



NARRATIVE REVIEW

How Does Fetal Acidosis Result in Cerebral Palsy?

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ABSTRACT

Cerebral palsy presents enormous challenges to the physical and emotional wellbeing of the affected individual as well as the immediate family. The average lifetime cost per person with cerebral palsy exceeds one million dollars in the United States. While highly dependent on the degree of severity, children severely impaired in cognition, motor skills, hearing, and vision have a markedly shortened life span. Whereas hypoxia-ischemia is considered to be etiologic in fetal/newborn cerebral palsy, the cellular mechanisms by which acidosis, hypoxemia and hypercapnia induce neuronal damage are complex. Questions remain as to why most newborns survive intact despite severe metabolic acidosis (base deficit greater than 12 mmol/L) and whether early newborn biochemical correction of metabolic acidosis may reduce long term injury. This review aims to address the putative pathways by which acidosis, hypoxemia and hypercapnia may result in cell damage. A knowledge of these mechanisms is essential for the development of therapeutic interventions for the prevention of cerebral palsy in at risk newborns.

Acidosis and Cell Dysfunction:

Acidosis causes cell dysfunction mainly by disturbing protein structure, ion gradients, and energy metabolism, which then impair signaling, contraction, and survival. The tertiary conformation of proteins and enzymes are dependent on protonation of amino acids. Amino acids with basic side chains, (lysine, arginine and histidine) are protonated at physiological pH, while acidic amino acids aspartate and glutamate are protonated in low pH environments. With changes in optimal protein conformation, enzyme activity is often reduced. These findings are of heightened significance in regards to energy homeostasis, as low pH impairs mitochondrial respiratory capacity and thus ATP production⁴. Subsequently, cell ATP-dependent ion pump dysfunction may result in a loss of cell membrane potential and volume regulation, and ultimately cell death⁴. Specifically, reduced extracellular pH inhibits neuronal voltage-gated and ligand-gated ion channels⁵, allowing calcium and sodium to enter the cells. The resulting cell depolarization and neuronal excitation activates cellular injury pathways⁶. In addition to ion channel effects, downstream protein binding to acid-sensing ion channels (ASIC) activates cell death pathways (apoptosis, necroptosis). Ironically, a critical component of the neuronal cell death pathway is a serine/threonine kinase receptor interaction protein 1, which is aptly named RIP1.

Intracellular acidosis also interferes with many pH-sensitive processes, including metabolism, motility, immune function and growth, in multiple cell types (e.g., neurons, astrocytes, colon, skeletal muscle, macrophage)⁷. Specifically impacted pathways which regulate these functions include MAPK⁸, PI3K-Akt-mTOR⁹ as well as TRAIL-death receptor signaling¹⁰. Thus, prolonged acidosis may shift cells from adaptation toward apoptosis or necrosis. Without question acidosis itself is a putative neural injury factor.

Hypoxia and Cell Dysfunction:

Similar to metabolic acidosis, hypoxia causes cell dysfunction primarily by limiting mitochondrial ATP production, altering gene expression, disturbing ion homeostasis, and activating cell death pathways¹¹. Hypoxia primarily injures cells through oxygen lack and energy failure, while metabolic acidosis mainly alters pH-sensitive proteins, ion transport, and

signaling, even when oxygen is present¹². The mechanism of response to hypoxia involves hypoxia inducible factors (HIFs)¹³, which are stabilized by low oxygen availability and control the expression of a multitude of genes, including those involved in cell survival, angiogenesis, and glycolysis.

If persistent and/or severe, hypoxia may produce secondary metabolic acidosis via anaerobic glycolysis and lactate accumulation; thus, many hypoxic injuries combine energy failure with pH-mediated enzyme and ion-transport disturbances (see above). However, mild acidosis may transiently reconfigure mitochondrial efficiency to maintain mitochondrial function and cell survival¹⁴. Thus, mild extracellular acidosis actually may be a protective agent¹⁵, but severe or prolonged acidosis worsens dysfunction and promotes cell death¹⁴.

Hypercapnia (Elevated CO₂) and Cell Dysfunction:

Although fetal/newborn metabolic acidosis is often accompanied by hypercapnia, the biochemical and physiologic consequences of hypercapnia, as well as the interaction with acidosis and hypoxia, are far less studied than that of acidosis or hypoxemia. Whereas metabolic acidosis disturbs enzyme activity, ion gradients and muscle contractility, hypercapnia changes cell function via both CO₂ itself and via respiratory acidosis, as CO₂ hydration to carbonic acid causes respiratory acidosis, lowering intra- and extracellular pH. As compared to modest physiologic or pathologic increases in pCO₂ under conditions of hypoventilation or increased CO₂ production in children and adults, fetal/newborn umbilical artery blood may demonstrate far greater pCO₂ levels, at times exceeding 100 mmHg. Adverse labor events resulting in absent umbilical placental clearance (e.g., complete cord compression, total abruption, uterine rupture) results in an increase of pCO₂ of ~7 mmHg per minute¹⁶. As metabolic acidosis (i.e., base deficit) increases ~0.5 mmol/L per minute under these conditions¹⁶, newborns with severe acidosis most often present with a mixed respiratory and metabolic acidosis. The preferential diffusion of bicarbonate ion (resulting from CO₂ hydration) as compared to H⁺ from the blood to the extracellular compartment mandates the accurate calculation of base deficit with

correction for this extracellular exchange (Base Deficit extracellular fluid)^{17,18}.

Similar to metabolic acidosis, elevated CO₂ reduces mitochondrial oxygen consumption, membrane potential, and electron transport chain activity, leading to lower ATP output from oxidative phosphorylation^{19,20}. Elevated CO₂ also may downregulate metabolic pathway enzymes, blunting hypoxia-driven upregulation of glycolytic enzymes and glucose transporters²¹.

Criteria for acute hypoxia induced encephalopathy, and thus risk for cerebral palsy (CP), are commonly defined as umbilical artery or newborn pH <7.0 and/or base deficit ≥ 12 mmol/L²². Metabolic acidosis is much more strongly linked to fetal brain injury than isolated respiratory acidosis from hypercapnia alone, though the pH criteria suggests that respiratory acidosis may be contributive. A 10 minute interruption of umbilical blood flow in a previously normal fetus would predictably increase umbilical artery pCO₂ to ~120 mmHg, but only increase base deficit to levels of 7 to 9 mmol/L, values below the base deficit threshold for HIE injury. This would result in an umbilical artery pH ~6.91, placing the infant within the high risk category based upon pH.

There is limited data regarding the risk for CP based on predominant respiratory acidosis with sub-injury BD threshold values, as the occurrence is uncommon. Tuuli et al demonstrated that umbilical cord arterial lactate (as a measure of metabolic acidosis) was a more discriminating measure of term infant neonatal morbidity than was pH²³, though other studies indicate that umbilical cord arterial lactate/base excess and pH predict short-term neonatal outcomes with similar efficacies^{24,25}.

In view of the diverse metabolic effects, the duration of metabolic and/or respiratory acidosis potentially also may impact on the risk of neonatal metabolic injury. Umbilical cord artery hypercarbia is typically corrected within minutes of life with adequate neonatal ventilation. In contrast, newborns do not significantly clear metabolic acidosis until beyond 1 ½ to 2 hours²⁶, due to immaturity of hepatic and renal mechanisms at birth. Thus, the cellular effects of metabolic acidosis are likely to impact far longer than that of acute hypercarbia. Importantly, efforts to chemically normalize metabolic acidosis (e.g., bicarbonate

administration) have not demonstrated efficacy, but may, in fact, result in adverse effects^{27,28,29}. For example, intravenous alkali infusions to acidotic dogs induced cerebral vasoconstriction and reduced cerebral blood flow³⁰. However, with ischemic stroke-induced brain acidosis, treatment with NaHCO₃ significantly reduced infarct volume³¹.

Despite the potential cellular effects, it is likely that severe metabolic acidosis creates a markedly greater risk of neurologic injury than does hypercapnia. However, there may well be an interplay between the effects of hypoxia, metabolic acidosis and respiratory acidosis.

Brain Regions Impacted by Fetal Hypoxemia and Acidosis

Classic patterns of fetal hypoxic-ischemic brain damage have differentiated Acute-Profound hypoxia patterns from that of Partial-Prolonged hypoxic damage. The former, a consequence of sudden sentinel events (e.g., abruptio, cord prolapse, uterine rupture) often cause symmetric injury to the basal ganglia and thalami (deep gray nuclei) and sometimes brainstem involvement. In contrast, Partial-Prolonged events often damage parasagittal watershed regions between major cerebral arteries, with relative sparing of deep nuclei^{32,33}. Depending upon the severity and mechanism of injury (fetal hemorrhage vs hypoxia-ischemia) other affected regions may include the hippocampus, brainstem and cerebellum. (Figure 2)

Both these patterns of injury may result in neonatal encephalopathy and ultimately cerebral palsy, due to the motor nerve pathways from cortical gray matter to the deep gray nuclei. Similarly, preterm infant periventricular white matter damage due to hypoxemia or intraventricular hemorrhage may result in similar motor dysfunction due to oligodendrocyte and nerve fiber damage. Maturation of cell types and circuits (oligodendrocytes, excitatory synapses) contribute to the changes in the spatial pattern of injury across gestation³⁴.

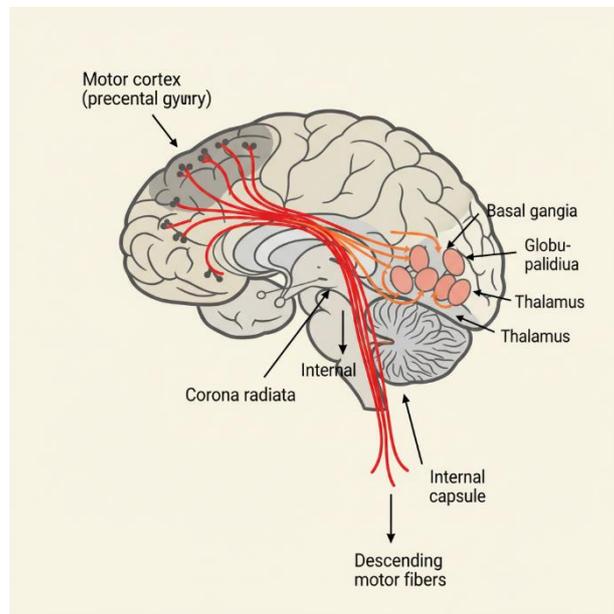


Figure 2: Patterns of motor fibers from the motor cortex to deep gray matter

The susceptibility of the basal ganglia to acute hypoxemia is believed to be due to the high metabolic demand, dense excitatory input, and vascular features that limit their reserve during sudden drops in oxygen or perfusion³³. Having a high resting metabolic rate, the basal ganglia depend on continuous oxidative ATP production; even brief interruptions in oxygen may rapidly exhaust energy stores³⁵. The basal ganglia neurons receive massive glutamatergic input from cortex and thalamus; during hypoxia–ischemia, energy failure impairs glutamate reuptake and increases synaptic glutamate, over activating NMDA/AMPA receptors with the excitotoxic cascade leading to Ca⁺⁺ overload³⁶.

Watershed regions lie at the junction of two of the three major cerebral arterial territories (anterior, middle and posterior). As such, being at the farthest, lowest-pressure ends of arterial territories and having the poorest collateral supply, sustained decreases in oxygen delivery (perfusion or oxygenation) may drop below thresholds necessary to prevent injury³⁷.

Acidosis Thresholds for Cerebral Palsy

As noted above, to attribute hypoxic-ischemic encephalopathy and cerebral palsy (CP) to an intrapartum event requires values of umbilical artery or newborn pH <7.0 and/or base deficit >12 mmol/L, or both. As pH is an inverse log of the hydrogen ion concentration, the relation with the degree of acidosis is logarithmic. Linear changes in umbilical artery pH are of limited value in

predicting the severity of injury, as pH combines the degree of metabolic acidosis with the often transient respiratory acidosis. In contrast, base deficit is a linear measure of the degree of metabolic acidosis, and base deficit extracellular fluid corrects for the effect of CO₂ levels. Base deficit measures indicate a linear or exponential correlation of the degree of metabolic acidosis with the risk of neonatal death, though a threshold phenomenon for the risk of cerebral palsy³⁸. Among infants with severe acidosis, the rate of death (up to 2 years of age) increased from 5% at BD 12-15.9 mmol/L, to 10% at 16-19.9 mmol/L, to 26% at BD >20 mmol/L. In contrast, among infants with a BD of 12-15.9 mmol/L, the rate of cerebral palsy was 2.1%, which did not increase significantly (4.0%) among those with BD 16-19.9 mmol/L, while there was a marked increase (33%) in the rate of cerebral palsy at >20 mmol/L³⁸.

The apparent threshold levels of 12 and 20 mmol/L may be reflective of the complex mechanisms by which acidosis results in neurologic injury as well as the contribution from fetal/newborn cardiovascular impairment (i.e., increased ischemia) which occurs at the highest levels of acidosis³⁹.

Therapeutic Approaches to Newborn Hypoxic Injury:

Certainly, prevention of fetal hypoxic-ischemic injury prenatally or intrapartum is the ultimate objective. However, should severe acidosis result, current pediatric care approaches have had significant success in preventing or reducing the

degree of damage. Certainly, quality resuscitation and stabilization may limit the hypoxic event. Therapeutic hypothermia (whole-body or head cooling) of nearterm or term infants with moderate–severe hypoxic-ischemic encephalopathy reduces death and severe disability and lowers rates of cerebral palsy compared with normothermia^{40,41}. Therapeutic hypothermia efficacy is focused on interrupting the delayed injury cascade that unfolds hours to days after the initial asphyxial event. Cooling the brain reduces cerebral metabolic rate, helping preserve ATP and delay or blunt the secondary energy failure that normally peaks 12–48 hours after hypoxia–ischemia⁴². Hypothermia also dampens glutamate release, NMDA/AMPA receptor activation, and downstream calcium-dependent enzyme activation, thereby limiting excitotoxic damage in vulnerable regions⁴³. Cooling also lowers production of reactive oxygen and nitrogen species and stabilizes mitochondrial membranes, attenuates microglial activation and pro-inflammatory cytokine release, and interferes with apoptotic pathways⁴⁴.

Pharmacologic adjuncts to therapeutic hypothermia have attempted to address the multiple pathways of neuronal damage induced by hypoxia, acidosis and hypercapnia, including energy production and utilization, excitatory stimulation, inflammatory cytokines, reactive oxygen species and apoptotic pathways. Studies of agents including erythropoietin, melatonin, allopurinol, magnesium sulfate, xenon, topiramate, and mesenchymal stem cells show potential promise, but demonstrated benefit has not been confirmed^{45,41}.

Conclusion

Fetal/newborn hypoxic-ischemic brain injury is a consequence of direct cellular effects of hypoxia coupled with secondary effects of metabolic acidosis and, potentially, that of hypercapnia. The complexity of mechanisms inducing neuronal damage include altered protein structure, abnormal ion gradients, reduced cellular energy metabolism, increased excitatory neuronal signals, inflammatory cytokines, activation of apoptotic pathways, and reoxygenation injury. Major questions remain unanswered, including what factors predict infant tolerance/susceptibility to severe metabolic acidosis, the relative impact of acidosis vs. hypoxemia and hypercapnia, and

whether persistence of acidosis during the newborn period impacts cellular injury. Whereas therapeutic hypothermia likely acts broadly to reduce cellular energy requirements and suppress damage pathways, the identification of these putative damage pathways indicates opportunities for therapeutic pharmacologic targets.

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