



REVIEW ARTICLE

Regulation of Cystathionine γ -Lyase Expression in the Cardiovascular System: Insights into Exogenous Hydrogen Sulfide, Hydrogen Peroxide, Hypoxia, and Nuclear Factor κ B Signaling

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ABSTRACT

Cystathionine γ -lyase is a key enzyme in the transsulfuration pathway responsible for endogenous hydrogen sulfide production in the cardiovascular system. As the third gaseous signaling molecule, hydrogen sulfide plays crucial roles in maintaining vascular homeostasis, regulating vasodilation, and protecting against ischemia-reperfusion injury. This review comprehensively analyzes the regulatory mechanisms governing cystathionine γ -lyase expression under various physiological and pathological conditions. Exogenous hydrogen sulfide exhibits concentration-dependent bidirectional regulation of cystathionine γ -lyase expression, with lower concentrations (10-80 μ M) suppressing cystathionine γ -lyase through feedback inhibition, while higher concentrations (120-160 μ M) upregulating its expression as a protective response. Hydrogen peroxide, at moderate concentrations (5 μ M), significantly enhances cystathionine γ -lyase promoter activity and mRNA/protein expression, suggesting a potential feedback loop where cystathionine γ -lyase-derived hydrogen sulfide scavenges reactive oxygen species. Hypoxia regulates cystathionine γ -lyase through transcriptional and post-transcriptional mechanisms, with increased cystathionine γ -lyase expression potentially protecting cells by elevating hydrogen sulfide levels and buffering oxygen consumption. Furthermore, lipopolysaccharide-induced cystathionine γ -lyase expression critically depends on the Nuclear Factor κ B transcription factor binding site (GGACATTCC) within the cystathionine γ -lyase promoter, establishing a direct link between inflammatory signaling and hydrogen sulfide biosynthesis. Based on these findings, we propose a mechanistic hypothesis wherein hypoxia-induced cardiomyocyte apoptosis releases hydrogen peroxide, which activates Nuclear Factor κ B signaling in vascular endothelial cells to upregulate cystathionine γ -lyase expression, leading to enhanced hydrogen sulfide production and subsequent vasodilation. Understanding these regulatory networks provides theoretical foundations for developing therapeutic strategies targeting the cystathionine γ -lyase/hydrogen sulfide pathway in cardiovascular diseases, including myocardial infarction, hypertension, and atherosclerosis.

Keywords: Cystathionine γ -lyase; Hydrogen sulfide; Hypoxia; Nuclear Factor κ B; Cardiovascular regulation

1. Introduction

Hydrogen sulfide (H₂S), a well-known toxic gas, has recently been recognized as the third endogenous gaseous signal molecule in recent years. Cystathionine γ -lyase (CSE), one of the three enzymes in the transsulfuration pathway, is responsible for synthesizing endogenous H₂S using L-cysteine or L-homocysteine as a substrate^{1,2}. The production of CSE mRNA must occur in the rat aorta in association with H₂S provided with exogenous L-cysteine. Typically, cystathionine β -synthase (CBS) plays a crucial role in developing and maintaining the central nervous system, and the radial glia/astrocyte dysfunction may be involved in the complex neuropathological features³. Moreover, 3-mercaptopyruvate sulfurtransferase (3MST), another H₂S-producing enzyme, is localized to brain neurons and the vascular endothelium⁴. The vascular smooth muscle cells (SMCs) from the CSE gene knockout mice are more susceptible to apoptosis induced by exogenous H₂S at physiologically relevant concentrations⁵. H₂S-induced S-sulfhydration on proteins are potential novel targets for therapeutic intervention and drug design for the skin, which may lead to the development and application of H₂S-related drugs for dermatological diseases⁶. Since H₂S has been explored to exhibit a wide range of physiologic functions to maintain vascular homeostasis, it is not surprising that H₂S may play beneficial effects in the progression of atherosclerosis⁷. In-depth research on the regulation of CSE gene expression can not only clarify the molecular mechanism of the occurrence of related diseases but also discover how different exogenous factors regulate CSE gene expression, and it is likely to find the molecular mechanism of the occurrence of related diseases and current targets of drug action.

2. Regulation of Cystathionine γ -lyase Expression by Exogenous Hydrogen Sulfide

The protective effect of exogenous H₂S at low concentrations (30 or 50 μ M) on rat aortic SMCs against cytotoxicity and damage induced by homocysteine has been demonstrated, resulting in enhanced cell viability⁸. Conversely, upregulation of CSE leads to SMC apoptosis due to an elevation in endogenous H₂S production⁹. Moreover, mice with cardiac overexpression of CSE exhibit significant resistance against ischemia-reperfusion injury. This protection is accompanied by reduced myocardial inflammation and preserved mitochondrial function¹⁰. After suppressing the endogenous background expression of CSE, direct administration of exogenous H₂S at a concentration of 100 μ M can induce apoptosis in human aortic SMCs. The activity of CSE has been reported to be regulated by Calmodulin in the presence of 1-2 mM Ca²⁺¹¹. Vascular SMCs from CSE gene knockout mice are more susceptible to apoptosis induced by physiologically relevant concentrations of exogenous H₂S⁵. Treatment with low levels of exogenous H₂S (\leq 100 μ M) can protect cells from radiation damage¹². However, further investigation is necessary to conclusively determine whether CSE is regulated by Ca²⁺/Calmodulin². GYY4137 (a slow-releasing H₂S donor) may serve as a novel therapeutic tool for preventing diabetes-associated vascular dysfunction¹³. Modulating H₂S signaling represents a potential novel therapeutic approach for managing hypertension; however, additional experimental and clinical studies on the role of H₂S in hypertension are required¹⁴.

Cystathionine γ -lyase mainly mediates the endogenous generation of H₂S in the cardiovascular system¹⁵. After inhibiting endogenous background CSE expression, the direct administration of 100 μ M exogenous H₂S can induce the apoptosis of human aorta smooth muscle cells⁹. Mice overexpressing CSE in the heart exhibit resistance to ischemia-reperfusion

injury, and this protection is accompanied by a decrease in myocardial inflammation¹⁰. Vascular smooth muscle cells (VSMCs) from CSE gene-knockout mice are more susceptible to apoptosis induced by exogenous H₂S at physiologically relevant concentrations than those from wild-type mice⁵. Hydrogen sulfide is synthesized within the vasculature and facilitates vascular homeostasis, vasodilation, and endothelial cell proliferation¹⁶. Elevated levels of homocysteine induce dysfunction in endothelial cells, while the metabolic and physiological functions of H₂S enable it to function as a protective agent¹⁷. Hydrogen sulfide is recognized as both a signaling molecule and cytoprotectant that safeguards various tissues and organs against oxidative stress and ischemia-reperfusion injury¹⁸. Endogenous H₂S plays modulatory roles in hypoxia-induced cardiovascular responses by inhibiting spontaneously hypertensive (SH) rats¹⁹. The production of H₂S primarily through endothelial CSE contributes to cardiovascular homeostasis²⁰.

Several studies have investigated the effects of H₂S in human vessels. Hydrogen sulfide-induced relaxation has been demonstrated in internal mammary²¹, pulmonary²², mesenteric²³, and intrarenal arteries²⁴ as well as in perfused human placentas²⁵. Upregulation of CSE expression during hypoxia may increase the production and concentration of H₂S in cells and protect cells from hypoxia²⁶. A controlled release formulation of S-propargyl-cysteine exerted protective effects against myocardial infarction (MI) via the CSE/H₂S pathway²⁷. Nicotinamide adenine dinucleotide phosphate (NADPH) oxidase 4 is a positive transcriptional regulator of CSE in endothelial cells, and some researchers propose that it may modulate the production of endogenous H₂S²⁸. Cystathionine γ -lyase-derived H₂S production by endothelial cells is critical for maintaining endothelial function and exercise capacity and protecting against myocardial ischemia/reperfusion injury²⁹.

An in vitro study showed that an exogenous H₂S donor attenuated hypoxia-induced apoptosis in primary rat nucleus pulposus (NP) cells³⁰. Hydrogen sulfide has been identified as an excitatory mediator of hypoxic sensing in carotid bodies³¹. Incubation with sodium hydrosulfide (NaHS), an H₂S donor, increased the expression of miR-21 and attenuated the reduced cell viability and increased apoptosis caused by ischemia-reperfusion (I/R) in Buffalo rat liver (BRL) cells³². The exogenous administration of NaHS might be a potential strategy for treating nickel-induced lung cancer progression³³. Pretreatment with NaHS or aspirin (ATB-340) in aged rats fed a high-fructose diet (HFD) and animals exposed to water-immersion restraint stress (WIRS) attenuated gastric damage compared to vehicle treatment³⁴.

Some studies have demonstrated that appropriate levels of exogenous H₂S can impact the regulation of cystathionine gamma-lyase (CSE) expression. The application of exogenous H₂S at concentrations of 50, 100, and 200 μ M can reduce the action potential duration in healthy papillary muscles. Moreover, pretreatment with glibenclamide partially attenuates the effects induced by 100 μ M exogenous H₂S³⁵. Exposing mammalian cells to exogenous H₂S within a range of 10-80 μ M downregulates both transcription and expression levels of CSE. Conversely, an exposure to exogenous H₂S at a concentration of 120 μ M significantly increases CSE transcription and expression³⁶. Additionally, hypoxia has been found to upregulate CSE expression to some extent²⁶. Therefore, it is crucial to investigate how exogenous H₂S regulates CSE expression in vascular endothelial cells during hypoxia. In this study, we examined the effects of applying 100 μ M exogenous H₂S on CSE expression in human umbilical vascular endothelial cells (HUVECs) under hypoxic conditions.

Since the higher concentration of exogenous H₂S is very harmful to cells³⁷, the down-regulation of exogenous H₂S at 160 μ M on CSE expression is perhaps unavoidable. Moreover, exogenous H₂S

(100 μ M) can alone inhibit HEK-293 cells proliferation³⁸. Exogenous H₂S could suppress CSE expression concentration-dependently while exogenous H₂S changed from 10 to 80 μ M. In the concentration range, exogenous H₂S might be feedback inhibition of CSE expression. The up-regulation of CSE expression may be derived from the protective effect of endogenous H₂S on cells at the higher concentration (from 120 to 160 μ M) of exogenous H₂S³⁶. Hydrogen sulfide donor sulforaphane (SFN) reshapes the CBS-H₂S signaling axis, activates mitochondrial autophagy, and suppresses inflammation, offering novel insights into multi-target therapeutic approaches for Parkinson's disease (PD) and underscoring the essential role of H₂S in neuroprotection³⁹.

There is no doubt that the CSE expression can respond specifically to exogenous H₂S within a certain concentration range. The up-regulation of CSE expression may be attributed, in part, to the detrimental effects of higher concentrations of exogenous H₂S on mammalian cells. CSE expression exhibits feedback regulation at lower concentrations of exogenous H₂S and can be upregulated at higher concentrations, while emphasizing that exceeding a certain level of exogenous H₂S concentration could pose harm to biological macromolecules.

3. Cystathionine γ -Lyase Expression Is Regulated by Exogenous Hydrogen Peroxide

Since H₂S is a reducing agent that readily reacts with hydrogen peroxide (H₂O₂), endogenous H₂S may scavenge reactive oxygen species (ROS). A state of moderately increased levels of intracellular ROS is referred to as oxidative stress such as H₂O₂. Although it can cause pathological damage, both H₂O₂ and endogenous H₂S also function in normal cell regulation^{40,41}. Endogenous H₂S acts as a reducing agent that readily reacts with H₂O₂ and scavenges ROS⁴². Administration of exogenous H₂S effectively protects myocytes and contractile activity by scavenging ROS⁴³ and accelerates the

removal of H₂O₂ generated by homocysteine in isolated mitochondria⁴⁴. The administration of H₂O₂ restored enzyme activity levels and accelerated the elimination of both H₂O₂ and superoxide anion generated by homocysteine in isolated mitochondria⁴⁴. Moreover, hydrogen peroxide protects neurons from oxidative stress by increasing the production of the antioxidant glutathione⁴², while also safeguarding cells against oxidative injury induced by H₂O₂^{45,46}.

Hydrogen sulfide synthesis is downregulated in the rostral ventrolateral medulla (RVLM)/Botzinger complex during hypoxia, and H₂S synthesis is diminished in the RVLM, facilitating hypothermia during hypoxia^{47,48}. H₂S protects cells against H₂O₂-induced oxidative injury^{45,46} and exerts its cytoprotective effect by increasing GSH production and scavenging ROS⁴⁹. Hydrogen sulfide has been identified as an excitatory mediator of hypoxic sensing in carotid bodies³¹. Hydrogen sulfide-induced gastroprotection against I/R-injury is due to increased gastric microcirculation and antioxidative properties²⁹. Hydrogen sulfide increases the generation of ROS in the carotid body enhances the chemosensory reflex, triggers hypertension⁵⁰, is an extremely reactive molecule, and may efficiently react with other compounds, especially reactive oxygen and nitrogen species⁵¹. Exogenous H₂S donor attenuated hypoxia-induced apoptosis in primary rat nucleus pulposus (NP) cells (Sun, Qi et al. 2018). Incubation with Na₂S³⁰. Incubation with NaHS increased the expression of miR-21 and attenuated the reduced cell viability and increased apoptosis caused by ischemia-reperfusion (I/R) in buffalo rat liver (BRL) cells³². Oxidative stress injury in I/R injury rats with CSE knockout was aggravated, while the increased expression of H₂S and CSE in the aortic tissues alleviated the oxidative stress injury⁵². Unexpected relationship between oxidative stress homeostasis and H₂S overproduction in Down Syndrome (DS), extending beyond human chromosome 21 (Hsa21) trisomy⁵³.

The effect of exogenously applied H₂O₂ on CSE expression in several mammalian cell lines demonstrates that exogenously applied H₂O₂ regulates CSE genes at promoter mRNA and protein levels. In particular, it is noteworthy that the treatment of a medium concentration (5 μM) of H₂O₂ at a longer time (1.5 h) upregulated CSE expression in the mammalian cells at levels of the promoter, mRNA, and protein⁵⁴. It can be speculated that the up-regulation of CSE expression induced by exogenous H₂O₂ may contribute to the increase in production and concentration of endogenous H₂S, which partially scavenges ROS like H₂O₂ in mammalian cell. Collectively, exogenously applied H₂O₂ significantly affects CSE mRNA and protein expression and affects CSE promoter activity in mammalian cells. Exogenous H₂O₂ can up-regulate the expression of the CSE gene in mammalian cells, which would open the possibility of indirect scavenging effects of the CSE gene on ROS in mammalian cells.

4. Regulation of Cystathionine γ-lyase Expression by Hypoxia

An adequate supply of O₂ is essential for the survival of the mammalian cells, and hypoxia profoundly impacts physiological systems⁵⁵. The H₂S-induced animation-state can protect mice from lethal hypoxia^{55,56}, and the anti-ischemic protection can be achieved by applying exogenous H₂S and up-regulating the endogenous production of H₂S⁵⁷. The regulatory effects of H₂S have been shown to modulate neuronal activity, relax smooth muscle, regulate insulin release, induce angiogenesis, suppress inflammation and protect cells⁵⁸. Reduced H₂S production mediated hypo-adrenocortical responsiveness and NLRP3 inflammasome activation during hypoxia⁵⁹.

The cystathionine γ-lyase/hydrogen sulfide pathway is indirectly linked to hypoxia, and H₂S protects mammalian cells against hypoxia-induced injury. Hydrogen sulfide is the physiologic gas transmitter of the carotid body and enhances its

sensory response to hypoxia⁶⁰, and is also an essential mediator of the hypoxic response in a variety of O₂-sensitive tissues^{61,62}. Sodium hydrosulfide administration reduced myocardial infarct size and prevented cardiomyocyte apoptosis⁶³, and S-propargyl-cysteine has cardioprotective properties in myocardial infarction rats and preserved cell viability when cultured cells were exposed to hypoxia⁶⁴. Experiments using cultured cardiomyocytes suggested hypoxia causes apoptosis, and apoptosis may play essential roles in ischemic heart diseases⁶⁵, and increased tissue content of H₂S protects the heart from ischemia/reperfusion damage⁵⁷. Hydrogen sulfide protects human skin keratinocytes against CoCl₂-induced injuries⁶⁶. Cystathionine γ-lyase transcription undergoes a period of repression and recovery, while CSE mRNA and protein levels increase during hypoxia². Hydrogen sulfide has the potential to restore aging-induced loss of cardioprotective effects of RIPC by up-regulating HIF-1α/Nrf2 signaling⁶⁷.

Cystathionine γ-lyase can respond to hypoxia through transcriptional and post-transcriptional regulation, and hypoxia can up-regulate CSE expression to some extent. Thus, the up-regulation of CSE expression during hypoxia may help increase the production and concentration of H₂S in mammalian cells, indirectly protecting cells from hypoxia. The author would like to propose a hypothesis that the up-regulation of CSE in mammalian cells may partially protect cells from lethal hypoxia, probably by increasing the H₂S content, buffering oxygen consumption within mammalian cells. Recently, The administration of exogenous H₂S further diminished CSE expression at various time points in HUVECs under hypoxic conditions. Technical terms were defined based on their initial applications. These findings strongly suggest that vascular endothelial cells respond to fluctuations in blood H₂S levels during oxygen-deficient periods⁶⁸.

5. Nuclear Factor κ B Transcription Factor Binding Site on Cystathionine γ -lyase Promoter Is Critical for LPS-Induced CSE Expression

The generation of CSE mRNA must occur along with H₂S in the rat aorta^{69,70} provided with exogenous L-cysteine. The CSE/H₂S signaling pathway is involved in the inflammation induced by endotoxins such as lipopolysaccharides (LPS). The inhibitory effects of LPS on endothelium-dependent relaxation results in pulmonary hypertension might also be mediated by H₂S⁷¹. Pulmonary surfactant (PS) decrease is the essential physiopathologic process of acute lung injury (ALI) induced by LPS. Exogenously applied H₂S can attenuate the process of ALI that likely because H₂S can adjust the composing and secretion of PS⁷². Hydrogen sulfide may represent a novel endogenous mechanism of cytoprotection in the inflamed joint, suggesting a potential opportunity for therapeutic intervention⁷³. Sodium hydrosulfide dose-dependently inhibited LPS-induced chemokine receptor C-X3-C motif chemokine receptor 1 expression in macrophages⁷⁴. Hydrogen sulfide breathing prevents inflammation and improves survival after the LPS challenge by altering sulfide metabolism in mice⁷⁵. Ralph A. Weinberg 264.7 murine macrophages treated with LPS mediated early apoptosis through TNF- α and the late apoptotic events through the production of H₂S⁷⁶. Endogenous H₂S deficiency contributed to sepsis-induced myocardial dysfunction (SIMD) and exogenous H₂S ameliorated sepsis-induced myocardial dysfunction by suppressing inflammation and endoplasmic reticulum stress (ERS) via inhibition of the TLR4 pathway⁷⁷. Deodorized garlic (DG) treatment decreases heart damage caused by LPS through the cross-talk between the H₂S and NO systems⁷⁸. Hydrogen sulfide inhibits the proliferation of A1 astrocytes induced by LPS-based neuroinflammation following cerebral I/R and promotes the transformation of astrocytes into A2 subtype, which may be related to up-regulation

of the large-conductance calcium- and voltage-activated K⁺ channel (BK_{Ca})⁷⁹. Protective effects of Alpha-lipoic acid (ALA) are dependent on the reduction in CSE expression in LPS-stimulated RAW 264.7 macrophages⁸⁰.

Nuclear Factor κ B is a heterodimer involving a variety of signaling pathways⁸¹. The transcription factor NF- κ B regulates inflammatory responses by inducing the expression of various genes⁸². The NF- κ B pathway can be rapidly activated by a large spectrum of chemically diverse agents and stress conditions, including bacterial LPS, microbial and viral pathogens, cytokines and growth factors⁸². Hydrogen sulfide can inhibit NF- κ B activation in LPS-stimulated macrophages⁸³. An increase in plasma H₂S concentration alongside augmented liver H₂S biosynthesis from exogenous cysteine is also apparent in animals four h after LPS injection^{84,85}. Serine (Ser) 276 phosphorylation of p65 is increased by LPS-mediated PKA (Protein kinase A) activation in Raw 264.7 murine macrophages⁸⁶. Secreted protein acidic and rich in cysteine (SPARC) afford beneficial actions in inhibitions I κ B- α degradation and phospho-transcription factors of the nuclear factor κ B p65 activation induced by LPS⁸⁷. Secreted protein acidic and rich in cysteine (SPARC) produces anti-inflammatory effects in LPS-stimulated H9c2 cells partly through the CSE/H₂S pathway by impairing I κ B- α /NF- κ B signaling⁸⁸. 4-trifluoromethylquinoline derivative (TKL002) exerts potent antiglioblastoma activity via modulation of the CTH/H₂S/NF κ B/epithelial-mesenchymal transition (EMT) signaling axis, highlighting its potential as a quinoline-based therapeutic candidate to overcome intrinsic glioblastoma (GBM) resistance and invasiveness⁸⁹.

Cystathionine γ -lyase / hydrogen sulfide signaling pathway is involved in the inflammation induced by endotoxins such as LPS^{73,83,87,88,90,91}. The transcriptional and post-transcriptional regulation of the CSE gene was involved in the NF κ B transcription factor binding site on the promoter in the mammalian cells under the treatment of LPS.

Some studies have indicated that H₂S plays an essential role in inflammation, and LPS stimulates the expression of the CSE gene and H₂S production rate⁹². Hydrogen sulfide regulates LPS-induced inflammation and apoptosis by activating the PI3K/Akt/NF- κ B signaling pathway⁹³. The source of H₂S, either endogenous (via CSE) or exogenous (via GYY4137), supports or inhibits the LPS-induced NF- κ B activity and Glut1 expression, respectively⁹⁴.

Exogenous H₂S inhibits wound-induced macrophage activation via the NF- κ B pathway, and appropriate H₂S supplementation may help control inflammation⁹⁵. SPRC exerts an anti-inflammatory effect in LPS-stimulated rat cardiomyocyte H9c2 cells partly through the CSE/H₂S pathway impairing I κ B- α /NF- κ B signaling⁸⁸. Lipopolysaccharide increases the biosynthesis of CSE and H₂S in macrophages, mainly in a TLR-4- and NF- κ B-dependent manner⁹⁶. Sodium hydrosulfide (an H₂S donor) suppresses the degradation of I κ B- α and the activity of NF- κ B⁹⁷. Endogenous H₂S inactivates IKK beta to inhibit the NF- κ B pathway and control pulmonary artery endothelial cell (PAEC) inflammation⁹⁸. tumor necrosis factor- α affects CSE gene expression, such that vascular endothelial cells respond to TNF- α in the blood by regulating CSE expression. The regulatory mechanisms associated with the effects of TNF- α on the transcriptional regulation of the CSE gene in HUVECs and the NF- κ B pathway warrant further investigation⁹⁹.

Exogenous H₂S attenuates angiotensin II-induced inflammation and cytotoxicity by inhibiting the ET1/NF- κ B signaling pathway in HUVECs¹⁰⁰. The effects of H₂S on the modulation of smooth muscle relaxation, inflammation suppression, and cell protection have been extensively studied¹⁰¹, and the CSE/H₂S signaling pathway is involved in the inflammation induced by endotoxins, such as LPS. The NF- κ B-binding site in the CSE promoter is critical for LPS-induced CSE expression in mammalian cells⁹². Moreover, the CSE/H₂S system protects against LPS-induced inflammation and

cell hyperpermeability by blocking NF- κ B transactivation¹⁰². Lipopolysaccharide-induced inflammation in murine macrophages is associated with H₂S production, and CSE/H₂S attenuates LPS-induced sepsis by inhibiting oxidative stress^{76,103}. Exogenous H₂S ameliorates intestinal injury by downregulating inflammation and activation of autophagy, suggesting the potential of NaHS as a therapeutic agent for ulcerative colitis (UC)¹⁰⁴. Methionine restriction (MR) attenuated LPS-induced lung injury through CSE and H₂S modulation¹⁰⁵.

Lipopolysaccharide could significantly increase the expression levels of mRNA and protein of the CSE gene after J774.1A cells and RAW264.7 cells under the treatment of LPS for 6 h⁹². Since LPS affected the post-transcriptional regulation of the CSE gene, there may be the NF- κ B transcription factor binding site on the promoter of the CSE gene in the mammalian cells. The DNA sequence GGACATTCC on the promoter of the CSE gene has been heavily associated with transcriptional regulation of the CSE gene in mammalian cells undergoing LPS therapy. The NF- κ B transcription factor on the CSE promoter is critical for LPS-induced CSE expression in mammalian cells.

6. Conclusion

Cystathionine γ -lyase is the dominant enzyme responsible for the production of endogenous H₂S in the cardiovascular system, where hypoxia is one of the pathogenic conditions regularly encountered. Hypoxia-regulated CSE expression in mammalian cells is complex, with CSE transcription undergoing periods of repression and restoration, while total mRNA and protein levels increase during hypoxia. Based on the above analysis, we proposed a hypothesis about the mechanism of CSE protection on the myocardial cells (Figure 1), and this assumption explains the mechanism as follows. Under hypoxic conditions, apoptosis is induced in some cardiomyocytes, a process that releases H₂O₂ into vascular endothelial cells in the minor arteries of the heart.

Regulation of CSE in the Cardiovascular System.

Proper concentrations of H_2O_2 promote the resumption of CSE expression in vascular endothelial cells via NF- κ B signaling pathway. The up-regulation of CSE expression in vascular endothelial cells produces additional hydrogen sulfide H_2S , which accelerates vasodilation in SMCs.

The CSE gene is primarily expressed in the cardiovascular system. Its close relationship to the occurrence of cardiovascular disease suggests that

the study of the regulatory mechanisms of CSE gene expression has significant theoretical and clinical value in the study of cardiovascular disease. Through the analysis of the CSE gene and its promoter in different populations, if it can be confirmed at the molecular level that some cardiovascular diseases, such as myocardial infarction, are caused by abnormalities of the CSE gene and its promoter, it can provide new ideas for effective screening and active intervention treatment of some cardiovascular diseases.

Figure 1

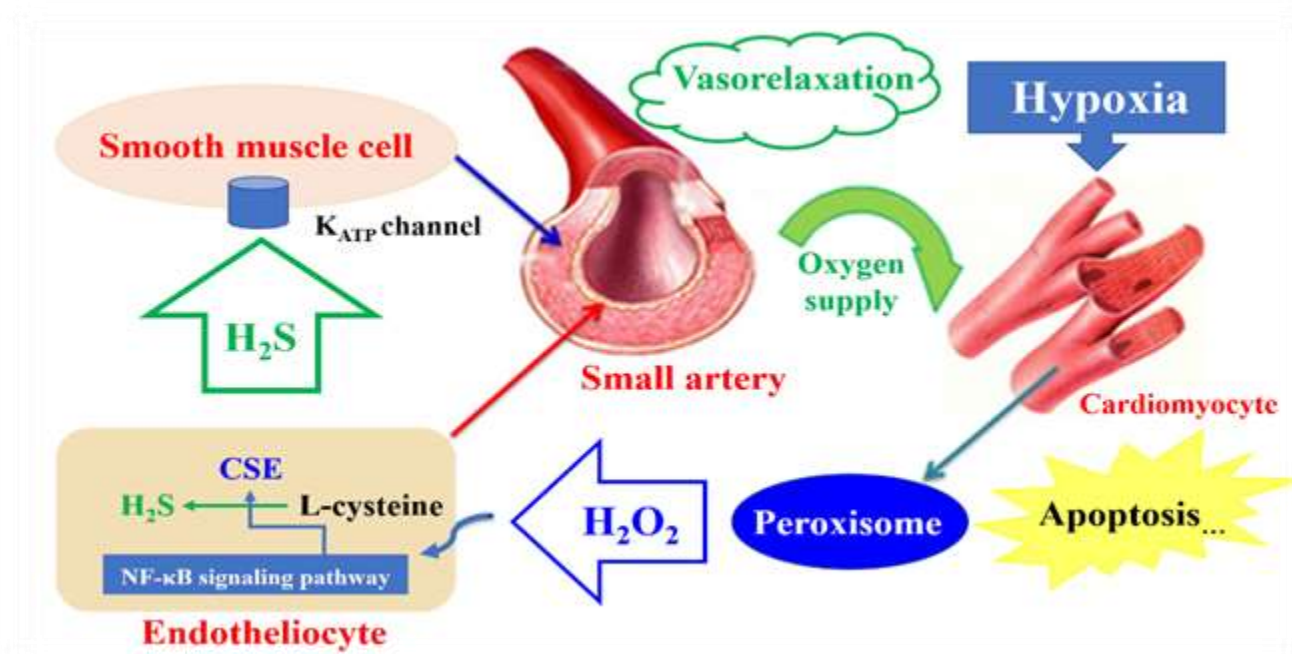


Figure 1 Legends

A hypothesis on the mechanism of cystathionine γ -Lyase protection of myocardial cells during hypoxia. Under hypoxia, cardiomyocyte apoptosis releases H_2O_2 into vascular endothelial cells, where appropriate levels restore CSE expression via NF- κ B signaling. The resulting H_2S promotes vasodilation of smooth muscle cells, improving circulation—this increases cardiac oxygen supply, reduces infarct size, and attenuates apoptosis

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

Maoxian Wang made the contributions to this study, writing, reviewing and approving the final version of the manuscript alone.

Ethics approval and consent to participate

Not applicable.

Conflict of interest statement

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Additional files

Not applicable

References

1. Wang, R. The gasotransmitter role of hydrogen sulfide. *Antioxid Redox Signal* 5, 493-501 (2003).
2. Kimura, H. Hydrogen sulfide: its production and functions. *Exp Physiol* 96, 833-835 (2011).
3. Enokido, Y., *et al.* Cystathionine beta-synthase, a key enzyme for homocysteine metabolism, is preferentially expressed in the radial glia/astrocyte lineage of developing mouse CNS. *FASEB J* 19, 1854-1856 (2005).
4. Shibuya, N., *et al.* 3-Mercaptopyruvate sulfurtransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 11, 703-714 (2009).
5. Yang, G., *et al.* Cystathionine gamma-lyase deficiency and overproliferation of smooth muscle cells. *Cardiovascular research* 86, 487-495 (2010).
6. Xu, M., *et al.* Hydrogen sulfide: Recent progress and perspectives for the treatment of dermatological diseases. *J Adv Res* 27, 11-17 (2021).
7. Gall, T., *et al.* Overview on hydrogen sulfide-mediated suppression of vascular calcification and hemoglobin/heme-mediated vascular damage in atherosclerosis. *Redox Biol* 57, 102504 (2022).
8. Yan, S.K., *et al.* Effects of hydrogen sulfide on homocysteine-induced oxidative stress in vascular smooth muscle cells. *Biochem Biophys Res Commun* 351, 485-491 (2006).
9. Yang, G., Wu, L. & Wang, R. Pro-apoptotic effect of endogenous H₂S on human aorta smooth muscle cells. *FASEB J* 20, 553-555 (2006).
10. Elrod, J.W., *et al.* Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. *Proc Natl Acad Sci U S A* 104, 15560-15565 (2007).
11. Yang, G., *et al.* H₂S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gamma-lyase. *Science* 322, 587-590 (2008).
12. Zhang, J., Xie, Y., Xu, Y., Pan, Y. & Shao, C. Hydrogen sulfide contributes to hypoxia-induced radioresistance on hepatoma cells. *J Radiat Res* 52, 622-628 (2011).
13. Alshahwan, H., *et al.* Hydrogen sulfide donor GYY4137 attenuates vascular complications in mesenteric bed of streptozotocin-induced diabetic rats. *Eur J Pharmacol* 933, 175265 (2022).
14. Beltowski, J. & Kowalczyk-Boltuc, J. Hydrogen sulfide in the experimental models of arterial hypertension. *Biochem Pharmacol* 208, 115381 (2023).
15. Geng, B., *et al.* H₂S generated by heart in rat and its effects on cardiac function. *Biochemical and biophysical research communications* 313, 362-368 (2004).
16. Osmond, J.M. & Kanagy, N.L. Modulation of hydrogen sulfide by vascular hypoxia. *Hypoxia (Auckl)* 2, 117-126 (2014).
17. Pushpakumar, S., Kundu, S. & Sen, U. Endothelial dysfunction: the link between homocysteine and hydrogen sulfide. *Curr Med Chem* 21, 3662-3672 (2014).
18. Kimura, H. Signaling molecules: hydrogen sulfide and polysulfide. *Antioxid Redox Signal* 22, 362-376 (2015).
19. Sabino, J.P., Traslavina, G.A. & Branco, L.G. Role of central hydrogen sulfide on ventilatory and cardiovascular responses to hypoxia in spontaneous hypertensive rats. *Respir Physiol Neurobiol* 231, 21-27 (2016).
20. Leucker, T.M., *et al.* Cystathionine gamma-lyase protects vascular endothelium: a role for inhibition of histone deacetylase 6. *Am J Physiol Heart Circ Physiol* 312, H711-H720 (2017).
21. Webb, G.D., *et al.* Contractile and vasorelaxant effects of hydrogen sulfide and its biosynthesis in the human internal mammary artery. *J Pharmacol Exp Ther* 324, 876-882 (2008).

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22. Ariyaratnam, P., Loubani, M. & Morice, A.H. Hydrogen sulphide vasodilates human pulmonary arteries: a possible role in pulmonary hypertension? *Microvasc Res* 90, 135-137 (2013).
23. Materazzi, S., *et al.* Vasodilator activity of hydrogen sulfide (H₂S) in human mesenteric arteries. *Microvasc Res* 109, 38-44 (2017).
24. Cacanyiova, S., *et al.* Nitroso-sulfide coupled signaling triggers specific vasoactive effects in the intrarenal arteries of patients with arterial hypertension. *J Physiol Pharmacol* 68, 527-538 (2017).
25. Cindrova-Davies, T., *et al.* Reduced cystathionine gamma-lyase and increased miR-21 expression are associated with increased vascular resistance in growth-restricted pregnancies: hydrogen sulfide as a placental vasodilator. *Am J Pathol* 182, 1448-1458 (2013).
26. Wang, M., Guo, Z. & Wang, S. Regulation of cystathionine gamma-lyase in mammalian cells by hypoxia. *Biochem Genet* 52, 29-37 (2014).
27. Tran, B.H., *et al.* Cardioprotective effects and pharmacokinetic properties of a controlled release formulation of a novel hydrogen sulfide donor in rats with acute myocardial infarction. *Biosci Rep* 35(2015).
28. Mistry, R.K., *et al.* Transcriptional Regulation of Cystathionine-gamma-Lyase in Endothelial Cells by NADPH Oxidase 4-Dependent Signaling. *J Biol Chem* 291, 1774-1788 (2016).
29. Magierowski, M., *et al.* Exogenous and Endogenous Hydrogen Sulfide Protects Gastric Mucosa against the Formation and Time-Dependent Development of Ischemia/Reperfusion-Induced Acute Lesions Progressing into Deeper Ulcerations. *Molecules* 22(2017).
30. Sun, H., Qi, L., Wang, S., Li, X. & Li, C. Hydrogen sulfide is expressed in the human and the rat cultured nucleus pulposus cells and suppresses apoptosis induced by hypoxia. *PLoS One* 13, e0192556 (2018).
31. Wu, B., *et al.* Interaction of Hydrogen Sulfide with Oxygen Sensing under Hypoxia. *Oxid Med Cell Longev* 2015, 758678 (2015).
32. Lu, M., *et al.* MicroRNA-21-Regulated Activation of the Akt Pathway Participates in the Protective Effects of H₂S against Liver Ischemia-Reperfusion Injury. *Biol Pharm Bull* 41, 229-238 (2018).
33. Ye, M., *et al.* Exogenous hydrogen sulfide donor NaHS alleviates nickel-induced epithelial-mesenchymal transition and the migration of A549 cells by regulating TGF-beta1/Smad2/Smad3 signaling. *Ecotoxicol Environ Saf* 195, 110464 (2020).
34. Pavlovskiy, Y., Yashchenko, A. & Zayachkivska, O. H₂S Donors Reverse Age-Related Gastric Malfunction Impaired Due to Fructose-Induced Injury via CBS, CSE, and TST Expression. *Front Pharmacol* 11, 1134 (2020).
35. Xu, M., Wu, Y.M., Li, Q., Wang, F.W. & He, R.R. Electrophysiological effects of hydrogen sulfide on guinea pig papillary muscles in vitro. *Sheng Li Xue Bao* 59, 215-220 (2007).
36. Wang, M., Guo, Z. & Wang, S. The effect of certain conditions in the regulation of cystathionine gamma-lyase by exogenous hydrogen sulfide in mammalian cells. *Biochem Genet* 51, 503-513 (2013).
37. Reiffenstein, R.J., Hulbert, W.C. & Roth, S.H. Toxicology of hydrogen sulfide. *Annual review of pharmacology and toxicology* 32, 109-134 (1992).
38. Yang, G., Cao, K., Wu, L. & Wang, R. Cystathionine gamma-lyase overexpression inhibits cell proliferation via a H₂S-dependent modulation of ERK1/2 phosphorylation and p21Cip/WAK-1. *J Biol Chem* 279, 49199-49205 (2004).
39. Xie, W., *et al.* The H₂S donor sulforaphane inhibits NLRP(3) inflammasome activation by inducing mitochondrial autophagy and mitigating CBS-H₂S axis damage in in-vitro

- and in-vivo models of Parkinson's disease. *Bioorg Chem* 174, 109708 (2026).
40. Denu, J.M. & Tanner, K.G. Specific and reversible inactivation of protein tyrosine phosphatases by hydrogen peroxide: evidence for a sulfenic acid intermediate and implications for redox regulation. *Biochemistry* 37, 5633-5642 (1998).
 41. Schreck, R., Rieber, P. & Baeuerle, P.A. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappa B transcription factor and HIV-1. *EMBO J* 10, 2247-2258 (1991).
 42. Kimura, Y. & Kimura, H. Hydrogen sulfide protects neurons from oxidative stress. *FASEB J* 18, 1165-1167 (2004).
 43. Geng, B., *et al.* Endogenous hydrogen sulfide regulation of myocardial injury induced by isoproterenol. *Biochemical and biophysical research communications* 318, 756-763 (2004).
 44. Chang, L., *et al.* Hydrogen sulfide inhibits myocardial injury induced by homocysteine in rats. *Amino acids* 34, 573-585 (2008).
 45. Lu, M., Hu, L.F., Hu, G. & Bian, J.S. Hydrogen sulfide protects astrocytes against H₂O₂-induced neural injury via enhancing glutamate uptake. *Free Radic Biol Med* 45, 1705-1713 (2008).
 46. Xu, Z.S., *et al.* Hydrogen sulfide protects MC3T3-E1 osteoblastic cells against H₂O₂-induced oxidative damage-implications for the treatment of osteoporosis. *Free Radic Biol Med* 50, 1314-1323 (2011).
 47. Donatti, A.F., Soriano, R.N., Sabino, J.P. & Branco, L.G. Endogenous hydrogen sulfide in the rostral ventrolateral medulla/Botzinger complex downregulates ventilatory responses to hypoxia. *Respir Physiol Neurobiol* 200, 97-104 (2014).
 48. Donatti, A.F., Soriano, R.N., Sabino, J.P. & Branco, L.G. Involvement of endogenous hydrogen sulfide (H₂S) in the rostral ventrolateral medulla (RVLM) in hypoxia-induced hypothermia. *Brain Res Bull* 108, 94-99 (2014).
 49. Kimura, H. Hydrogen sulfide and polysulfides as signaling molecules. *Proc Jpn Acad Ser B Phys Biol Sci* 91, 131-159 (2015).
 50. Yuan, G., *et al.* H₂S production by reactive oxygen species in the carotid body triggers hypertension in a rodent model of sleep apnea. *Sci Signal* 9, ra80 (2016).
 51. Wen, Y.D., Wang, H. & Zhu, Y.Z. The Drug Developments of Hydrogen Sulfide on Cardiovascular Disease. *Oxid Med Cell Longev* 2018, 4010395 (2018).
 52. Luo, Y., *et al.* Activation of the CaR-CSE/H₂S pathway confers cardioprotection against ischemia-reperfusion injury. *Exp Cell Res* 398, 112389 (2021).
 53. Mouli, K., *et al.* SOD1 at the Crossroads: Co-Overexpression of Canonical Antioxidant Response and Noncanonical Hydrogen Sulfide Generation Pathways in Down Syndrome, With Immune Cell Implications. *Res Sq* (2026).
 54. Wang, M., Guo, Z. & Wang, S. Cystathionine gamma-lyase expression is regulated by exogenous hydrogen peroxide in the mammalian cells. *Gene Expr* 15, 235-241 (2012).
 55. Blackstone, E., Morrison, M. & Roth, M.B. H₂S induces a suspended animation-like state in mice. *Science* 308, 518 (2005).
 56. Blackstone, E. & Roth, M.B. Suspended animation-like state protects mice from lethal hypoxia. *Shock* 27, 370-372 (2007).
 57. Wang, R. Hydrogen sulfide: the third gasotransmitter in biology and medicine. *Antioxid Redox Signal* 12, 1061-1064 (2010).
 58. Kimura, H. Hydrogen sulfide: its production, release and functions. *Amino acids* 41, 113-121 (2011).
 59. Zhang, N., *et al.* Reduced hydrogen sulfide production contributes to adrenal insufficiency induced by hypoxia via modulation of NLRP3

- inflammasome activation. *Redox Rep* 28, 2163354 (2023).
60. Peng, Y.J., *et al.* H₂S mediates O₂ sensing in the carotid body. *Proc Natl Acad Sci U S A* 107, 10719-10724 (2010).
61. Dombkowski, R.A., *et al.* Hydrogen sulfide (H₂S) and hypoxia inhibit salmonid gastrointestinal motility: evidence for H₂S as an oxygen sensor. *J Exp Biol* 214, 4030-4040 (2011).
62. Dombkowski, R.A., Doellman, M.M., Head, S.K. & Olson, K.R. Hydrogen sulfide mediates hypoxia-induced relaxation of trout urinary bladder smooth muscle. *J Exp Biol* 209, 3234-3240 (2006).
63. Yao, L.L., *et al.* Hydrogen sulfide protects cardiomyocytes from hypoxia/reoxygenation-induced apoptosis by preventing GSK-3 β -dependent opening of mPTP. *Am J Physiol Heart Circ Physiol* 298, H1310-1319 (2010).
64. Wang, Q., Liu, H.R., Mu, Q., Rose, P. & Zhu, Y.Z. S-propargyl-cysteine protects both adult rat hearts and neonatal cardiomyocytes from ischemia/hypoxia injury: the contribution of the hydrogen sulfide-mediated pathway. *J Cardiovasc Pharmacol* 54, 139-146 (2009).
65. Takemura, G., Ohno, M. & Fujiwara, H. [Ischemic heart disease and apoptosis]. *Rinsho byori. The Japanese journal of clinical pathology* 45, 606-613 (1997).
66. Yang, C., *et al.* Hydrogen sulfide protects against chemical hypoxia-induced cytotoxicity and inflammation in HaCaT cells through inhibition of ROS/NF- κ B/COX-2 pathway. *PLoS One* 6, e21971 (2011).
67. Wang, H., Shi, X., Cheng, L., Han, J. & Mu, J. Hydrogen sulfide restores cardioprotective effects of remote ischemic preconditioning in aged rats via HIF-1 α /Nrf2 signaling pathway. *Korean J Physiol Pharmacol* 25, 239-249 (2021).
68. Wang, M. Exogenous H₂S Regulates CSE Expression in HUVECs under Hypoxic Conditions. *Journal of Clinical, Medical, and Diagnostic Research* 4, 1-8 (2026).
69. Zhao, W., Zhang, J., Lu, Y. & Wang, R. The vasorelaxant effect of H₂S as a novel endogenous gaseous K(ATP) channel opener. *EMBO J* 20, 6008-6016 (2001).
70. Hosoki, R., Matsuki, N. & Kimura, H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. *Biochemical and biophysical research communications* 237, 527-531 (1997).
71. Huang, X.L., Zhou, X.H., Wei, P., Xian, X.H. & Ling, Y.L. [The role of hydrogen sulfide in acute lung injury during endotoxic shock and its relationship with nitric oxide and carbon monoxide]. *Zhonghua Yi Xue Za Zhi* 88, 2240-2245 (2008).
72. Wang, P., *et al.* [Effects of hydrogen sulfide on pulmonary surfactant in rats with acute lung injury induced by lipopolysaccharide]. *Zhongguo Ying Yong Sheng Li Xue Za Zhi* 27, 485-489 (2011).
73. Fox, B., *et al.* Inducible hydrogen sulfide synthesis in chondrocytes and mesenchymal progenitor cells: is H₂S a novel cytoprotective mediator in the inflamed joint? *J Cell Mol Med* 16, 896-910 (2012).
74. Zhang, H., *et al.* Hydrogen sulfide inhibits the development of atherosclerosis with suppressing CX3CR1 and CX3CL1 expression. *PLoS One* 7, e41147 (2012).
75. Tokuda, K., *et al.* Inhaled hydrogen sulfide prevents endotoxin-induced systemic inflammation and improves survival by altering sulfide metabolism in mice. *Antioxid Redox Signal* 17, 11-21 (2012).
76. George, L., Ramasamy, T., Sirajudeen, K. & Manickam, V. LPS-induced Apoptosis is Partially Mediated by Hydrogen Sulphide in RAW 264.7 Murine Macrophages. *Immunol Invest* 48, 451-465 (2019).
77. Chen, Y.H., *et al.* Hydrogen Sulfide Attenuated Sepsis-Induced Myocardial Dysfunction

- Through TLR4 Pathway and Endoplasmic Reticulum Stress. *Front Physiol* 12, 653601 (2021).
78. Perez-Torres, I., *et al.* Deodorized Garlic Decreases Oxidative Stress Caused by Lipopolysaccharide in Rat Heart through Hydrogen Sulfide: Preliminary Findings. *Int J Mol Sci* 23(2022).
79. Li, X., Yin, X., Pang, J., Chen, Z. & Wen, J. Hydrogen sulfide inhibits lipopolysaccharide-based neuroinflammation-induced astrocyte polarization after cerebral ischemia/reperfusion injury. *Eur J Pharmacol* 949, 175743 (2023).
80. Shahid, A., Chambers, S., Scott-Thomas, A., Zawari, M. & Bhatia, M. Anti-Inflammatory Effects of Alpha-Lipoic Acid Modulate Cystathionine-gamma-Lyase Expression in RAW 264.7 Macrophages. *Int J Mol Sci* 27(2026).
81. Pahl, H.L. Activators and target genes of Rel/NF-kappaB transcription factors. *Oncogene* 18, 6853-6866 (1999).
82. Baeuerle, P.A. & Baltimore, D. NF-kappa B: ten years after. *Cell* 87, 13-20 (1996).
83. Oh, G.S., *et al.* Hydrogen sulfide inhibits nitric oxide production and nuclear factor-kappaB via heme oxygenase-1 expression in RAW264.7 macrophages stimulated with lipopolysaccharide. *Free Radic Biol Med* 41, 106-119 (2006).
84. Zhu, X.Y., Liu, S.J., Liu, Y.J., Wang, S. & Ni, X. Glucocorticoids suppress cystathionine gamma-lyase expression and H₂S production in lipopolysaccharide-treated macrophages. *Cell Mol Life Sci* 67, 1119-1132 (2010).
85. Li, L., Whiteman, M. & Moore, P.K. Dexamethasone inhibits lipopolysaccharide-induced hydrogen sulphide biosynthesis in intact cells and in an animal model of endotoxic shock. *Journal of cellular and molecular medicine* 13, 2684-2692 (2009).
86. Moon, E.Y., Lee, J.H., Lee, J.W., Song, J.H. & Pyo, S. ROS/Epac1-mediated Rap1/NF-kappaB activation is required for the expression of BAFF in Raw264.7 murine macrophages. *Cell Signal* 23, 1479-1488 (2011).
87. Gong, Q.H., *et al.* S-propargyl-cysteine, a novel hydrogen sulfide-modulated agent, attenuates lipopolysaccharide-induced spatial learning and memory impairment: involvement of TNF signaling and NF-kappaB pathway in rats. *Brain Behav Immun* 25, 110-119 (2011).
88. Pan, L.L., Liu, X.H., Gong, Q.H. & Zhu, Y.Z. S-Propargyl-cysteine (SPRC) attenuated lipopolysaccharide-induced inflammatory response in H9c2 cells involved in a hydrogen sulfide-dependent mechanism. *Amino Acids* 41, 205-215 (2011).
89. Luo, Z., *et al.* Hijacking the Hydrogen Sulfide Axis: A Novel 4-Trifluoromethylquinoline Derivative Suppresses Glioblastoma via Cystathionine gamma-Lyase Suppression. *J Med Chem* 69, 3457-3476 (2026).
90. Huang, X.L., *et al.* Role of endogenous hydrogen sulfide in pulmonary hypertension induced by lipopolysaccharide. *Sheng li xue bao : [Acta physiologica Sinica]* 60, 211-215 (2008).
91. Zhang, J., Xie, Y., Xu, Y. & Shao, C. Suppression of endogenous hydrogen sulfide contributes to the radiation-induced bystander effects on hypoxic HepG2 cells. *Radiat Res* 178, 395-402 (2012).
92. Wang, M., Guo, Z. & Wang, S. The binding site for the transcription factor, NF-kappaB, on the cystathionine gamma-lyase promoter is critical for LPS-induced cystathionine gamma-lyase expression. *Int J Mol Med* 34, 639-645 (2014).
93. Wang, Y., *et al.* H₂S mediates apoptosis in response to inflammation through PI3K/Akt/NFkappaB signaling pathway. *Biotechnol Lett* 42, 375-387 (2020).
94. Cornwell, A., Fedotova, S., Cowan, S. & Badiei, A. Cystathionine gamma-lyase and hydrogen sulfide modulates glucose transporter Glut1 expression via NF-kappaB and PI3k/Akt in

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- macrophages during inflammation. *PLoS One* 17, e0278910 (2022).
95. Zhuang, R., *et al.* Exogenous hydrogen sulfide inhibits oral mucosal wound-induced macrophage activation via the NF-kappaB pathway. *Oral Dis* 24, 793-801 (2018).
96. Zheng, Y., *et al.* Lipopolysaccharide regulates biosynthesis of cystathionine gamma-lyase and hydrogen sulfide through Toll-like receptor-4/p38 and Toll-like receptor-4/NF-kappaB pathways in macrophages. *In Vitro Cell Dev Biol Anim* 49, 679-688 (2013).
97. Rao, C.Y., *et al.* H₂S mitigates severe acute pancreatitis through the PI3K/AKT-NF-kappaB pathway in vivo. *World J Gastroenterol* 21, 4555-4563 (2015).
98. Zhang, D., *et al.* Endogenous hydrogen sulfide sulfhydrates IKKbeta at cysteine 179 to control pulmonary artery endothelial cell inflammation. *Clin Sci (Lond)* 133, 2045-2059 (2019).
99. Wang, M. TNFalpha regulates the expression of the CSE gene in HUVEC. *Exp Ther Med* 22, 1233 (2021).
100. Hu, H.J., Jiang, Z.S., Zhou, S.H. & Liu, Q.M. Hydrogen sulfide suppresses angiotensin II-stimulated endothelin-1 generation and subsequent cytotoxicity-induced endoplasmic reticulum stress in endothelial cells via NF-kappaB. *Mol Med Rep* 14, 4729-4740 (2016).
101. Kimura, H. The physiological role of hydrogen sulfide and beyond. *Nitric Oxide* 41, 4-10 (2014).
102. Bourque, C., *et al.* H₂S protects lipopolysaccharide-induced inflammation by blocking NFkappaB transactivation in endothelial cells. *Toxicol Appl Pharmacol* 338, 20-29 (2018).
103. Wang, X.L., *et al.* Endogenous Hydrogen Sulfide Ameliorates NOX4 Induced Oxidative Stress in LPS-Stimulated Macrophages and Mice. *Cell Physiol Biochem* 47, 458-474 (2018).
104. Liu, Y., *et al.* Exogenous H₂S Protects Colon Cells in Ulcerative Colitis by Inhibiting NLRP3 and Activating Autophagy. *DNA Cell Biol* 40, 748-756 (2021).
105. Duan, J., *et al.* Methionine Restriction Prevents Lipopolysaccharide-Induced Acute Lung Injury via Modulating CSE/H₂S Pathway. *Nutrients* 14(2022).