, Sahoo, AK, Dernedde, J, Pagel, K, Kaufer, BB, Netz, RR, Ballauff, M, Haag, R. Polysulfates Block SARS-CoV-2 Uptake through Electrostatic Interactions*. *Angew Chem Int Ed Engl* **60**, 15870-15878 (2021).

29 Kearns, F., Sandoval, DR, Casalino, L, Clausen, TM, Rosenfeld, MA, Spliid, CB, Amaro, RE, Esko, JD. Spike-heparan sulfate interactions in SARS-CoV-2 infection. *Curr Opin Struct Biol* **76**, 102439 (2022).

30 Bhat, E., Khan, J, Sajjad, N, Ali, A, Aldakeel, FM, Mateen, A, Alqahtani, MS, Syed, R. SARS-CoV-2: Insight in genome structure, pathogenesis and viral receptor binding analysis - An updated review. *Int Immunopharmacol* **95**, 107493 (2021).

31 Guney, C., Akar, F. Epithelial and Endothelial Expressions of ACE2: SARS-CoV-2 Entry Routes. *J Pharm Pharm Sci* **24**, 84-93 (2021).

32 Rusnati, M., Urbini, C, Caputo, A, Possati, L, Lortat-Jacob, H, Giacca, M, Ribatti, D, Presta, M. Pentosan polysulfate as an inhibitor of extracellular HIV-1 Tat. *J Biol Chem* **276**, 22420-22425 (2001).

33 Tandon, R., Sharp, JS, Zhang, F, Pomin, VH, Ashpole, NM, Mitra, D, McCandless, MG, Jin, W, Liu, H, Sharma, P, Linhardt, RJ. Effective Inhibition of SARS-CoV-2 Entry by Heparin and Enoxaparin Derivatives. *J Virol* **95**, e01987-01920 (2021).

34 Yadav, R., Chaudhary, JK, Jain, N, Chaudhary, PK, Khanra, S, Dhamija, P, Sharma, A, Kumar, A, Handu, S. Role of Structural and Non-Structural Proteins and Therapeutic Targets of SARS-CoV-2 for COVID-19. *Cells* **10**, 821 (2021).

35 Yuan, H., Wen, HL. Research progress on coronavirus S proteins and their receptors. . *Arch Virol* **28** (2021).

36 Mahmoud, I., Jarrar, YB. Targeting the intestinal TMPRSS2 protease to prevent SARS-CoV-2 entry into enterocytes-prospects and challenges. *Mol Biol Rep* doi: [10.1007/s11033-021-06390-1](https://doi.org/10.1007/s11033-021-06390-1) (2021).

37 Cantuti-Castelvetri, L., Ojha, R, Pedro, LD, Djannatian, M, Franz, J, Kuivanen, S, van der Meer, F, Kallio, K, Kaya, T, Anastasina, M, Smura, T, Levanov, L, Szirovicza, L, Tobi, A, Kallio-Kokko, H, Österlund, P, Joensuu, M, Meunier, FA, Butcher, SJ, Winkler, MS, Mollenhauer, B, Helenius, A, Gokce, O, Teesalu, T, Hepojoki, J, Vapalahti, O, Stadelmann, C, Balistreri, G, Simons, M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science* **370**, 856-860 (2020).

38 Chekol Abebe, E., Mengie Ayele, T, Tilahun Muche, Z, Asmamaw Dejenie, T. . Neuropilin 1: A Novel Entry Factor for SARS-CoV-2 Infection and a Potential Therapeutic Target. *Biologics* **15**, 143-152 (2021).

39 Daly, J., Simonetti, B, Klein, K, Chen, KE, Williamson, MK, Antón-Plágaro, C, Shoemark, DK, Simón-Gracia, L, Bauer, M, Hollandi, R, Greber, UF, Horvath, P, Sessions, RB, Helenius, A, Hiscox, JA, Teesalu, T, Matthews, DA, Davidson, AD, Collins, BM, Cullen, PJ, Yamauchi, Y. Neuropilin-1 is a host

factor for SARS-CoV-2 infection. *Science* **370**, 861-865 (2020).

40 Sajuthi, S., DeFord, P, Li, Y, Jackson, ND, Montgomery, MT, Everman, JL, Rios, CL, Pruesse, E, Nolin, JD, Plender, EG, Wechsler, ME, Mak, ACY, Eng, C, Salazar, S, Medina, V, Wohlford, EM, Huntsman, S, Nickerson, DA, Germer, S, Zody, MC, Abecasis, G, Kang, HM, Rice, KM, Kumar, R, Oh, S, Rodriguez-Santana, J, Burchard, EG, Seibold, MA. Type 2 and interferon inflammation regulate SARS-CoV-2 entry factor expression in the airway epithelium. *Nat Commun* **11**, 5139 (2020).

41 Zmora, P., Moldenhauer, AS, Hofmann-Winkler, H, Pöhlmann, S. TMPRSS2 Isoform 1 Activates Respiratory Viruses and Is Expressed in Viral Target Cells. *PLoS One* **10**, e0138380 (2015).

42 Bertram, S., Glowacka, I, Steffen, I, Kühl, A, Pöhlmann, S. Novel insights into proteolytic cleavage of influenza virus hemagglutinin. *Rev Med Viro* **20**, 298-310 (2010).

43 Bertram, S., Glowacka, I, Blazejewska, P, Soilleux, E, Allen, P, Danisch, S, et al. . . TMPRSS2 and TMPRSS4 facilitate trypsin-independent spread of influenza virus in Caco-2 cells. *J Viro* **84**, 10016-10025 (2010).

44 Muralidhar, S., Gopal, G, Visaga, Ambi, S. Targeting the viral-entry facilitators of SARS-CoV-2 as a therapeutic strategy in COVID-19. *J Med Viro* doi: 10.1002/jmv.27019 (2021).

45 Davies, J., Randeva, HS, Chatha, K, Hall, M, Spandidos, DA, Karteris, E, Kyrou, I. . Neuropilin 1 as a new potential SARS CoV 2 infection mediator implicated in the neurologic features and central nervous system involvement of COVID 19. *Mol Med Rep* **22**, 4221-4226 (2020).

46 Berrou, E., Quarck, R, Fontenay-Roupie, M, Lévy-Toledano, S, Tobelem, G, Bryckaert, M. Transforming growth factor-beta 1 increases internalization of basic fibroblast growth factor by smooth muscle cells: implication of cell-surface heparan sulphate proteoglycan endocytosis. *Biochem J* **311**, 393-399 (1995).

47 Christianson, H., Belting, M. . Heparan sulfate proteoglycan as a cell-surface endocytosis receptor. *Matrix Biol* **35**, 51-55 (2014).

48 Neill, T., Schaefer, L, Iozzo, RV. Decoding the Matrix: Instructive Roles of Proteoglycan Receptors. *Biochemistry* **54**, 4583-4598 (2015).

49 Balistreri, G., Yamauchi, Y, Teesalu, T. A widespread viral entry mechanism: The C-end Rule motif-neuropilin receptor interaction. *Proc Natl Acad Sci USA* **118**, e2112457118 (2021).

50 Danthi, P., Holm, GH, Stehle, T, Dermody, TS. Reovirus receptors, cell entry, and proapoptotic signaling. *Adv Exp Med Biol* **790**, 42-71 (2013).

51 Inoue, J., Ninomiya, M, Shimosegawa, T, McNiven, MA. Cellular Membrane Trafficking Machineries Used by the Hepatitis Viruses. *Hepatology* **68**, 751-762 (2018).

52 Mikulić, S., Florin, L. The endocytic trafficking pathway of oncogenic papillomaviruses. *Papillomavirus Res* **7**, 135-137 (2019).

53 Perera-Lecoin, M., Meertens, L, Carnec, X, Amara, A. Flavivirus entry receptors: an update. *Viruses* **6**, 69-88 (2013).

54 Altgärde, N., Eriksson, C, Peerboom, N, Block, S, Altgärde, N, Wahlsten, O, Möller, S, Schnabelrauch, M, Trybala, E, Bergström, T, Bally, M. Binding Kinetics and Lateral Mobility of HSV-1 on End-Grafted Sulfated Glycosaminoglycans. *Biophys J* **113**, 1223-1234 (2017).

55 Egedal, J., Xie, G, Packard, TA, Laustsen, A, Neidleman, J, Georgiou, K, Pillai, SK, Greene, WC, Jakobsen, MR, Roan, NR. . Hyaluronic acid is a negative regulator of mucosal fibroblast-mediated enhancement of HIV infection. *Mucosal Immunol* **14**, 1203-1213 (2021).

56 Li, P., Fujimoto, K, Bourguignon, L, Yukl, S, Deeks, S, Wong, JK. Exogenous and endogenous hyaluronic acid reduces HIV infection of CD4(+) T cells. *Immunol Cell Biol* **92**, 770-780 (2014).

57 Peerboom, N., Phan-Xuan, T, Moeller, S, Schnabelrauch, M, Svedhem, S, Trybala, E, Bergström, T, Bally, M. Mucin-like Region of Herpes Simplex Virus Type 1 Attachment Protein

Glycoprotein C (gC) Modulates the Virus-Glycosaminoglycan Interaction. *J Biol Chem* **290**, 21473-21485 (2015).

58 Turville, S. Blocking of HIV entry through CD44-hyaluronic acid interactions. *Immunol Cell Biol* **92**, 735-736 (2014).

59 Smith, M., Melrose, J. Impaired instructive and protective barrier functions of the endothelial cell glycocalyx pericellular matrix is impacted in COVID-19 disease. *J Cell Mol Med* **28**, e70033. doi: 70010.71111/jcmm.70033. (2024).

60 Libby, P., and Luscher, T. COVID-19 is, in the end, an endothelial disease. *Eur Heart J* **41**, 3038-3044 (2020).

61 Yang, J., LeBlanc, ME, Cano, I, Saez-Torres, KL, Saint-Geniez, M, Ng, YS, et al. . ADAM10 and ADAM17 proteases mediate proinflammatory cytokine-induced and constitutive cleavage of endomucin from the endothelial surface. *J Biol Chem* **295**, 6641-6651 (2020).

62 Baggen, J. e. a. Genome-wide CRISPR screening identifies TMEM106B as a proviral host factor for SARS-CoV-2. *Nat Genet* <https://doi.org/10.1038/s41588-021-00805-2> (2021).

63 Liu, L., Chopra, P, Li, X, Bouwman, KM, Tompkins, SM, Wolfert, MA, de Vries, RP, Boons, GJ. Heparan Sulfate Proteoglycans as Attachment Factor for SARS-CoV-2. *ACS Cent Sci* **7**, 1009-1018 (2021).

64 Zhang, Q. e. a. Heparan sulfate assists SARS-CoV-2 in cell entry and can be targeted by approved drugs in vitro. *Cell Discov* **6**, 80 (2020).

65 Bao, L. e. a. The pathogenicity of SARS-CoV-2 in hACE2 transgenic mice. *Nature* **583**, 830-833 (2020).

66 Hoffmann, M. e. a. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* <https://doi.org/10.1016/j.cell.2020.02.052> (2020).

67 Lan, J. e. a. Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. *Nature* **581**, 215-220 (2020).

68 Schneider, W. e. a. Genome-scale identification of SARS-CoV-2 and pan-coronavirus host factor networks. *Cell* **184**, 120-132 (2021).

69 Sun, S. e. a. A mouse model of SARS-CoV-2 infection and pathogenesis. *Cell Host Microbe* **28**, 124-133 (2020).

70 Zhou, P. e. a. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* **579**, 270-273 (2020).

71 Balistreri, G., Yamauchi, Y, Teesalu, T. A widespread viral entry mechanism: The C-end Rule motif-neuropilin receptor interaction. . *Proc Natl Acad Sci USA* **118**, e2112457118 (2021).

72 Cantuti-Castelvetri, L., Ojha, R, Pedro, LD, Djannatian, M, Franz, J, Kuivanen, S, van der Meer, F, Kallio, K, Kaya, T, Anastasina, M, Smura, T, Levanov, L, Szirovicza, L, Tobi, A, Kallio-Kokko, H, Österlund, P, Joensuu, M, Meunier, FA, Butcher, SJ, Winkler, MS, Mollenhauer, B, Helenius, A, Gokce, O, Teesalu, T, Hepojoki, J, Vapalahti, O, Stadelmann, C, Balistreri, G, Simons, M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science* **370**, 856-860 (2020).

73 Shi, Q., Jiang, J, Luo, G. Syndecan-1 serves as the major receptor for attachment of hepatitis C virus to the surfaces of hepatocytes. *J Virol* **87**, 6866-6875. doi: 6810.1128/JVI.03475-03412. (2013).

74 Okamoto, K., Kinoshita, H, Parquet, MDC, Raekiansyah, M, Kimura, D, Yui, K, et al Dengue virus strain DEN2 16681 utilizes a specific glycochain of syndecan-2 proteoglycan as a receptor. *J Gen Virol* **93**, 761-770. doi: 710.1099/vir.1090.037853-037850. (2012).

75 Tripathi, S., Li, Y, Luo, G. Syndecan 2 proteoglycan serves as a hepatitis B virus cell attachment receptor. *J Virol* **e0079625**. doi: 10.1128/JVI.00796-25 (2025).

76 de Witte, L., Bobardt, M, Chatterji, U, Degeest, G, David, G, Geijtenbeek, TB, Gallay, P. . Syndecan-3 is a dendritic cell-specific attachment receptor for HIV-1. *Proc Natl Acad Sci U S A* **104**, 19464-19469. doi: 19410.11073/pnas.0703747104. (2007).

77 Hudák, A., Roach, M, Pusztai, D, Pettkó-Szandtner, A, Letoha, A, Szilák, L, et al Syndecan-4 Mediates the Cellular Entry of Adeno-Associated

Virus 9. *Int J Mol Sci* **24**, 3141. doi: 3110.3390/ijms24043141. (2023).

78 Wang, R., Wang, X, Ni, B, Huan, CC, Wu, JQ, Wen, LB, et al Syndecan-4, a PRRSV attachment factor, mediates PRRSV entry through its interaction with EGFR. . *Biochem Biophys Res Commun* **475**, 230-237. doi: 210.1016/j.bbrc.2016.1005.1084. (2016).

79 Verrier, E., Colpitts, CC, Bach, C, Heydmann, L, Weiss, A, Renaud, M, et al. . A targeted functional RNA interference screen uncovers glypican 5 as an entry factor for hepatitis B and D viruses. *Hepatology* **63**, 35-48. doi: 10.1002/hep.28013. (2016).

80 Hu, S., Zhao, K, Lan, Y, Shi, J, Guan, J, Lu, H, et al Cell-surface glycans act as attachment factors for porcine hemagglutinating encephalomyelitis virus. *Vet Microbio* **265**, 109315. doi: 109310.1010/16/j.vetmic.102021.109315. (2022).

81 Shafti-Keramat, S., Handisurya, A, Kriehuber, E, Meneguzzi, G, Slupetzky, K, Kirnbauer, R. Different heparan sulfate proteoglycans serve as cellular receptors for human papillomaviruses. *J Viro* **77**, 13125-13135. doi: 13110.11128/jvi.13177.13124.13125-13135.12003 (2003).

82 Shafti-Keramat, S., Handisurya, A, Kriehuber, E, Meneguzzi, G, Slupetzky, K, Kirnbauer, R. Different heparan sulfate proteoglycans serve as cellular receptors for human papillomaviruses. *J Viro* **77**, 13125-13135. doi: 13110.11128/jvi.13177.13124.13125-13135.12003. (2003).

83 Bermejo-Jambrina, M., Eder, J, Kaptein, TM, van Hamme, JL, Helgers, LC, Vlaming, KE, et al Infection and transmission of SARS-CoV-2 depend on heparan sulfate proteoglycans. . *EMBO J* **40**, e106765. doi: 106710.115252/embj.2020106765 (2021).

84 Gallay, P. Syndecans and HIV-1 pathogenesis. *Microbes Infect* **6**, 617-622. doi: 610.1016/j.micinf.2004.1002.1004. (2004).

85 Bobardt , M., Saphire, AC, Hung, HC, Yu, X, Van der Schueren, B, Zhang, Z, et al Syndecan captures, protects, and transmits HIV to T lymphocytes. *Immunity* **18**, 27-39. doi: 10.1016/s1074-7613(1002)00504-00506. (2003).

86 Saphire, A., Bobardt, MD, Zhang, Z, David, G, Gallay, PA. Syndecans serve as attachment receptors for human immunodeficiency virus type 1 on macrophages. *J Virol* **75**, :9187-9200. doi: 9110.1128/JVI.9175.9119.9187-9200.2001. (2001).

87 Chhabra, M., Shanthamurthy, CD, Kumar ,NV, Mardhekar, S, Vishweshwara, SS, Wimmer, N, Modhiran, N, Watterson, D, Amarilla, AA, Cha, JS, Beckett, JR, De Voss, JJ, Kayal, Y, Vlodavsky, I, Dorsett ,LR, Smith, RAA, Gandhi, NS, Kikkeri, R, Ferro, V. Amphiphilic Heparinoids as Potent Antiviral Agents against SARS-CoV-2. *J Med Chem* **67**, 11885-11916. doi: 11810.11021/acs.jmedchem.11884c00487. (2024).

88 De Clercq, E. Potential drugs for the treatment of AIDS. *J Antimicrob Chemother* **23 Suppl A**, 35-46. doi: 10.1093/jac/1023.suppl_a.1035 (1989).

89 Vert, M. The non-specific antiviral activity of polysulfates to fight SARS-CoV-2, its mutants and viruses with cationic spikes. *J Biomater Sci Polym Ed* **32**, 1466-1471. doi: 1410.1080/09205063.09202021.01925391. (2021).

90 Pavan, M., Fanti ,CD, Lucia, AD, Canato, E, Acuasaliente, L, Sonvico, F, Delgado, J, Hicks, A, Torrelles, JB, Kulkarni ,V, Dwivedi ,V, Zanellato, AM, Galesso, D, Pasut, G, Buttini, F, Martinez-Sobrido, L, Guarise, C. Aerosolized sulfated hyaluronan derivatives prolong the survival of K18 ACE2 mice infected with a lethal dose of SARS-CoV-2. *Eur J Pharm Sci* **187**, 106489. doi: 106410.101016/j.ejps.102023.106489. (2023).

91 Baba, M., Nakajima, M, Schols, D, Pauwels, R, Balzarini, J, De Clercq, E. . Pentosan polysulfate, a sulfated oligosaccharide, is a potent and selective anti-HIV agent in vitro. *Antiviral Res* **9**, 335-343. doi: 310.1016/0166-3542(1088)90035-90036. (1988).

92 Rusnati, M., Urbinati, C, Caputo, A, Possati, L, Lortat-Jacob, H, Giacca, M, Ribatti, D, Presta, M. Pentosan polysulfate as an inhibitor of extracellular HIV-1 Tat. *J Biol Chem* **276**, 22420-22425 (2001).

93 Biesert, L., Suhartono, H., Winkler, I., Meichsner, C., Helsberg, M., Hewlett, G., Klimetzek, V., Mölling, K., Schlumberger, HD., Schrinner, E., et al. Inhibition of HIV and virus replication by polysulphated polyxyran: HOE/BAY 946, a new antiviral compound. *AIDS* **2**, 449-457. doi: 410.1097/00002030-198812000-198800007. (1988).

94 Biesert, L., Adamski, M., Zimmer, G., Suhartono, H., Fuchs, J., Unkelbach, U., Mehlhorn, RJ., Hideg, K., Milbradt, R., Rübsamen-Waigmann, H. . Anti-human immunodeficiency virus (HIV) drug HOE/BAY946 increases membrane hydrophobicity of human lymphocytes and specifically suppresses HIV-protein synthesis. *Med Microbiol Immunol* **179**, 307-321. doi: 310.1007/BF00189609. (1990).

95 von Briesen, H., Meichsner, C., Andreesen, R., Esser, R., Schrinner, E., Rübsamen-Waigmann, H. . The polysulphated polyxyran Hoe/Bay-946 inhibits HIV replication on human monocytes/macrophage s. *Res Virol* **141**, 251-257. doi: 210.1016/0923-2516(1090)90029-i. (1990).

96 Peters, M., Witvrouw, M., De Clercq, E., Ruf, B. Pharmacokinetics of intravenous pentosan polysulphate (HOE/BAY 946) in HIV-positive patients. *AIDS* **5**, 1534-1535. doi: 1510.1097/00002030-199112000-199100021. (1991).

97 Pearce-Pratt, R., Phillips, DM. . Sulfated polysaccharides inhibit lymphocyte-to-epithelial transmission of human immunodeficiency virus-1. *Biol Reprod* **54**, 173-182. doi: 110.1095/biolreprod1054.1091.1173. (1996).

98 Witvrouw, M., Desmyter, J., De Clercq, E. . Antiviral portrait series: 4. Polysulfates as inhibitors of HIV and other enveloped viruses. . *Antiviral Chemistry & Chemotherapy* **5**, 345-359 (1994).

99 Jurkiewicz E, P. P., Jentsch KD, Hartmann H, Hunsmann G. In vitro anti-HIV-1 activity of chondroitin polysulphate. *AIDS* **3**, 423-427. doi: 410.1097/00002030-198907000-198900003. (1989).

100 Vanderlinden, E., Boonen, A., Noppen, S., Schoofs, G., Imbrechts, M., Geukens, N., Snoeck, R., Stevaert, A., Naesens, L., Andrei, G., Schols, D. PRO-2000 exhibits SARS-CoV-2 antiviral activity by interfering with spike-heparin binding. *Antiviral Res* **217**, 105700. doi: 105710.101016/j.antiviral.102023.105700. (2023).

101 Huskens, D., Profy, AT., Vermeire, K., Schols, D. PRO 2000, a broadly active anti-HIV sulfonated compound, inhibits viral entry by multiple mechanisms. *Retrovirology* **7 Suppl 1**, P17. doi: 10.1186/1742-4690-1187-S1181-P1117 (2010).

102 Nakamura, T., Satoh, K., Fukuda, T., Kinoshita, I., Nishiura, Y., Nagasato, K., Yamauchi, A., Kataoka, Y., Nakamura, T., Sasaki, H., Kumagai, K., Niwa, M., Noguchi, M., Nakamura, H., Nishida, N., Kawakami, A. Pentosan polysulfate treatment ameliorates motor function with increased serum soluble vascular cell adhesion molecule-1 in HTLV-1-associated neurologic disease. *J Neurovirol* **20**, 269-277. doi: 210.1007/s13365-13014-10244-13368. (2014).

103 Callaway, E. Heavily mutated Omicron variant puts scientists on alert. *Nature* **600**, 21 (2021).

104 WHO. Classification of Omicron (B.1.1.529): SARS-CoV-2 Variant of Concern. [https://www.who.int/news-room/detail/26-11-2021-classification-of-omicron-\(b.1.1.529\)-sars-cov-2-variant-of-concern](https://www.who.int/news-room/detail/26-11-2021-classification-of-omicron-(b.1.1.529)-sars-cov-2-variant-of-concern). (2021).

105 Smith, M., Melrose, J. . Pentosan Polysulfate Affords Pleotropic Protection to Multiple Cells and Tissues. *Pharmaceuticals (Basel)* **16**, 437 (2023).

106 GISAID. <https://www.gisaid.org/hcov19-variants/>. (2021).

107 Basheer, A., Zahoor, I., Yaqub, T. . Genomic architecture and evolutionary relationship of BA.2.75: A Centaurus subvariant of Omicron SARS-CoV-2. *PLoS One* **18**, e0281159. doi: 0281110.0281371/journal.pone.0281159. (2023).

108 Caputo, E., Mandrich, L. SARS-CoV-2: Searching for the Missing Variants. *Viruses* **14**, 2364. doi: 2310.3390/v14112364. (2022).

109 Zappa, M., Verdecchia, P., Angeli, F. Knowing the new Omicron BA.2.75 variant ('Centaurus'): A simulation study. *Eur J Intern Med* **105**, 107-108. doi: 110.1016/j.ejim.2022.1008.1009. (2022).

110 Sabbatucci, M., Vitiello, A., Clemente, S., Zovi, A., Boccellino, M., Ferrara, F., Cimmino, C., Langella, R., Ponzo, A., Stefanelli, P., Rezza, G. . Omicron variant evolution on vaccines and monoclonal

antibodies. *Inflammopharmacology* **31**, 1779-1788 . doi: 1710.1007/s10787-10023-01253-10786 (2023).

111 Fernandes, Q., Inchakalody, VP, Merhi, M, Mestiri, S, Taib, N, Moustafa, Abo El-Ella, D, Bedhiafi, T, Raza, A, Al-Zaidan, L, Mohsen, MO, Yousuf Al-Nesf, MA, Hssain, AA, Yassine, HM, Bachmann, MF, Uddin, S, Dermime, S. Emerging COVID-19 variants and their impact on SARS-CoV-2 diagnosis, therapeutics and vaccines. *Ann Med* **54**, 524-540 (2022).

112 Dechechchi, M., Tamanini, A, Bonizzato, A, Cabrini, G. Heparan sulfate glycosaminoglycans are involved in adenovirus type 5 and 2-host cell interactions. *Virology* **268**, 382-390 (2000).

113 Lerch, T., Chapman, MS. . Identification of the heparin binding site on adeno-associated virus serotype 3B (AAV-3B). . *Virology* **423**, 6-13 (2012).

114 Qiu, J., Handa, A, Kirby, M, Brown, KE. . The interaction of heparin sulfate and adeno-associated virus 2. *Virology* **269**, 137-147 (2000).

115 Murakami, S., Takenaka-Uema, A, Kobayashi, T, Kato, K, Shimojima, M, Palmarini, M, Horimoto, T. Heparan Sulfate Proteoglycan Is an Important Attachment Factor for Cell Entry of Akabane and Schmallenberg Viruses. *J Virol* **91**, pii: e00503-00517 (2017).

116 McAllister , N., Liu, Y, Silva, LM, Lentscher, AJ, Chai, W, Wu, N, Griswold, KA, Raghunathan, K, Vang, L, Alexander ,J, Warfield, KL, Diamond, MS, Feizi,T, Silva, LA, Dermody, TS. Chikungunya Virus Strains from Each Genetic Clade Bind Sulfated Glycosaminoglycans as Attachment Factors. *J Virol* **94**, e01500-01520. (2020).

117 Merilahti, P., Karelehto, E, Susi, P. Role of Heparan Sulfate in Cellular Infection of Integrin-Binding Coxsackievirus A9 and Human Parechovirus 1 Isolates. . *PLoS One* **11**, e0147168 (2016).

118 Pourianfar , H., Kirk, K, Grollo, L. Initial evidence on differences among Enterovirus 71, Coxsackievirus A16 and Coxsackievirus B4 in binding to cell surface heparan sulphate. *Virusdisease* **25**, 277-284 (2014).

119 Zautner, A., Jahn, B, Hammerschmidt, E, Wutzler, P, Schmidtke, M. N- and 6-O-sulfated heparan sulfates mediate internalization of coxsackievirus B3 variant PD into CHO-K1 cells. *J Virol* **80**, 6629-6636 (2006).

120 Artpradit, C., Robinson, LN, Gavrilov, BK, Rurak, TT, Ruchirawat, M, Sasisekharan, R. Recognition of heparan sulfate by clinical strains of dengue virus serotype 1 using recombinant subviral particles. *Virus Res* **176**, 69-77 (2013).

121 Hilgard, P., Stockert, R. Heparan sulfate proteoglycans initiate dengue virus infection of hepatocytes. *Hepatology* **32**, 1069-1077 (2000).

122 Saksono, B., Dewi, BE, Nainggolan, L, Suda, Y. A Highly Sensitive Diagnostic System for Detecting Dengue Viruses Using the Interaction between a Sulfated Sugar Chain and a Virion. *PLoS One* **10**, e0123981 (2015).

123 Wu, S., Wu, Z, Wu, Y, Wang, T, Wang, M, Jia, R, Zhu, D, Liu, M, Zhao, X, Yang, Q, Wu, Y, Zhang, S, Liu, Y, Zhang, L, Yu, Y, Pan, L, Chen, S, Cheng, A. . Heparin sulfate is the attachment factor of duck Tembus virus on both BHK21 and DEF cells. *Viro J* **16**, 134 (2019).

124 Tamhankar, M., Gerhardt, DM, Bennett, RS, Murphy, N, Jahrling, PB, Patterson, JL. Heparan sulfate is an important mediator of Ebola virus infection in polarized epithelial cells. *Viro J* **15**, 135 (2018).

125 Israelsson, S., Gullberg, M, Jonsson, N, Roivainen, M, Edman, K, Lindberg, AM. Studies of Echovirus 5 interactions with the cell surface: heparan sulfate mediates attachment to the host cell. *Virus Res* **151**, 170-176 (2010).

126 Tan, C., Poh, CL, Sam, IC, Chan, YF. Enterovirus 71 uses cell surface heparan sulfate glycosaminoglycan as an attachment receptor. . *J Virol* **87**, 611-620 (2013).

127 Tan, C., Sam, IC, Lee, VS, Wong, HV, Chan, YF. VP1 residues around the five-fold axis of enterovirus A71 mediate heparan sulfate interaction. *Virology* **501**, 79-87 (2017).

128 Salvador, B., Sexton, NR, Carrion, R Jr, Nunneley, J, Patterson, JL, Steffen, I, Lu, K,

Muench, MO, Lembo, D, Simmons, G. . Filoviruses utilize glycosaminoglycans for their attachment to target cells. *J Virol* **87**, 3295-3304 (2013).

129 Mathieu, C., Dhondt, KP, Châlons, M, Mély, S, Raoul, H, Negre, D, Cosset, FL, Gerlier, D, Vivès, RR, Horvat, B. Heparan sulfate-dependent enhancement of henipavirus infection. *mBio* **6**, e02427 (2015).

130 Schulze, A., Gripon, P, Urban, S. . Hepatitis B virus infection initiates with a large surface protein-independent binding to heparan sulfate proteoglycans. *Hepatology* **46**, 1759-1766 (2007).

131 Lamas Longarela, O., Schmidt, TT, Schöneweis, K, Romeo, R, Wedemeyer, H, Urban, S, Schulze, A. P. Proteoglycans act as cellular hepatitis delta virus attachment receptors. . *PLoS One* **8**, e58340 (2013).

132 Olenina, L., Kuzmina, TI, Sobolev, BN, Kuraeva, TE, Kolesanova, EF, Archakov, AI. . Identification of glycosaminoglycan-binding sites within hepatitis C virus envelope glycoprotein E2. . *J Viral Hepat* **12**, 584-593 (2005).

133 Xu, Y., Martinez, P, Séron, K, Luo, G, Allain, F, Dubuisson, J, Belouzard, S. Characterization of hepatitis C virus interaction with heparan sulfate proteoglycans. *J Virol* **89**, 3846-3858 (2015).

134 Akula, S., Pramod, NP, Wang, FZ, Chandran, B. Human herpesvirus 8 envelope-associated glycoprotein B interacts with heparan sulfate-like moieties. *Virology* **284**, 235-249 (2001).

135 Feyzi, E., Trybala, E, Bergström, T, Lindahl, U, Spillmann, D. . Structural requirement of heparan sulfate for interaction with herpes simplex virus type 1 virions and isolated glycoprotein C. *J Biol Chem* **372**, 24850-24857 (1997).

136 Thammawat, S., Sadlon, TA, Hallsworth, PG, Gordon, DL. . Role of cellular glycosaminoglycans and charged regions of viral G protein in human metapneumovirus infection. *J Virol* **82**, 11767-11774 (2008).

137 Karasneh, G., Ali, M, Shukla, D. . An important role for syndecan-1 in herpes simplex virus type-1 induced cell-to-cell fusion and virus spread. *PLoS One* **6**, e25252 (2011).

138 Terry-Allison, T., Montgomery, RI, Warner, MS, Geraghty, RJ, Spear, PG. Contributions of gD receptors and glycosaminoglycan sulfation to cell fusion mediated by herpes simplex virus 1. *Virus Res* **74**, 39-45 (2001).

139 Connell, B. a. L.-J., H. Human immunodeficiency virus and heparan sulfate: from attachment to entry inhibition. *Frontiers in Immunology* **4**, 385 (2013).

140 Guibinga, G., Miyanohara, A, Esko, JD, Friedmann, T. Cell surface heparan sulfate is a receptor for attachment of envelope protein-free retrovirus-like particles and VSV-G pseudotyped MLV-derived retrovirus vectors to target cells. *Mol Ther* **5**, 538-546 (2002).

141 Trkola, A., Gordon, C, Matthews, J, Maxwell, E, Ketas, T, Czaplewski, L, Proudfoot, AE, Moore, JP. The CC-chemokine RANTES increases the attachment of human immunodeficiency virus type 1 to target cells via glycosaminoglycans and also activates a signal transduction pathway that enhances viral infectivity. *J Virol* **73**, 6370-6379 (1999).

142 Nasimuzzaman, M., Persons, DA. Cell Membrane-associated heparan sulfate is a receptor for prototype foamy virus in human, monkey, and rodent cells. *Mol Ther*. 2012 Jun;20(6):1158-66. Cell Membrane-associated heparan sulfate is a receptor for prototype foamy virus in human, monkey, and rodent cells. *Mol Ther* **20**, 1158-1166 (2012).

143 Plochmann, K., Horn, A, Gschmack, E, Armbruster, N, Krieg, J, Wiktorowicz, T, Weber, C, Stirnnagel, K, Lindemann, D, Rethwilm, A, Scheller, C. Heparan sulfate is an attachment factor for foamy virus entry. *J Virol* **86**, 10028-10035 (2012).

144 Bousarghin, L., Hubert, P, Franzen, E, Jacobs, N, Boniver, J, Delvenne, P. Human papillomavirus 16 virus-like particles use heparan sulfates to bind dendritic cells and colocalize with langerin in Langerhans cells. *J Gen Virol* **86**, 1297-1305 (2005).

145 Feldman, S., Audet, S, Beeler, JA. . The fusion glycoprotein of human respiratory syncytial virus

facilitates virus attachment and infectivity via an interaction with cellular heparan sulfate. *J Virol* **74**, 6442-6447 (2000).

146 Chang, A., Masante, C, Buchholz, UJ, Dutch, RE. Human metapneumovirus (HMPV) binding and infection are mediated by interactions between the HMPV fusion protein and heparan sulfate. *J Virol* **86**, 3230-3243 (2012).

147 Su, C., Liao, CL, Lee, YL, Lin, YL. Highly sulfated forms of heparin sulfate are involved in japanese encephalitis virus infection. *Virology* **286**, 206-215 (2001).

148 Chowalter, R., Pastrana, DV, Buck, CB. Glycosaminoglycans and sialylated glycans sequentially facilitate Merkel cell polyomavirus infectious entry. *PLoS Pathog* **7**, e1002161 (2011).

149 Kureishi, N., Faruque, D, Porter, CD. Primary attachment of murine leukaemia virus vector mediated by particle-associated heparan sulfate proteoglycan. *Biochem J* **400**, 421-430 (2006).

150 Gillet, L, Adler, H, Stevenson, PG. Glycosaminoglycan interactions in murine gammaherpesvirus-68 infection. *PLoS One* **2**, e347 (2007).

151 Huan, C., Wang, Y, Ni, B, Wang, R, Huang, L, Ren, XF, Tong, GZ, Ding, C, Fan, HJ, Mao, X. Porcine epidemic diarrhea virus uses cell-surface heparan sulfate as an attachment factor. *Arch Virol* **160**, 1621-1628 (2015).

152 Trybala, E., Bergström, T, Spillmann, D, Svennerholm, B, Olofsson, S, Flynn, SJ, Ryan, P. Mode of interaction between pseudorabies virus and heparan sulfate/heparin. *Virology* **218**, 35-42 (1996).

153 Sasaki, M., Anindita, PD, Ito, N, Sugiyama, M, Carr, M, Fukuhara, H, Ose, T, Maenaka, K, Takada, A, Hall, WW, Orba, Y, Sawa, H. The Role of Heparan Sulfate Proteoglycans as an Attachment Factor for Rabies Virus Entry and Infection. *J Infect Dis* **217**, 1740-1749 (2018).

154 Hallak, L., Spillmann, D, Collins, PL, Peebles, ME. Glycosaminoglycan sulfation requirements for respiratory syncytial virus infection. *J Virol* **74**, 10508-10513 (2000).

155 Shields, B., Mills, J, Ghildyal, R, Gooley, P, Meanger, J. Multiple heparin binding domains of respiratory syncytial virus G mediate binding to mammalian cells. *Arch Virol* **148**, 1987-2003 (2003).

156 Ennemoser, M., Rieger, J, Muttenthaler, E, Gerlza, T, Zatloukal, K, Kungl, AJ. Enoxaparin and pentosan polysulfate bind to the SARS-CoV-2 spike protein and human ACE2 receptor, inhibiting Vero cell infection. *Biomedicines* **10**, 49 (2022).

157 Escribano-Romero, E., Jimenez-Clavero, MA, Gomes, P, García-Ranea, JA, Ley, V. Heparan sulphate mediates swine vesicular disease virus attachment to the host cell. *J Gen Virol* **85**, 653-663 (2004).

158 Byrnes, A., Griffin, DE. Binding of Sindbis virus to cell surface heparan sulfate. *J Virol* **72**, 7349-7356 (1998).

159 Hulst, M., van Gennip, HG, Moormann, RJ. . Passage of classical swine fever virus in cultured swine kidney cells selects virus variants that bind to heparan sulfate due to a single amino acid change in envelope protein E(rns). *J Virol* **74**, 9553-9561 (2000).

160 Luteijn, R., van Diemen, F, Blomen, VA, Boer, IGJ, Manikam Sadasivam, S, van Kuppevelt, TH, Drexler, I, Brummelkamp, TR, Lebbink, RJ, Wiertz, EJ. . A Genome-Wide Haploid Genetic Screen Identifies Heparan Sulfate-Associated Genes and the Macropinocytosis Modulator TMED10 as Factors Supporting Vaccinia Virus Infection. *J Virol* **93**, e02160-02118 (2019).

161 Tan, C., Sam, IC, Chong, WL, Lee, VS, Chan, YF. Polysulfonate suramin inhibits Zika virus infection. *Antiviral Res* **143**, 186-194 (2017).

162 García-Villalón, D., Gil-Fernández, C. . Antiviral activity of sulfated polysaccharides against African swine fever virus. *Antiviral Res* **15**, 139-148 (1991).

163 Baba, M., Snoeck, R, Pauwels, R, de Clercq, E. Sulfated polysaccharides are potent and selective inhibitors of various enveloped viruses, including herpes simplex virus, cytomegalovirus, vesicular stomatitis virus, and human immunodeficiency virus. *Antimicrob Agents Chemother* **32**, 1742-1745 (1988).

164 Baba, M., Nakajima, M, Schols, D, Pauwels, R, Balzarini, J, De Clercq, E. . Pentosan polysulfate, a sulfated oligosaccharide, is a potent and selective anti-HIV agent in vitro. *Antiviral Res* **9**, 335-343 (1988).

165 Ma, G., Yasunaga, JI, Ohshima, K, Matsumoto, T, Matsuoka, M. Pentosan Polysulfate Demonstrates Anti-human T-Cell Leukemia Virus Type 1 Activities In Vitro and In Vivo. *J Virol* **93**, e00413-00419 (2019).

166 Herrero, L., Foo, SS, Sheng, KC, Chen, W, Forwood, MR, Bucala, R, Mahalingam, S. Pentosan Polysulfate: a Novel Glycosaminoglycan-Like Molecule for Effective Treatment of Alphavirus - Induced Cartilage Destruction and Inflammatory Disease. *J Virol* **89**, 8063-8076 (2015).

167 Krishnan, R., Duiker, M, Rudd, PA, Skerrett, D, Pollard, JGD, Siddel, C, Rifat, R, Ng, JHK, Georgius, P, Hererro, LJ, Griffin, P. Pentosan polysulfate sodium for Ross River virus-induced arthralgia: a phase 2a, randomized, double-blind, placebo-controlled study. *BMC Musculoskelet Disord* **22**, 271 (2021).

168 Tavares-Júnior, J., de Souza, ACC, Borges, JWP, Oliveira, DN, Siqueira-Neto, JI, Sobreira-Neto, MA, Braga-Neto, P. COVID-19 associated cognitive impairment: A systematic review. . *Cortex* **152**, 77-97 (2022).

169 Kumagai, K., Shirabe, S, Miyata, N, Murata, M, Yamauchi, A, Kataoka, Y, Niwa, M. Sodium pentosan polysulfate resulted in cartilage improvement in knee osteoarthritis--an open clinical trial. *BMC Clin Pharmacol* **10**, 7. doi: 10.1186/1472-6904-1110-1187. (2010).

170 Eita, M., Ashour, RH, El-Khawaga, OY. . Pentosan polysulfate exerts anti-inflammatory effect and halts albuminuria progression in diabetic nephropathy: Role of combined losartan. . *Fundam Clin Pharmacol* **36**, 801-810. doi: 810.1111/fcp.12781 (2022).

171 Rudd, P., Lim, EXY, Stapledon, CJM, Krishnan, R, Herrero, LJ. Pentosan polysulfate sodium prevents functional decline in chikungunya infected mice by modulating growth factor signalling and lymphocyte activation. *PLoS ONE* **16**, e0255125 (2021).

172 Krishnan, R., Stapledon, CJM, Mostafavi, H, Freitas, JR, Liu, X, Mahalingam, S, Zaid, A. Anti-inflammatory actions of Pentosan polysulfate sodium in a mouse model of influenza virus A/PR8/34-induced pulmonary inflammation. *Front Immunol* **14**, 1030879. doi: 1030810.1033389/fimmu.1032023.1030879 (2023).

173 Bertini, S., Alekseeva, A, Elli, S, Pagani, I, Zanzoni, S, Eisele, G, Krishnan, R, Maag, KP, Reiter, C, Lenhart, D, Gruber, R, Yates, EA, Vicenzi, E, Naggi, A, Bisio, A, Guerrini, M. Pentosan Polysulfate Inhibits Attachment and Infection by SARS-CoV-2 In Vitro: Insights into Structural Requirements for Binding. . *Thromb Haemost* **122**, 984-997. doi: 910.1055/a-1807-0168 (2022).

174 Wool, G., Miller, JL. . The Impact of COVID-19 Disease on Platelets and Coagulation. *Pathobiology* **88**, 15-27, doi:doi: 10.1159/000512007. (2021).

175 Iba, T., Levy, JH, Levi, M, Thachil, J. Coagulopathy in COVID-19. *J Thromb Haemost* **18**, 2103-2109, doi:doi: 10.1111/jth.14975 (2020).

176 Grobler, C., Maphumulo, SC, Grobbelaar, LM, Bredenkamp, JC, Laubscher, GJ, Lourens, PJ, Steenkamp, J, Kell, DB, Pretorius, E. Covid-19: The Rollercoaster of Fibrin(Ogen), D-Dimer, Von Willebrand Factor, P-Selectin and Their Interactions with Endothelial Cells, Platelets and Erythrocytes. *Int J Mol Sci* **21**, 5168, doi: doi: 10.3390/ijms21145168. (2020).

177 Vögtle, T., Sharma, S, Mori, J, Nagy, Z, Semeniak, D, Scandola, C, Geer, MJ, Smith, CW, Lane, J, Pollack, S, Lassila, R, Jouppila, A, Barr, AJ, Ogg, DJ, Howard, TD, McMiken, HJ, Warwicker, J, Geh C, Rowlinson, R, Abbott, WM, Eckly, A, Schulze, H, Wright, GJ, Mazharian, A, Fütterer, K, Rajesh, S, Douglas, MR, Senis, YA. Heparan sulfates are critical regulators of the inhibitory megakaryocyte-platelet receptor G6b-B. . *Elife* **8**, e46840 (2019).

178 Koupenova, M., Corkrey, HA, Vitseva, O, Tanriverdi, K, Somasundaran, M, Liu, P, Soofi, S, Bhandari, R, Godwin, M, Parsi, KM, Cousineau, A,

Maehr, R, Wang, JP, Cameron, SJ, Rade, J, Finberg, RW, Freedman, JE. SARS-CoV-2 Initiates Programmed Cell Death in Platelets. *Circ Res* **129**, 631-646 (2021).

179 Lord, M., Tang, F, Rnjak-Kovacina, J, Smith, JGW, Melrose, J, Whitelock, JM. . The multifaceted roles of perlecan in fibrosis. . *Matrix Biol* **68-69**, 150-166 (2018).

180 Lord, M., Chuang, CY, Melrose, J, Davies, MJ, Iozzo, RV, Whitelock, JM. . The role of vascular-derived perlecan in modulating cell adhesion, proliferation and growth factor signaling. *Matrix Biol* **35**, 112-122 (2014).

181 Whitelock, J., Melrose, J, Iozzo, RV. Diverse cell signaling events modulated by perlecan. *Biochemistry* **47**, 11174-11183 (2008).

182 Hayashida, K., Aquino, RS, Park, PW. Coreceptor functions of cell surface heparan sulfate proteoglycans. *Am J Physiol Cell Physiol* **322**, C896-C912 (2022).

183 Smits, N., Shworak, NW, Dekhuijzen, PN, van Kuppevelt, TH. Heparan sulfates in the lung: structure, diversity, and role in pulmonary emphysema. *Anat Rec (Hoboken)* **293**, 955-967 (2010).

184 Burgess, J., Mauad, T, Tjin, G, Karlsson, JC, Westergren-Thorsson, G. The extracellular matrix - the under-recognized element in lung disease? *J Pathol* **240**, 397-409 (2016).

185 Burgstaller, G., Oehrle, B, Gerckens, M, White, ES, Schiller, HB, Eickelberg, O. The instructive extracellular matrix of the lung: basic composition and alterations in chronic lung disease. . *Eur Respir J* **50**, 1601805 (2017).

186 Zhou, Y., Horowitz, JC, Naba, A, Ambalavanan, N, Atabai, K, Balestrini, J, Bitterman, PB, Corley, RA, Ding, BS, Engler, AJ, Hansen, KC, Hagood, JS, Kheradmand, F, Lin, QS, Neptune, E, Niklason, L, Ortiz, LA, Parks, WC, Tschumperlin, DJ, White, ES, Chapman, HA, Thannickal, VJ. . Extracellular matrix in lung development, homeostasis and disease. *Matrix Biol* **73**, 77-104 (2018).

187 Hackett, T., Osei, ET. Modeling Extracellular Matrix-Cell Interactions in Lung Repair and Chronic Disease. *Cells* **10**, 2145 (2021).

188 Phogat, S., Thiam, F, Al Yazeedi, S, Abokor, FA, Osei, ET. 3D in vitro hydrogel models to study the human lung extracellular matrix and fibroblast function. *Respir Res* **24**, 242 (2023).

189 Ma, H., Wu, X, Li, Y, Xia, Y. . Research Progress in the Molecular Mechanisms, Therapeutic Targets, and Drug Development of Idiopathic Pulmonary Fibrosis. *Front Pharmacol* **13**, 963054 (2022).

190 Patel, D., Snelgrove, RJ. The multifaceted roles of the matrikine Pro-Gly-Pro in pulmonary health and disease. *Eur Respir Rev* **27**, 180017 (2018).

191 Burgess, J., Weckmann, M. Matrikines and the lungs. *Pharmacol Ther* **134**, 317-337 (2012).

192 Gressett, T., Hossen, ML, Talkington, G, Volic, M, Perez, H, Tiwari, PB, Chapagain, P, Bix, G. Molecular interactions between perlecan LG3 and the SARS-CoV-2 spike protein receptor binding domain. *Protein Sci* **33**, e4843 (2024).

193 Lavorgna, T., Gressett, TE, Chastain, WH, Bix, GJ. . Perlecan: a review of its role in neurologic and musculoskeletal disease. *Front Physiol* **14**, 1189731 (2023).

194 Hayes, A., Farrugia, BL, Biose, IJ, Bix, GJ, Melrose, J. Perlecan, A Multi-Functional, Cell-Instructive, Matrix-Stabilizing Proteoglycan With Roles in Tissue Development Has Relevance to Connective Tissue Repair and Regeneration. *Front Cell Dev Biol* **10**, 856261 (2022).

195 Kunnathattil, M., Rahul, P, Skaria, T. Soluble vascular endothelial glycocalyx proteoglycans as potential therapeutic targets in inflammatory diseases. . *Immunol Cell Biol* **102**, 97-116 (2024).

196 Yoshida, H., Nagaoka, A, Kusaka-Kikushima, A, Tobiishi, M, Kawabata, K, Sayo, T, Sakai, S, Sugiyama, Y, Enomoto, H, Okada, Y, Inoue, S. KIAA1199, a deafness gene of unknown function, is a new hyaluronan binding protein involved in hyaluronan depolymerization. *Proc Natl Acad Sci U S A* **110**, 5612-5617 (2013).

197 Shimizu, H., Shimoda, M., Mochizuki, S., Miyamae, Y., Abe, H., Chijiwa, M., Yoshida, H., Shiozawa, J., Ishijima, M., Kaneko, K., Kanaji, A., Nakamura, M., Toyama, Y., Okada, Y. Hyaluronan-Binding Protein Involved in Hyaluronan Depolymerization Is Up-Regulated and Involved in Hyaluronan Degradation in Human Osteoarthritic Cartilage. *Am J Pathol* **188**, 2109-2119 (2018).

198 Song, Y., He, P., Rodrigues, AL, Datta, P., Tandon, R., Bates, JT, Bierdeman, MA, Chen, C., Dordick, J., Zhang, F., Linhardt, RJ. . Anti-SARS-CoV-2 Activity of Rhamnan Sulfate from *Monostroma nitidum*. *Mar Drugs* **19**, 685 (2021).

199 Koumpa, F., Forde, CT, Manjaly, JG. Sudden irreversible hearing loss post COVID-19. *BMJ Case Rep* **13**, e238419 (2020).

200 Francis, D., Hutadilok, N., Kongtawelert, P., Ghosh, P. Pentosan polysulphate and glycosaminoglycan polysulphate stimulate the synthesis of hyaluronan in vivo. *Rheumatol Int* **13**, 61-64 (1993).

201 Shen, B., Shimmon, S., Smith, MM, Ghosh, P. Biosensor analysis of the molecular interactions of pentosan polysulfate and of sulfated glycosaminoglycans with immobilized elastase, hyaluronidase and lysozyme using surface plasmon resonance (SPR) technology. *J Pharm Biomed Anal* **31**, 83-93. doi: 10.1016/s0731-7085(00)00606-00604. (2003).

202 Slevin, M., Krupinski, J., Gaffney, J., Matou, S., West, D., Delisser, H., Savani, RC, Kumar, S. Hyaluronan-mediated angiogenesis in vascular disease: uncovering RHAMM and CD44 receptor signaling pathways. *Matrix Biol* **26**, 58-68 (2007).

203 Genasetti, A., Vigetti, D., Viola, M., Karousou, E., Moretto, P., Rizzi, M., Bartolini, B., Clerici, M., Pallotti, F., De Luca, G., Passi, A. Hyaluronan and human endothelial cell behavior. *Connect Tissue Res* **49**, 120-123 (2008).

204 Matou-Nasri, S., Gaffney, J., Kumar, S., Slevin, M. Oligosaccharides of hyaluronan induce angiogenesis through distinct CD44 and RHAMM-mediated signalling pathways involving Cdc2 and gamma-adducin. *Int J Oncol* **35**, 761-773 (2009).

205 Slevin, M., Krupinski, J., Kumar, S., Gaffney, J. Angiogenic oligosaccharides of hyaluronan induce protein tyrosine kinase activity in endothelial cells and activate a cytoplasmic signal transduction pathway resulting in proliferation. *Lab Invest* **78**, 987-1003 (1998).

206 Wang, Y., Cao, ML, Liu, YW, He, YQ, Yang, CX, Gao, F. CD44 mediates oligosaccharides of hyaluronan-induced proliferation, tube formation and signal transduction in endothelial cells. *Exp Biol Med (Maywood)* **236**, 84-90 (2011).

207 Castor, C., Fremuth, TD, Furlong, AM, Jourdian, GW. Hyaluronic acid and proteoglycan synthesis by lung fibroblasts in basal and activated states. *In Vitro* **19**, 462-470 (1983).

208 Ambrosino, P., Calcaterra, IL, Mosella, M., Formisano, R., D'Anna, SE, Bachetti, T., Marcuccio, G., Galloway, B., Mancini, FP, Papa, A., Motta, A., Di Minno, MND, Maniscalco, M. Endothelial Dysfunction in COVID-19: A Unifying Mechanism and a Potential Therapeutic Target. *Biomedicines* **10**, 812 (2022).

209 Liu, N., Long, H., Sun, J., Li, H., He, Y., Wang, Q., Pan, K., Tong, Y., Wang, B., Wu, Q., Gong, L. New laboratory evidence for the association between endothelial dysfunction and COVID-19 disease progression. *J Med Viro* **94**, 3112-3120 (2022).

210 Robles, J., Zamora, M., Adan-Castro, E., Siqueiros-Marquez, L., Martinez de la Escalera, G., Clapp, C. The spike protein of SARS-CoV-2 induces endothelial inflammation through integrin $\alpha 5\beta 1$ and NF- κB signaling. *J Biol Chem* **298**, 101695 (2022).

211 Sbirkov, Y., Dzharov, V., Todorova, K., Hayrabyan, S., Sarafian, V. Endothelial inflammation and dysfunction in COVID-19. *Vasa* **51**, 62-70. doi: 10.1024/0301-1526/a000991. (2022).

212 Smith, M., Melrose, J. Xylan Prebiotics and the Gut Microbiome Promote Health and Wellbeing: Potential Novel Roles for Pentosan Polysulfate. *Pharmaceuticals (Basel)* **15**, 1151 (2022).

213 Al-Zahrani, A., Gajewski, JB. Long-term efficacy and tolerability of pentosan polysulphate

sodium in the treatment of bladder pain syndrome. *Can Urol Assoc J* **5**, 113-118 (2011).

214 Anderson, J., Edelman, J, Ghosh, P. Effects of pentosan polysulphate on peripheral blood leukocyte populations and mononuclear cell procoagulant activity in patients with osteoarthritis. *Curr Therap Res* **58**, 93-107 (1997).

215 Nickel, J., Forrest, JB, Tomera, K, Hernandez-Graulau, J, Moon, TD, Schaeffer, AJ, Krieger, JN, Zeitlin, SI, Evans, RJ, Lama, DJ, Neal, DE Jr, Sant, GR. Pentosan polysulfate sodium therapy for men with chronic pelvic pain syndrome: a multicenter, randomized, placebo controlled study. *J Urol* **173**, 1252-1255 (2005).

216 Nickel, J., Barkin, J, Forrest, J, Mosbaugh, PG, Hernandez-Graulau, J, Kaufman, D, Lloyd, K, Evans, RJ, Parsons, CL, Atkinson, LE; Elmiron Study Group. Randomized, double-blind, dose-ranging study of pentosan polysulfate sodium for interstitial cystitis. *Urology* **65**, 654-658 (2005).

217 Nickel, J., Herschorn, S, Whitmore, KE, Forrest, JB, Hu, P, Friedman, AJ, Baseman, AS. Pentosan polysulfate sodium for treatment of interstitial cystitis/bladder pain syndrome: insights from a randomized, double-blind, placebo controlled study. *J Urol* **193**, 857-862 (2015).

218 Senthil, D., Malini, MM, Varalakshmi, P. Sodium pentosan polysulphate--a novel inhibitor of urinary risk factors and enzymes in experimental urolithiatic rats.. *Ren Fail* **20**, 573-580 (1998).

219 Wallius, B., Tidholm, AE. Use of pentosan polysulphate in cats with idiopathic, non-obstructive lower urinary tract disease: a double-blind, randomised, placebo-controlled trial. *J Feline Med Surg* **11**, 409-412 (2009).

220 Baggen, J., Vanstreels, E, Jansen, S, Daelemans, D. Cellular host factors for SARS-CoV-2 infection. *Nat Microbiol* **6**, 1219-1232 (2021).

221 Takizawa, M., Ohuchi, E, Yamanaka, H, Nakamura, H, Ikeda, E, Ghosh, P, Okada, Y. Production of tissue inhibitor of metalloproteinases 3 is selectively enhanced by calcium pentosan polysulfate in human rheumatoid synovial fibroblasts. *Arthritis Rheum* **43**, 812-820 (2000).

222 Takizawa, M., Yatabe, T, Okada, A, Chijiwa, M, Mochizuki, S, Ghosh, P, Okada, Y. Calcium pentosan polysulfate directly inhibits enzymatic activity of ADAMTS4 (aggrecanase-1) in osteoarthritic chondrocytes. *FEBS Lett* **582**, 2945-2949 (2008).

223 Troeberg, L., Fushimi, K, Khokha, R, Emonard, H, Ghosh, P, Nagase, H. Calcium pentosan polysulfate is a multifaceted exosite inhibitor of aggrecanases. *FASEB J* **22**, 3515-3524 (2008).

224 Bwalya, E., Kim, S, Fang, J, Wijekoon, HMS, Hosoya, K, Okumura, M. Pentosan polysulfate inhibits IL-1 β -induced iNOS, c-Jun and HIF-1 α upregulation in canine articular chondrocytes. *PLoS One* **12**, e0177144 (2017).

225 Daly, C., Ghosh, P, Zannettino, ACW, Badal, T, Shimmon, R, Jenkin, G, Oehme, D, Jain, K, Sher, I, Vais, A, Cohen, C, Chandra, RV, Goldschlager, T. Mesenchymal progenitor cells primed with pentosan polysulfate promote lumbar intervertebral disc regeneration in an ovine model of microdiscectomy. *Spine J* **18**, 491-506 (2018).

226 Kumagai, K., Shirabe, S, Miyata, N, Murata, M, Yamauchi, A, Kataoka, Y, Niwa, M. Sodium pentosan polysulfate resulted in cartilage improvement in knee osteoarthritis--an open clinical trial. *BMC Clin Pharmacol* **10**, 7 (2010).

227 Oehme, D., Ghosh, P, Goldschlager, T, Itescu, S, Shimon, S, Wu, J, McDonald, C, Troupis, JM, Rosenfeld, JV, Jenkin, G. Reconstitution of degenerated ovine lumbar discs by STRO-3-positive allogeneic mesenchymal precursor cells combined with pentosan polysulfate. *J Neurosurg Spine* **24**, 715-726 (2016).

228 Andrews, J., Ghosh, P, Lentini, A, Ternai, B. The interaction of pentosan polysulphate (SP54) with human neutrophil elastase and connective tissue matrix components. *Chem Biol Interact* **47**, 157-173 (1983).

229 Suranji Wijekoon, H., Kim, S, Bwalya, EC, Fang, J, Aoshima, K, Hosoya, K, Okumura, M. Anti-arthritis

effect of pentosan polysulfate in rats with collagen-induced arthritis. *Res Vet Sci* **122**, 179-185 (2019).

230 Frith, J., Cameron, AR, Menzies, DJ, Ghosh, P, Whitehead, DL, Gronthos, S, Zannettino, AC, Cooper-White, JJ. An injectable hydrogel incorporating mesenchymal precursor cells and pentosan polysulphate for intervertebral disc regeneration. *Biomaterials* **34**, 9430-9440 (2013).

231 Frith, J., Menzies, DJ, Cameron, AR, Ghosh, P, Whitehead, DL, Gronthos, S, Zannettino, AC, Cooper-White, JJ. Effects of bound versus soluble pentosan polysulphate in PEG/HA-based hydrogels tailored for intervertebral disc regeneration. *Biomaterials* **35**, 1150-1162 (2014).

232 Lohmann, N., Schirmer, L, Atallah, P, Wandel, E, Ferrer, RA, Werner, C, Simon, JC, Franz, S, Freudenberg, U. Glycosaminoglycan-based hydrogels capture inflammatory chemokines and rescue defective wound healing in mice. *Sci Transl Med* **9**, pii: eaai9044 (2017).

233 Kilgore, K., Naylor, KB, Tanhehco, EJ, Park, JL, Booth, EA, Washington, RA, Lucchesi, BR. The semisynthetic polysaccharide pentosan polysulfate prevents complement-mediated myocardial injury in the rabbit perfused heart. *J Pharmacol Exp Ther* **285**, 987-994 (1998).

234 Klegeris, A., Singh, EA, McGeer, PL. . Effects of C-reactive protein and pentosan polysulphate on human complement activation. *Immunology* **106**, 381-388 (2002).

235 Fischer, A., Barrowcliffe, TW, Thomas, DP. . A comparison of pentosan polysulphate (SP54) and heparin. I: Mechanism of action on blood coagulation. *Thromb Haemost* **47**, 104-108 (1982).

236 Giedrojć, J., Radziwon, P, Klimiuk, M, Bielawiec, M, Breddin, HK, Kłoczko, J. Experimental studies on the anticoagulant and antithrombotic effects of sodium and calcium pentosan polysulphate. *J Physiol Pharmacol* **50**, 111-119 (1999).

237 Orme, C., Harris, RC. . A comparison of the lipolytic and anticoagulative properties of heparin and pentosan polysulphate in the thoroughbred horse. *Acta Physiol Scand* **59**, 179-185 (1997).

238 Vinazzer, H. Effect of pentosan polysulfate on fibrinolysis: basic tests and clinical application. *Semin Thromb Hemost* **17**, 375-378 (1991).

239 Goad, K., Horne, MK 3rd, Gralnick, HR. Pentosan-induced thrombocytopenia: support for an immune complex mechanism. *Br J Haematol* **88**, 803-808 (1994).

240 Tardy-Poncet, B., Tardy, B, Grelac, F, Reynaud, J, Mismetti, P, Bertrand, JC, Guyotat, D. Pentosan polysulfate-induced thrombocytopenia and thrombosis. *Am J Hematol* **45**, 252-257 (1994).

241 Verbruggen, G., Veys, EM. Intra-articular injection pentosan polysulphate results in increased hyaluronan molecular weight in joint fluid. *Clin Exp Rheumatol* **10**, 249-254 (1992).

242 Stapledon, C., Tsangari, H, Solomon, LB, Campbell, DG, Hurtado, P, Krishnan, R, Atkins, GJ. Human osteocyte expression of Nerve Growth Factor: The effect of Pentosan Polysulphate Sodium (PPS) and implications for pain associated with knee osteoarthritis. *PLoS One* **14**, e0222602 (2019).

243 Sampson, M., Kabbani, M, Krishnan, R, Nganga, M, Theodoulou, A, Krishnan, J. Improved clinical outcome measures of knee pain and function with concurrent resolution of subchondral Bone Marrow Edema Lesion and joint effusion in an osteoarthritic patient following Pentosan Polysulphate Sodium treatment: a case report. *BMC Musculoskelet Disord* **18**, 396 (2017).

244 Pluda, J., Shay, LE, Foli, A, Tannenbaum, S, Cohen, PJ, Goldspiel, BR, Adamo, D, Cooper, MR, Broder, S, Yarchoan, R. Administration of pentosan polysulfate to patients with human immunodeficiency virus-associated Kaposi's sarcoma. *J Natl Cancer Inst* **85**, 1585-1592 (1993).

245 Alberca, G., Solis-Castro, RL, Solis-Castro, ME, Alberca, RW. . Coronavirus disease-2019 and the intestinal tract: An overview. *World J Gastroenterol* **27**, 1255-1266, doi:doi: 10.3748/wjg.v27.i13.1255. (2021).

246 He, F., Zhang, T, Xue, K, Fang, Z, Jiang, G, Huang, S, Li, K, Gu, Z, Shi, H, Zhang, Z, Zhu, H, Lin, L, Li, J, Xiao, F, Shan, H, Yan, R, Li, X, Yan, Z. . Fecal multi-omics analysis reveals diverse molecular

alterations of gut ecosystem in COVID-19 patients. *Anal Chim Acta* **1180**, 338881, doi:doi: 10.1016/j.aca.2021.338881 (2021).

247 Zhang, B., Zhong, Y., Dong, D., Zheng, Z., Hu, J. Gut microbial utilization of xylan and its implication in gut homeostasis and metabolic response. *Carbohydr Polym* **286**, 119271 (2022).

248 Lee JY, T. R., Bäumler AJ. The microbiome and gut homeostasis. *Science* **377**, eabp9960 (2022).

249 Xiao, Z., Pan, M., Li, X., Zhao, C. Impact of SARS-CoV2 infection on gut microbiota dysbiosis. *Microbiome Res Rep* **3**, 7 (2023).

250 Nordberg Karlsson, E., Schmitz, E., Linares-Pastén, JA, Adlercreutz, P. Endo-xylanases as tools for production of substituted xylooligosaccharides with prebiotic properties. *Appl Microbiol Biotechnol* **102**, 9081-9088 (2018).

251 Zhang, M., Chekan, JR, Dodd, D., Hong, PY, Radlinski, L., Revindran, V., Nair, SK, Mackie, RI, Cann, I. Xylan utilization in human gut commensal bacteria is orchestrated by unique modular organization of polysaccharide-degrading enzymes. *Proc Natl Acad Sci U S A* **111**, E3708-3717 (2014).

252 Childs, C., Röytiö, H., Alhoniemi, E., Fekete, AA, Forssten, SD, Hudjec, N., Lim, YN, Steger, CJ, Yaqoob, P., Tuohy, KM, Rastall, RA, Ouwehand, AC, Gibson, GR. Xylo-oligosaccharides alone or in symbiotic combination with *Bifidobacterium animalis* subsp. *lactis* induce bifidogenesis and modulate markers of immune function in healthy adults: a double-blind, placebo-controlled, randomised, factorial cross-over study. *Br J Nutr* **111**, 1945-1956 (2014).

253 Gesaka, S., Obimbo, MM, Wanyoro, A. . Coronavirus disease 2019 and the placenta: A literature review. *Placenta* **126**, 209-223 (2022).

254 Ojha, R., Gurjar, K., Ratnakar, TS, Mishra, A., Prajapati, VK. . Designing of a bispecific antibody against SARS-CoV-2 spike glycoprotein targeting human entry receptors DPP4 and ACE2. *Hum Immunol* **83**, 346-355 (2022).

255 Chaves-Medina, M., Gómez-Ospina, JC, García-Perdomo, HA. . Molecular mechanisms for understanding the association between TMPRSS2 and beta coronaviruses SARS-CoV-2, SARS-CoV and MERS-CoV infection: scoping review. *Arch Microbiol* **204**, 77 (2021).

256 Jacobs, J., Booth, GS. COVID-19 and Immune-Mediated RBC Destruction. *Am J Clin Pathol* **157**, 844-851 (2022).

257 Wendisch, D., Dietrich O, Mari T, von Stillfried S, Ibarra IL, Mittermaier M, Mache C, Chua RL, Knoll R, Timm S, Brumhard S, Krammer T, Zauber H, Hiller AL, Pascual-Reguant A, Mothes R, Bülow RD, Schulze J, Leipold AM, Djedjaj S, Erhard F, Geffers R, Pott F, Kazmierski J, Radke J, Pergantis P, Baßler K, Conrad C, Aschenbrenner AC, Sawitzki B, Landthaler M, Wyler E, Horst D; Deutsche COVID-19 OMICS Initiative (DeCOI), Hippenstiel S, Hocke A, Heppner FL, Uhrig A, Garcia C, Machleidt F, Herold S, Elezkurtaj S, Thibeault C, Witzenrath M, Cochaint C, Suttorp N, Drosten C, Goffinet C, Kurth F, Schultze JL, Radbruch H, Ochs M, Eils R, Müller-Redetzky H, Hauser AE, Luecken MD, Theis FJ, Conrad C, Wolff T, Boor P, Selbach M, Saliba AE, Sander LE. SARS-CoV-2 infection triggers profibrotic macrophage responses and lung fibrosis. *Cell* **184**, 6243-6261.e6227 (2021).

258 Acharya, Y., Alameer, A., Calpin, G., Alkhattab, M., Sultan, S. . A comprehensive review of vascular complications in COVID-19. *J Thromb Thrombolysis* **53**, 586-593 (2022).

259 Veizades, S., Tso, A., Nguyen, PK. . Infection, inflammation and thrombosis: a review of potential mechanisms mediating arterial thrombosis associated with influenza and severe acute respiratory syndrome coronavirus 2. *Biol Chem* **403**, 231-241 (2021).

260 Plášek, J., Gumulec, J., Máca, J., Škarda, J., Procházka, V., Grézl, T., Václavík, J. COVID-19 associated coagulopathy: Mechanisms and host-directed treatment. *Am J Med Sci* **363**, 465-475 (2022).

261 Rizal, S., Joshi, BR, Regmi, S. . Raised D-dimer Level among COVID-19 Patients in a Tertiary Care Hospital: A Descriptive Cross-sectional Study. *JNMA J Nepal Med Assoc* **60**, 259-262 (2022).

262 Al-Kuraishy, H., Al-Gareeb, Al, Kaushik, A, Kujawska, M, Batiha, GE. Hemolytic anemia in COVID-19. *Ann Hematol* **101**, 1887-1895 (2022).

263 Wagener, F., Pickkers, P, Peterson SJ, Immenschuh, S, Abraham, NG. Targeting the Heme-Heme Oxygenase System to Prevent Severe Complications Following COVID-19 Infections.. *Antioxidants (Basel)* **9**, 540 (2020).

264 Ousaka, D., Nishibori, M. Is hemolysis a novel therapeutic target in COVID-19? *Front Immunol* **13**, 956671 (2022).

265 Natale, N., Lukens, JR, Petri, WA Jr. The nervous system during COVID-19: Caught in the crossfire. *Immunol Rev* doi: [10.1111/imr.13114](https://doi.org/10.1111/imr.13114). (2022).

266 Valdes, E., Fuchs, B, Morrison, C, Charvet, L, Lewis, A, Thawani, S, Balcer, L, Galetta, SL, Wisniewski, T, Frontera, JA. Demographic and social determinants of cognitive dysfunction following hospitalization for COVID-19. *J Neurol Sci* **438**, 120146 (2022).

267 Meyer, P., Hellwig, S, Blazhenets, G, Hosp, JA. Molecular Imaging Findings on Acute and Long-Term Effects of COVID-19 on the Brain: A Systematic Review. *J Nucl Med* **63**, 971-980 (2022).

268 Kao, J., Frankland, PW. COVID fog demystified. *Cell* **S0092-8674**, 00725-00725 (2022).

269 Maltezou, H., Pavli, A, Tsakris, A. . Post-COVID Syndrome: An Insight on Its Pathogenesis. *Vaccines (Basel)* **9**, 497 (2021).

270 García-Grimshaw, M., Sankowski, R, Valdés-Ferrer, SI. . Neurocognitive and psychiatric post-coronavirus disease 2019 conditions: pathogenic insights of brain dysfunction following severe acute respiratory syndrome coronavirus 2 infection. *Curr Opin Neuro* **35**, 375-383 (2022).

271 Anonymous. Long COVID and kids: more research is urgently needed. *Nature* **602**, 183 (2022).

272 Ortona, E., Malorni, W. Long COVID: to investigate immunological mechanisms and sex/gender related aspects as fundamental steps for tailored therapy. *Eur Respir J* **59**, 2102245 (2022).

273 Schober, M., Robertson, CL, Wainwright, MS, Roa, JD, Fink, EL. COVID-19 and the Pediatric Nervous System: Global Collaboration to Meet a Global Need. *Neurocrit Care* **35**, 283-290 (2021).

274 Deoni, S., Beauchemin, J, Volpe, A, Dâ Sa, V; RESONANCE Consortium. Impact of the COVID-19 Pandemic on Early Child Cognitive Development: Initial Findings in a Longitudinal Observational Study of Child Health. *medRxiv [Preprint]*. [08.10.21261846](https://doi.org/10.1101/21261846) (2021).

275 Dyer, O. Covid-19: Children born during the pandemic score lower on cognitive tests, study finds. *BMJ*. 2021 Aug 16;374:n2031. Dyer O. Covid-19: Children born during the pandemic score lower on cognitive tests, study finds. *BMJ* **374**, n2031 (2021).

276 Ehrler, M., Werninger, I, Schnider, B, Eichelberger, DA, Naef, N, Disselhoff, V, Kretschmar, O, Hagmann, CF, Latal, B, Wehrle, FM. Impact of the COVID-19 pandemic on children with and without risk for neurodevelopmental impairments. *Acta Paediatr* **110**, 1281-1288 (2021).

277 Liu, K., Howard, R, Banerjee, S, Comas-Herrera, A, Goddard, J, Knapp, M, Livingston, G, Manthorpe, J, O'Brien, JT, Paterson, RW, Robinson, L, Rossor, M, Rowe, JB, Sharp, DJ, Sommerlad, A, Suárez-González, A, Burns, A. Dementia wellbeing and COVID-19: Review and expert consensus on current research and knowledge gaps. *Int J Geriatr Psychiatry* **36**, 1597-1639 (2021).

278 Hampshire, A., Trender, W, Chamberlain, SR, Jolly, AE, Grant, JE, Patrick, F, Mazibuko, N, Williams, SC, Barnby, JM, Hellyer, P, Mehta, MA. Cognitive deficits in people who have recovered from COVID-19. *EClinicalMedicine* **39**, 101044 (2021).

279 Palakshappa, J., Krall, JTW, Belfield, LT, Files, DC. Long-Term Outcomes in Acute Respiratory Distress Syndrome: Epidemiology, Mechanisms, and Patient Evaluation. *Crit Care Clin* **37**, 895-911 (2021).

280 Schou, T., Joca, S, Wegener, G, Bay-Richter, C. Psychiatric and neuropsychiatric sequelae of COVID-19 - A systematic review. *Brain Behav Immun* **97**, 328-348 (2021).

281 Yang, F., Zhao, H, Liu, H, Wu, X, Li, Y. . Manifestations and mechanisms of central nervous

system damage caused by SARS-CoV-2. *Brain Res Bull* **S0361-9230**, 00284-00287 (2021).

282 Yong, S. Long COVID or post-COVID-19 syndrome: putative pathophysiology, risk factors, and treatments. *Infect Dis (Lond)* **53**, 737-754 (2021).

283 Aiyegbusi, O., Hughes, SE, Turner, G, Rivera, SC, McMullan, C, Chandan, JS, Haroon, S, Price, G, Davies, EH, Nirantharakumar, K, Sapey, E, Calvert, MJ; TLC Study Group. Symptoms, complications and management of long COVID: a review. *J R Soc Med* **114**, 428-442 (2021) (2021).

284 Chen, J., Zhang, W, Li, Y, Liu, C, Dong, T, Chen, H, et al Bat-infecting merbecovirus HKU5-

CoV lineage 2 can use human ACE2 as a cell entry receptor. *Cell* **188**, 1729-1742.e1716. doi: 1710.1016/j.cell.2025.1701.1042. (2025).

285 Srivastava, S., Sharma, D, Sridhar, SB, Kumar, S, Sahu, R, Ashique, S, et al HKU5-CoV-2: A silent threat looming in bats. *Travel Med Infect Dis* **65**, 102838. doi: 102810.101016/j.tmaid.102025.102838. (2025).

286 Wang, N., Ji, W, Jiao, H, Veit ,M, Sun, J, Wang, Y, et al A MERS-CoV-like mink coronavirus uses ACE2 as an entry receptor. *Nature* **642**, 739-746. doi: 710.1038/s41586-41025-09007-w. (2025).

Abbreviations used:

ACE2 Angiotensin-converting enzyme 2
ADAMTS A Disintegrin and Metalloproteinase with Thrombospondin motifs
AD Alzheimer's disease
ADHD Attention deficit hyperactivity disorder
ARDS Acute respiratory distress syndrome
AT Antithrombin
BSG/CD147 Basigin
BGN Biglycan
BMP Bone morphogenetic protein
CendR C-end Rule protein motif, interacts with NRP-1
CoV Coronavirus
COVID Coronavirus disease 2019
CoV RaTG13 Bat coronavirus RaTG13
CVI Chronic venous insufficiency
CSF Cerebrospinal fluid
DPP4/CD26 Dipeptidyl peptidase-4
DCN Decorin
DMOADDisease modifying anti-arthritis drug
DVT Deep vein thrombosis
ECM Extracellular matrix protein
ERK1/2 Extracellular signal-regulated kinase 1 and 2
GPC Glypican
HS Heparan sulfate
HSPG Heparan sulfate proteoglycan
HUVEC Human umbilical vein endothelial cells
IAV Influenza A virus
IFTM1-3 Interferon-induced transmembrane protein
IL Interleukin
iNOS Inducible nitric oxide synthase
IQ Intelligence quotient
ITI Inter--trypsin inhibitor
ITIM Immunoreceptor tyrosine-based inhibitory motif
KIAA1199 *CEMIP* (*cell migration inducing protein*) deafness gene
LDL Low density lipoprotein
LMW-HA Low molecular weight hyaluronan
MAPK Mitogen-activated protein kinase
MERS Middle East respiratory syndrome
MSCs Mesenchymal stromal stem cells
NDST-2 *N*-Deacetylase/*N*-Sulfotransferase-2
NF-κB Nuclear factor kappa-light-chain-enhancer of activated B cells
NGF Nerve growth factor
NMJ Neuromuscular junction
NRP-1 Neuropilin-1
NTD N-terminal domain (of Spike protein)

OA, RA Osteoarthritis, Rheumatoid arthritis
OCD Obsessive compulsive disorder
PAI-1, 2 Plasminogen activator inhibitor-1, 2
PCSK9 Proprotein convertase subtilisin/kexin type 9
PET Positron emission tomography
PPS Pentosan polysulfate
RBD Receptor binding domain (of Spike protein)
ROS Reactive Oxygen species
SARS CoV-2 *Severe acute respiratory syndrome coronavirus-2*
S Spike protein of SARS CoV-2
SDC Syndecan
SPECT Single-photon emission computed tomography
TAZ Transcriptional co-activator with PDZ binding motif
TLR4 Toll-like receptor-4
TMPRSS2 Transmembrane protease, serine 2
TNF Tumor necrosis factor alpha
tPA Tissue plasminogen activator
TRAF-6 Tumor necrosis factor receptor associated factor 6
TS Tourette Syndrome
TSG-6 Tumor necrosis factor-inducible gene 6 protein
YAP Yes associated protein